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Impaired exercise training-induced muscle fiber hypertrophy and Akt/mTOR pathway activation in hypoxemic patients with COPD

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COPD; skeletal muscle; hypoxia; exercise training

CHRONIC OBSTRUCTIVE PULMONARY disease (COPD) is one of the main causes of morbidity and a leading cause of death worldwide (32). Muscle dysfunction is an important systemic consequence of COPD (17, 47). It is characterized by a shift from type I to type II muscle fibers, a loss of oxidative capacity, a reduced capillary density, and an atrophy of muscle fibers resulting in a severe loss of muscle mass (49). All these factors significantly contribute to reduce a patient’s exercise capacity and quality of life, ultimately leading to greater mortality (5, 38). Although interventional strategies such as nutritional support and exercise training (ExTr) improve the quality of life and survival of patients with COPD when associated with a weight gain (29, 39), these strategies do not succeed in counteracting muscle dysfunction and mass loss in patients.

Skeletal muscle mass is tightly regulated by the Akt/mammalian target of rapamycin (mTOR) pathway. Stimulation of the Akt/mTOR pathway increases protein translation in skeletal muscle (7, 36) and inhibits protein degradation via the inhibition of both ubiquitin-proteasome (7, 37, 40) and autophagy-lysosome pathways (26, 50). This notably involves the regulation of FoxO1 and FoxO3 transcriptional activity on the promoters of MuRF1, Atrogin-1/MAFbx, and autophagy-related genes (26, 37, 40, 50). The available literature is conflicting regarding the regulation of this pathway in skeletal muscle of patients with COPD. A downregulation of the Akt/mTOR pathway has been reported in skeletal muscle of patients with COPD compared with healthy subjects (45), whereas other studies reported no difference (30) or even an upregulation of the pathway (11). By contrast, a high-intensity interval training program allows the reactivation of the Akt/mTOR pathway in skeletal muscle of patients with COPD (45). Furthermore, strength training in these patients increases muscle expression of insulin-like growth factor-1 (IGF-1), an upstream activator of the signaling pathway (24). Therefore, rehabilitation strategies incorporating resistance exercise may be helpful in limiting the extent of skeletal muscle mass loss in patients with COPD by activating the Akt/mTOR pathway.

However, some recent studies suggest that the severity of hypoxemia could be associated with a resistance of skeletal muscle to the activation of the Akt/mTOR pathway. Indeed, muscle atrophy resulting from ambient hypoxia in rodents involves a downregulation of the Akt/mTOR pathway and an upregulation of the ubiquitin/proteasome pathway (9, 18). In line with these findings, we reported a downregulation of the Akt/mTOR pathway in hypoxemic patients with COPD compared with normoxic patients with COPD (18). Taken together, these data strongly suggest that the response of Akt/mTOR pathway to ExTr could be compromised in hypoxemic patients with COPD.

In the present study, we therefore tested the hypothesis that the response of skeletal muscle to ExTr would be altered in patients with COPD and severe hypoxemia compared with normoxic patients with COPD. We particularly focused our attention on the regulation of muscle fiber size and Akt/mTOR pathway. To further delineate the role of hypoxia, an in vitro analysis of the effects of hypoxia on the regulation of Akt/mTOR pathway was also performed on C2C12 myotubes.
METHODS

Subjects

We included 23 consecutive patients with COPD who entered an outpatient pulmonary rehabilitation center (CHU Saint Etienne). Written consent in accordance with the policy statement regarding the use of human subjects was obtained from all patients. This investigation was approved by the Rhône-Alpes Loire regional Consultant Committee on Human Protection for Medical Research and received agreement from the French Health Minister (DGS 2005/023). Criteria for inclusion in the study were a stable COPD disease (absence of exacerbation during the last 4 wk), the ability to perform maximal exercise testing, and no contraindication to muscle biopsy (e.g., chronic anticoagulant treatment). All patients were treated with inhaled long-acting sympathomimetics and inhaled corticosteroids. Fifteen patients were also treated with tiotropium. None used oral corticosteroid regularly at the time of inclusion. Patients were considered to be hypoxemic (long-term oxygen therapy ≥6 h/day for more than 3 mo, resting arterial PO2 <55 mmHg at the initiation of the treatment, n = 8) or normoxemic (resting arterial PO2 > 60 mmHg, n = 15).

Pulmonary Function and Morphometric Characteristics of Patients

Lung volumes and airflows were measured (Bodybox; Medisoft, Dinant Belgium) according to European Respiratory Society recommendations (48). Postbronchodilator values were reported. Body composition was assessed by bioelectrical impedance at 50 Hz (Nutrigard Data Input; Pöcking, Germany) and fat free mass was calculated and compared with normal values according to those outlined by Kyle et al. (23). Body mass index (BMI) and fat-free mass index (FFMI) were also calculated.

Evaluation of Exercise Capacity and Muscle Strength

Incremental cycling exercise. After a 3-min warmup, the patients performed an incremental exercise test on a bicycle ergometer (5 to 10 W every min) while breathing room air (Ergocard; Medisoft, Dinant, Belgium). Breath-by-breath analysis of inspired and expired gases was used to determine oxygen consumption (VO2), CO2 output (VCO2), and minute ventilation (Ve). Peak power output (Wpeak) corresponded to the highest workload that could be sustained for more than 20 s. Electrocardiographic and arterial oxygen saturation readings were monitored continuously. Arterialized blood samples from the ear were used for blood gas analysis and lactate measurement (ABL 800; Radiometer, Copenhagen, Denmark). A Borg scale was used to assess dyspnea and fatigue. Exercise capacity was determined before and after ExTr.

Maximal muscle force. Patients sat on a bench and performed isometric maximal voluntary contractions of the quadriceps muscle with a 90° knee flexion while breathing room air. Muscle force was recorded with a dynamometer attached to the bench (Globus, Codognè, Italy). Handgrip strength was tested using a hand dynamometer (Jamar, Anaheim, CA). For each test, the best of three reproducible contractions (±10%) was recorded. Muscle strength was assessed before and after ExTr.

Multidisciplinary Pulmonary Rehabilitation Program

Patients participated in a multidisciplinary rehabilitation program consisting of 24 sessions (three sessions/week) under the supervision of a physiotherapist. ExTr included endurance bicycle exercise (20 to 30 min) and treadmill exercise (10 to 15 min). Patients were free to adapt resting periods as necessary. Exercise intensity was initially set to a heart rate corresponding to the ventilatory threshold (VT) measured during the initial maximal cycling test (42). When VT was not discernible (one patient in the hypoxemic group and two patients in the normoxemic group), the exercise intensity was arbitrarily fixed at 60% of peak workload. Heart rate and oxygen saturation were monitored every 10 min during the session. Exercise intensity was adjusted every week to maintain heart rate to the target value: the workload was increased by 5 W when the heart rate decreased by more than 5 beats/min during two consecutive training sessions. For patients with severe hypoxemia, oxygen was administered during exercise to maintain SpO2 ≥ 90%. Patients also performed resistance exercises of lower and upper limbs (three sets of 8-12 repetitions at 60% of their maximal isometric force). The workload was adjusted every week and the intensity was increased up to 85% of maximal force. Patients also participated in educational courses and relaxation sessions, and received dietary counseling. However, neither protein or essential amino acids supplementation nor hypocaloric diet was employed during the study.

Vastus Lateralis Muscle Biopsy: Immunohistochemical and Biochemical Analyses

Muscle biopsy. Biopsy of the vastus lateralis muscle was performed with a Weil-Blakesley forceps 24 h before the first training session and 24 h after the last training session. Patients with hypoxemia breathed ambient air for at least 1 h before the biopsy. Posttraining biopsy was taken 2 cm away from the pretraining biopsy site.

Immunohistology and morphological analysis. Muscle samples mounted in embedding medium were cut (10 μm) in a cryostat microtome (HM 560; Microm, Walldorf, Germany) at −20°C. Sections were immunostained with antibodies against myosin heavy chain type I (A4.951; Alexis Biochemicals) and myosin heavy chain type IIa (N2.261; Alexis Biochemicals) as previously described (44). Fibers were classified as type I, Ila, I-IIa, or IIX fibers. The cross-sectional area of at least 50 type I and type II fibers per biopsy was determined. Microvessels were identified using a CD31 antibody (Dako, Les Ulis, France). The number of capillaries in contact with each fiber was counted and expressed as the capillary-to-fiber ratio (19). Muscle sections were visualized under a light microscope (Eclipse E400; Nikon, Bes Hoevedorp, The Netherlands) connected to a digital camera (Nikon Coolpix 990). Photographs were analyzed using ImageJ software (http://rsb.info.nih.gov/ij/, 1997–2014; National Institutes of Health, Bethesda, MD).

Protein and DNA analysis. Total RNA was extracted from 20–30 mg of skeletal muscle samples conditioned in RNAlater (Qiagen, Courtaboeuf, France) using the Total RNA isolation kit (Ambion, Austin, TX) followed by purification on an RNAasy silica spin column (Qiagen). Complementary DNA was generated from 400 ng of RNA using the Transcriptor First Strand cDNA synthesis kit (Roche Diagnostics, Mannheim, Germany). The selected forward and reverse primer sequences are listed in Table 1. Real-time quantitative polymerase chain reaction (qPCR) was performed in a 20-μl final volume and optimized concentrations for each primer using Sensimix SYBR & Fluorescein (GC Biotech, Alphen aan den Rijn, The Netherlands) and a MyiQ single-color real-time thermal cycler (Bio-Rad, Hercules, CA). Expression stability of seven reference genes (glyceraldehyde-3-phosphate dehydrogenase, β-actin, cyclophilin A, large ribosomal protein ribosomal protein P0, large ribosomal protein, ribosomal protein 13A, β2-microglobulin, tyrosine 3-monooxygenase/tryptophan 5-monooxygenase activation protein zeta polypeptide) was assessed using GENORM software (Gent University, Gent, Belgium) (14, 43). All references genes were used for normalization.

Protein isolation. Muscle samples were homogenized at 4°C in a 20-volume buffer consisting of 50 mM Tris HCl (pH 7.4), 100 mM NaCl, 2 mM EDTA, 2 mM EGTA, 50 mM β-glycerophosphate, 50 mM NaF, 1 mM sodium orthovanadate, 120 mM okadaic acid, and 1% Triton X-100. Homogenates were centrifuged at 12,000 g for 20 min at 4°C. Protein concentration of the supernatant was spectrophotometrically measured at 750 nm using the Bio-Rad protein assay (Marnes-la-Coquette, France).
Enzyme activities. Citrate synthase (CS, EC 4.1.3.7) and lactate dehydrogenase (LDH, EC 1.1.1.27) enzyme activities were fluorometrically determined ($\lambda_{\text{exc}} = 340$ nm and $\lambda_{\text{em}} = 450$ nm) (13). Cathepsin B + L (EC 3.4.22.1 and EC 3.4.22.15), chymotrypsin-like (EC 3.4.21.1), trypsin-like (EC 3.4.21.4), and caspase-like (EC 3.4.13.17) enzyme activities of 20S proteasomes were fluorometrically measured ($\lambda_{\text{exc}} = 380$ nm and $\lambda_{\text{em}} = 460$ nm) by cleavage of specific amido-4-methylcoumarin-coupled substrates (Bachem, Weil am Rhein, Germany) as previously described (6, 12).

Western immunoblotting. Proteins (50 µg) were separated on 12.5% SDS-PAGE and transferred onto 0.45-µm nitrocellulose membranes. Gel loading was systematically checked by Coomassie staining. Immunoblot analysis of proteins was performed as previously described (21) with the following primary antibodies against Akt (1:1,000), GSK-3 (1:1,000), p70S6K (1:1,000), and caspase-3 (1:1,000). The clinical outcome initially designed for the present study was a training effect on maximal power output. We performed a statistical power analysis and calculated that eight patients in each group was enough to detect a 10-W difference in peak workload with a standard deviation of 6 W ($\alpha = 0.05$, $\beta = 90\%$). Data are means ± SE. The effect of ExTr and hypoxemia on exercise capacity, muscle strength, protein content, and mRNA level was assessed by two-way ANOVA (Statview 5.0) followed by a Scheffé protected least significance difference test to detect specific mean differences. Akt, GSK-3β, and p70S6K phosphorylation levels between normoxemic and hypoxemic patients were compared by a Mann-Whitney test. For in vitro experiments, mRNA and phosphorylated proteins levels were compared by two-way ANOVA (time × absolute value) followed by a Scheffé post hoc test. Statistical difference was established at $P < 0.05$.

RESULTS

Baseline Patient Characteristics and Functional Benefits of ExTr

Patients displayed moderate to severe airway obstruction and mild to moderate lung hyperinflation (Table 2). Hypox-
The training sessions was also similar in normoxemic and hypoxemic groups, respectively. Duration of 15 kg/m² for women) was present in five normoxemic patients from corresponding normoxemic group. bSignificantly different from baseline.

### Table 2. Baseline morphometric, spirometric and blood gases characteristics in normoxemic and hypoxemic patients with COPD

<table>
<thead>
<tr>
<th>Subject Characteristic</th>
<th>Normoxemic Group</th>
<th>Hypoxemic Group</th>
</tr>
</thead>
<tbody>
<tr>
<td>n, sex ratio</td>
<td>15, 12M/3F</td>
<td>8, 8M</td>
</tr>
<tr>
<td>Age, yr</td>
<td>60.5 ± 1.9</td>
<td>60.4 ± 2.4</td>
</tr>
<tr>
<td>Gold stage I/II/IV</td>
<td>4/0/2</td>
<td>0/0/8</td>
</tr>
<tr>
<td>FEV₁, liter</td>
<td>1.18 ± 0.07</td>
<td>1.05 ± 0.13</td>
</tr>
<tr>
<td>FEV₁, % pred</td>
<td>42 ± 3</td>
<td>34 ± 4</td>
</tr>
<tr>
<td>FVC, liter</td>
<td>2.85 ± 0.22</td>
<td>3.35 ± 0.23</td>
</tr>
<tr>
<td>FVC, % pred</td>
<td>77 ± 7</td>
<td>87 ± 6</td>
</tr>
<tr>
<td>FEV₁/FVC,%</td>
<td>43 ± 3</td>
<td>31 ± 3</td>
</tr>
<tr>
<td>RV, liter</td>
<td>4.12 ± 0.32</td>
<td>5.10 ± 0.50</td>
</tr>
<tr>
<td>RV, % pred</td>
<td>186 ± 12</td>
<td>224 ± 22</td>
</tr>
<tr>
<td>TLC, liter</td>
<td>7.20 ± 0.47</td>
<td>8.51 ± 0.58</td>
</tr>
<tr>
<td>TLC, % pred</td>
<td>112 ± 10</td>
<td>131 ± 6</td>
</tr>
<tr>
<td>PaO₂, mmHg</td>
<td>68.5 ± 1.5</td>
<td>57.0 ± 1.0</td>
</tr>
<tr>
<td>PaCO₂, mmHg</td>
<td>35.9 ± 1.2</td>
<td>36.9 ± 1.8</td>
</tr>
<tr>
<td>SaO₂, %</td>
<td>95.1 ± 0.4</td>
<td>90.9 ± 0.8</td>
</tr>
</tbody>
</table>

COPD, chronic obstructive pulmonary disease; FEV₁, forced expiratory volume in 1 s; % pred, percentage of predicted value; FVC, forced vital capacity; RV, residual volume; TLC, total lung capacity; PaO₂, oxygen arterial blood pressure; PaCO₂, carbon dioxide arterial blood pressure; SaO₂, oxygen arterial saturation. Data are means ± SE. *Significantly different from corresponding normoxemic patients.

Hypoxic patients had a lower PaO₂ and a tendency toward more severe airway obstruction and higher hyperinflation (Table 2). BMI and FFMI were similar in both normoxemic and hypoxemic groups (Table 3). A depleted state (FFMI < 17 for men, 15 kg/m² for women) was present in five normoxemic patients and four hypoxemic patients.

Before ExTr, hypoxic patients had a lower Wpeak (Table 3). VO₂peak also tended to be lower in patients with hypoxemia. Relative training intensities at the end of the first month of the training program were similar in both groups (76.5 ± 7.7% and 66.2 ± 11.6% of pretraining Wpeak in normoxemic and hypoxemic patients, respectively). At the end of the training program, training intensities were similar in normoxemic and hypoxemic patients (83.6 ± 7% vs. 76.5 ± 7% of pretraining Wpeak for normoxemic and hypoxemic groups, respectively). Duration of the training sessions was also similar in normoxemic and hypoxic patients (36 ± 3 vs. 30 ± 2 min, and 36 ± 3 vs. 33 ± 1 min for normoxemic and hypoxic groups, respectively, at the end of the first and second month of ExTr). ExTr significantly increased Wpeak, illustrating the efficacy of the rehabilitation program (Table 3). The relative increase in Wpeak was not significantly different between groups. VO₂peak tended to increase, but it did not reach the significance level. The VO₂/workload relationship was not significantly different between groups and did not change after ExTr (10.1 ± 0.7 and 11.2 ± 1.5 ml·min⁻¹·W⁻¹ in normoxemic and hypoxic groups, respectively). Dyspnea and fatigue Borg scores were similar in both groups of patients during incremental cycling exercise before and after ExTr (Table 3). Quadriceps muscle force increased significantly in response to ExTr both in normoxemic and hypoxic patients (Table 3). Finally, handgrip force, used as a control test, remained unchanged in response to ExTr.

### Effects of ExTr on CS and LDH Activities, Muscle Fiber Type Distribution, Muscle Fiber Size, and Capillarization

CS activity was significantly increased in the normoxemic group in response to ExTr, whereas it remained relatively unchanged in the hypoxic group (Fig. 1A). LDH activity, which was significantly higher in hypoxic patients before ExTr, was increased only in normoxemic patients in response to ExTr (Fig. 1B).

Muscle fiber type distribution was not significantly different between groups before ExTr, and it remained unchanged in response to ExTr (Table 4). Muscle fiber cross-sectional area was significantly larger in hypoxic patients compared with normoxic patients before ExTr (Fig. 2). ExTr elicited a significant increase in muscle fiber cross-sectional area in the normoxic group, whereas muscle fiber cross-sectional area remained unchanged in hypoxic patients (Fig. 2). Before training, the capillary-to-fiber ratio was significantly higher in the hypoxic group compared with that in the normoxic group. Capillary-to-fiber ratio increased significantly in the normoxic group with ExTr, whereas it remained unchanged in the hypoxic group (Table 4). Normalized to muscle fiber

### Table 3. Body composition and exercise tolerance in normoxemic and hypoxemic patients with COPD before and after exercise training

<table>
<thead>
<tr>
<th></th>
<th>Normoxic</th>
<th>Hypoxic</th>
<th>Normoxic</th>
<th>Hypoxic</th>
</tr>
</thead>
<tbody>
<tr>
<td>Before ExTr</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>BMI, kg/m²</td>
<td>23.8 ± 1.1</td>
<td>23.0 ± 1.6</td>
<td>24.1 ± 1.0</td>
<td>23.0 ± 1.5</td>
</tr>
<tr>
<td>FFMI, kg/m²</td>
<td>18.0 ± 0.7</td>
<td>18.0 ± 0.8</td>
<td>17.9 ± 0.6</td>
<td>17.8 ± 0.9</td>
</tr>
<tr>
<td>Wpeak, W</td>
<td>63.7 ± 6.7</td>
<td>43.1 ± 4.3</td>
<td>72.0 ± 8.7</td>
<td>52.5 ± 3.0</td>
</tr>
<tr>
<td>Wpeak, % pred</td>
<td>41 ± 4</td>
<td>26 ± 3*</td>
<td>45 ± 5*</td>
<td>32 ± 3*</td>
</tr>
<tr>
<td>VO₂peak, ml/min</td>
<td>933 ± 75</td>
<td>816 ± 81</td>
<td>998 ± 89</td>
<td>864 ± 60</td>
</tr>
<tr>
<td>VO₂peak, % pred</td>
<td>51 ± 4</td>
<td>41 ± 4</td>
<td>52 ± 4</td>
<td>45 ± 7</td>
</tr>
<tr>
<td>HRpeak, %pred</td>
<td>79 ± 2</td>
<td>78 ± 4</td>
<td>79 ± 2</td>
<td>77 ± 5</td>
</tr>
<tr>
<td>VR, %</td>
<td>12 ± 6</td>
<td>5 ± 6</td>
<td>15 ± 7</td>
<td>13 ± 7</td>
</tr>
<tr>
<td>Dyspnea Borg score</td>
<td>6.4 ± 0.5</td>
<td>5.9 ± 0.7</td>
<td>6.4 ± 0.7</td>
<td>6.8 ± 0.7</td>
</tr>
<tr>
<td>Fatigue Borg score</td>
<td>5.0 ± 0.7</td>
<td>6.5 ± 0.7</td>
<td>4.4 ± 0.6</td>
<td>5.9 ± 0.46</td>
</tr>
<tr>
<td>QMF, N</td>
<td>329 ± 27</td>
<td>309 ± 29</td>
<td>392 ± 28b</td>
<td>401 ± 50b</td>
</tr>
<tr>
<td>Handgrip, N</td>
<td>359 ± 21</td>
<td>388 ± 27</td>
<td>395 ± 28</td>
<td>407 ± 34</td>
</tr>
<tr>
<td>After ExTr</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

BMI, body mass index; ExTr, exercise training; FFMI, fat-free mass index; W, power output; VO₂, oxygen uptake; HR, peak heart rate expressed as a percentage of predicted value; VR, ventilatory reserve calculated as (MVV – Vmax)/MVV (MVV was predicted as 35 × FEV₁); dyspnea and fatigue Borg scores, symptoms were recorded at peak cycling exercise; QMF, quadriceps muscle force measured in Newtons (N). Data are means ± SE. *Significantly different from corresponding normoxemic group. bSignificantly different from baseline.
cross-sectional area, capillary-to-fiber ratio was nonsignificantly increased in the normoxemic group (Table 4).

**Ubiquitin-Proteasome and Autophagy-Lysosome Pathways**

We first determined whether some critical players in the proteolytic pathways were differentially regulated between normoxemic and hypoxemic patients with COPD. Messenger RNA levels of MuRF1, Atrogin-1, and Nedd4 were not different between groups before or after ExTr (Fig. 3A). In agreement with these data, chymotrypsin-like enzyme activity of 20S proteasome remained unchanged with ExTr in both groups (Fig. 3B). Similarly, mRNA levels of autophagy-related genes (Beclin, LC3, Bnip, Gabarapl), as well as cathepsin B-L enzyme activity, were similar in both groups and did not change after ExTr (Fig. 3, C and D). Finally, the plasma level of procatabolic (IL-1β, IL-6, IL-8, TNF-α, IFN-γ) and anticoncatabolic (IL-10, IL-15) cytokines did not differ between groups and remained unchanged with ExTr (data not shown).

**Impaired Akt/mTOR Pathway Activation by ExTr in Hypoxemic Patients with COPD**

We next determined whether expression of known regulators of the Akt/mTOR pathway was differentially regulated in response to ExTr between normoxemic and hypoxemic patients. The transcript level of IGF-1, a positive regulator of the Akt/mTOR pathway (36), and myostatin, a negative regulator of the Akt/mTOR pathway (3), did not differ between normoxemic and hypoxemic patients with COPD before or after ExTr (Fig. 4, A and B).

We next investigated the phosphorylation status of several downstream mediators of the Akt/mTOR pathway. The phosphorylation levels of AktSer473, GSK-3βSer9, and p70S6kThr389 were differentially regulated in response to ExTr between normoxemic and hypoxemic patients. The relative changes in the phosphorylation level of these proteins were decreased in normoxemic patients with COPD before and after exercise training (ExTr).

Table 4. **Vastus lateralis muscle fiber type distribution and capillarization in normoxemic and hypoxemic patients with COPD before and after exercise training**

<table>
<thead>
<tr>
<th>Fiber Type</th>
<th>Before ExTr</th>
<th>After ExTr</th>
</tr>
</thead>
<tbody>
<tr>
<td>Type I, %</td>
<td>Normoxemic</td>
<td>Hypoxemic</td>
</tr>
<tr>
<td></td>
<td>20.8 ± 6.3</td>
<td>17.6 ± 4.9</td>
</tr>
<tr>
<td>Type I-IIa, %</td>
<td>8.7 ± 2.7</td>
<td>4.8 ± 0.7</td>
</tr>
<tr>
<td>Type IIA, %</td>
<td>69.9 ± 5.0</td>
<td>68.2 ± 4.8</td>
</tr>
<tr>
<td>Type IIX, %</td>
<td>4.2 ± 2.1</td>
<td>6.9 ± 2.7</td>
</tr>
<tr>
<td>Capillary-to-fiber ratio</td>
<td>2.76 ± 0.05</td>
<td>2.95 ± 0.06</td>
</tr>
<tr>
<td>CF/CSA</td>
<td>1.35 ± 0.14</td>
<td>1.43 ± 0.20</td>
</tr>
<tr>
<td></td>
<td>Normoxemic</td>
<td>Hypoxemic</td>
</tr>
<tr>
<td></td>
<td>28.6 ± 4.5</td>
<td>19.3 ± 3.1</td>
</tr>
<tr>
<td></td>
<td>7.6 ± 2.0</td>
<td>6.2 ± 0.7</td>
</tr>
<tr>
<td></td>
<td>59.5 ± 5.4</td>
<td>67.6 ± 3.3</td>
</tr>
<tr>
<td></td>
<td>10.9 ± 6.7</td>
<td>9.9 ± 3.9</td>
</tr>
<tr>
<td></td>
<td>3.20 ± 0.06</td>
<td>2.91 ± 0.06</td>
</tr>
<tr>
<td></td>
<td>2.08 ± 0.70</td>
<td>1.16 ± 0.09</td>
</tr>
</tbody>
</table>

CF/CSA, capillary-to-fiber ratio corrected to fiber cross-sectional area. Data are means ± SE. *Significantly different from corresponding normoxemic group. 

Fig. 1. Citrate synthase (A) and lactate dehydrogenase (B) enzyme activities in vastus lateralis muscle in normoxemic and hypoxemic patients with chronic obstructive pulmonary disease (COPD) before and after exercise training (ExTr). Data are means ± SE. *Significantly different between groups of patients before exercise training (ExTr). **Significantly different with ExTr within a group of patients.

Fig. 2. Vastus lateralis muscle fiber cross-sectional area in normoxemic and hypoxemic patients with COPD before and after ExTr. Data are means ± SE. *Significantly different between groups of patients pre ExTr. **Significantly different with ExTr within a group of patients.
hypoxic patients with COPD in response to ExTr. Total protein content of Akt, GSK-3\beta, and p70S6k did not differ between groups and remained unchanged with ExTr (data not shown).

**Hypoxia Abolished IGF-1-Induced Stimulation of the Akt/mTOR Pathway in C2C12 Myotubes**

These data suggest that hypoxemia could be a factor that contributes to limiting the capacity of the Akt/mTOR pathway to respond to ExTr. We therefore further tested this hypothesis by determining in vitro whether culturing myotubes in hypoxia for 48 h could limit or blunt stimulation of the Akt/mTOR pathway induced by IGF-1. When cultured in normoxia, addition of IGF-1 induced an immediate and sustained increase in the phosphorylation of Akt, GSK-3\beta, and p70S6K (Fig. 5A). By contrast, hypoxia completely prevented IGF-1-induced phosphorylation of Akt and GSK-3\beta. Phosphorylation of p70S6K was also decreased in response to IGF-1 addition in hypoxia-preconditioned myotubes. Finally, the transcript level of MuRF1 decreased significantly 180 min after IGF-1 addition, whereas the variation in the level of Atrogin-1 did not reach statistical significance (Fig. 5B). No difference was observed between culture conditions.

**DISCUSSION**

In the present study, we have demonstrated that ExTr induced similar functional benefits in exercise capacity in normoxic and hypoxic patients. However, our biochemical analyses indicated that muscle fiber hypertrophy and activation of the Akt/mTOR pathway by ExTr was impaired in the skeletal muscle of hypoxic patients with COPD. Furthermore, in vitro analysis using C2C12 myotubes indicated that hypoxia prevented activation of the Akt/mTOR pathway in response to IGF-1 addition, suggesting that hypoxemia could be a factor that contributes to limiting the extent of skeletal muscle response to ExTr in hypoxic patients with COPD.

The training program was effective at increasing maximal power output in both groups of patients. The extent of improvement was of comparable amplitude to that previously described in patients with moderate to severe COPD (34) and exceeded the minimal perceived difference (31). These data confirmed that ExTr can similarly improve the exercise capacity of normoxic and hypoxic patients with COPD.

Such an observation has been also reported in cachectic and noncachectic patients with COPD in response to pulmonary rehabilitation (45). Exercise tolerance is influenced by many factors, including motivation and habituation to the exercise test. After ExTr, desensitization to dyspnea also contributes to the improved exercise tolerance due to the alleviation of the discomfort of breathing (1, 4). That this phenomenon could occur in the hypoxic group in the present study during submaximal exercise could explain their better exercise tolerance and greater sustained workload without the need for peripheral adaptations.

The increase in CS and LDH activities in response to ExTr in skeletal muscle of normoxic patients with COPD strongly suggests an increase in their overall capacity to produce ATP during exercise by increasing both oxidative and anaerobic metabolic capacities. ExTr was thus efficient in eliciting metabolic adaptations in skeletal muscle of normoxic patients. By contrast, skeletal muscle of hypoxic patients seems to be refractory, in that both CS and LDH activities remained unchanged in response to ExTr. This is in agreement with previous reports showing that chronic exposure to hypoxia did
not change (or even decreased) mitochondrial enzyme activity (27, 35) and mitochondrial content of skeletal muscle (20). Overall, this suggests that the extent of skeletal muscle metabolic response to ExTr is altered in hypoxemic patients with COPD.

Surprisingly, hypoxemic patients with COPD had larger fiber cross-sectional area compared with normoxic patients. This difference still existed when female patients from the normoxic group were excluded from the analysis, thus ruling out a gender effect. A lower muscle fiber size in normoxic patients with COPD compared with those who were hypoxic before ExTr could have been beneficial by increasing the capillary-to-muscle fiber ratio and thus improving oxygen delivery to skeletal muscle (15). However, the observation that the capillary-to-fiber ratio normalized to muscle fiber cross-sectional area was not different between groups does not support this hypothesis. The reported increase in muscle fiber cross-sectional area with ExTr in normoxic patients is in agreement with a recent study showing that ExTr in normoxic patients with COPD increased muscle fiber cross-sec-

Fig. 4. Insulin-like growth factor (IGF)-1 mRNA level, myostatin mRNA level, and the Akt/mTOR pathway. IGF-1 (A) and myostatin (B) mRNA levels remained unchanged in the vastus lateralis muscle of normoxic and hypoxic patients with COPD before (open bars) and after (black bars) ExTr. Messenger RNA level was determined by relative quantification with real-time PCR. C: relative changes in phosphorylation levels of AktSer473, GSK-3βSer9, and p70S6KThr421/Ser424 in response to ExTr in normoxic and hypoxic patients. Representative blots before and after ExTr for each phosphorylated protein measured appear on the left. Data are means ± SE. *Significantly different with ExTr.
ional area by 11% (46). ExTr also increased muscle fiber capillarization in skeletal muscle of normoxemic patients. By contrast, skeletal muscle of hypoxemic patients was refractory to the effects of ExTr because both muscle fiber cross-sectional area and capillarization remained unchanged in response to ExTr. Taken together, these data suggest that ExTr increases oxygen delivery in normoxemic patients above the capacity of hypoxemic patients. Interestingly, Vogiatzis et al. (45) previously reported a lower degree of muscle fiber hypertrophy in cachectic patients with COPD compared with noncachectic patients in response to ExTr. Therefore, a resistance of skeletal muscle to the beneficial effects of ExTr in patients with COPD could be a common feature linked to the severity of the disease either appreciated by the extent of cachexia, or by the degree of hypoxia, or both.

Several hypotheses could be evoked to explain the differential regulation in muscle fiber cross-sectional area between normoxemic and hypoxemic patients with COPD. One may first argue that a difference in muscle fiber type distribution could affect muscle fiber cross-sectional area. However, muscle fiber type distribution was similar in both groups before and after ExTr. Second, this observation could also result from an increase in protein degradation in hypoxemic patients with COPD. In the present study, markers of both ubiquitin-proteasome and autophagy-lysosome pathways were unchanged in response to ExTr. The invasive nature of the muscle biopsy precluded the inclusion of an earlier time point during ExTr, so we cannot rule out the possibility that an adaptive response of both ubiquitin-proteasome and autophagy-lysosome pathways may have occurred earlier during the rehabilitation procedure. Third, the different regulation of the Akt/mTOR signaling pathway between normoxemic and hypoxemic patients with COPD could also contribute to explaining the increase in muscle fiber cross-sectional area in hypoxemic patients.

We do not have definite evidence to assume that hypoxemia impairs the adaptive response of skeletal muscle in patients with COPD, but several arguments suggest that hypoxemia could be involved in the unresponsiveness of hypoxemic patients with COPD. First, we previously showed that chronic hypoxia in rodents and severe hypoxemia in patients with COPD downregulated the Akt/mTOR pathway (18). Second, a short-term hypoxia exposure in healthy volunteers (3.5 h) has been associated with a blunted muscle protein synthesis in

![Fig. 5. Effects of hypoxia on phosphorylation levels of AktSer473, GSK-3βSer9, and p70S6KThr421/Ser424 (A) and transcript levels of MuRF1 and Atrogin-1 (B). C2C12 myotubes (5 days of differentiation) were cultured in either normoxia or in hypoxia for 48 h. Myotubes were harvested 60 (T60) and 180 (T180) min after IGF-1 addition (20 nM) and analyzed as described in MATERIALS AND METHODS. Myotubes without IGF-1 were used as controls (T0). Values are means ± SE.

aDifferent from T0 in the same culture condition. bDifferent from the same time point in normoxia.](https://journals.physiology.org/journal/jappl)
response to acute resistance exercise (16). Finally, our in vitro analyses on C2C12 myotubes showed that hypoxia per se almost completely abolished the response of the Akt/mTOR pathway to IGF-1. The mechanisms that could be involved in hypoxia-induced skeletal muscle resistance to an anabolic stimulus are currently unknown. However, a recent in vitro study indicated that hypoxia reduced the sensitivity of the IGF receptor, leading to a decreased activation of Akt in myoblasts (25). Furthermore, insulin receptor substrate-1 has been shown to be phosphorylated on serine or threonine residues in hypoxia, thus preventing further activation of the pathway (22, 41). Whether such a mechanism occurs in vivo in adult skeletal muscle deserves further experiments.

Study Limitations

The limited number of hypoxemic patients (n = 8) is acknowledged as a limitation in our study. The clinical outcome initially designed for the present study was a training effect on maximal power output. The statistical power of analysis indicated that eight patients were necessary to detect a 10-W difference in peak workload with a standard deviation of 6 W (α = 0.05, β = 90%). Recent studies have shown significant training-induced adaptations in muscle fiber cross-sectional area and protein phosphorylation (43), as well as gene expression in subgroups of 6 to 10 patients with COPD (33), suggesting that despite the limited number of subjects in the hypoxic group, this would have been enough to detect muscle adaptations.

Another time point of analysis would have also been very informative to further decipher the kinetic response of intracellular signaling events. However, for a number of ethical reasons essentially linked to the invasive nature of the muscle biopsy, this was not possible. Finally, we do not have definite evidence to assume that hypoxemia impairs the adaptive response of skeletal muscle in patients with COPD. Hypoxic patients received long-term oxygen therapy for at least 3 mo before inclusion. Furthermore, clinical guidelines recommend adding O2 during ExTr sessions to maintain arterial O2 saturation in the 88% to 90% range (28). This was carried out in the present study. Therefore, hypoxic patients lived and exercised with O2 supplementation, which could minimize the effect of muscle hypoxia.

In conclusion, although hypoxic patients with COPD retained the capacity to improve their exercise capacity in response to ExTr as much as normoxic patients did, hypoxic patients with COPD were resistant to ExTr-induced skeletal muscle adaptations.

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DISCLOSURES

No conflicts of interest, financial or otherwise, are declared by the authors.

AUTHOR CONTRIBUTIONS


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