

# Lymphocyte-Sparing Radiotherapy

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# Lymphocyte-Sparing Radiotherapy: The Rationale for Protecting Lymphocyte-rich Organs When Combining Radiotherapy With Immunotherapy

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There is now strong clinical and preclinical evidence that lymphocytes, for example, CD8<sup>+</sup> T cells, are key effectors of immunotherapy and that irradiation of large blood vessels, the heart, and lymphoid organs (including nodes, spleen, bones containing bone marrow, and thymus in children) causes transient or persistent lymphopenia. Furthermore, there is extensive clinical evidence, across multiple cancer sites and treatment modalities, that lymphopenia correlates strongly with decreased overall survival. At the moment, we lack quantitative evidence to establish the relationship between dose-volume and dose-rate to critical normal structures and lymphopenia. Therefore, we propose that data should be systematically recorded to characterise a possible quantitative relationship. This might enable us to improve the efficacy of radiotherapy and develop strategies to predict and prevent treatment-related lymphopenia. In anticipation of more quantitative data, we recommend the application of the principle of As Low As Reasonably Achievable to lymphocyte-rich regions for radiotherapy treatment planning to reduce the radiation doses to these structures, thus moving toward “Lymphocyte-Sparing Radiotherapy.”  
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## Introduction

The Pacific trial, a randomised phase 3 trial in non-metastatic, advanced non-small cell lung cancer (NSCLC), represented a breakthrough in immuno-oncology (IO) treatment

within radiation oncology, convincingly demonstrating that adjuvant IO, after normofractionated chemoradiotherapy, can improve progression-free survival (PFS).<sup>1</sup> Remarkably, the radiotherapy (RT) schedules of the Pacific trial were neither

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medical, Oncoradiomics, ptTheragnostic, Health Innovation Ventures and DualTpharma. He received an advisor/presenter fee and/or reimbursement of travel costs/external grant writing fee and/or in kind manpower contribution from Oncoradiomics, BHV, Merck and Convert pharmaceuticals. Dr. Lambin has shares in the company Oncoradiomics SA and Convert pharmaceuticals SA and is co-inventor of two issued patents with royalties on radiomics (PCT/NL2014/050248, PCT/NL2014/050728) licensed to Oncoradiomics and one issue patent on mtDNA (PCT/EP2014/059089) licensed to ptTheragnostic/DNAmito, three non-patentable invention (softwares) licensed to ptTheragnostic/DNAmito, Oncoradiomics and Health Innovation Ventures.

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standardised nor optimised, as these were based only on investigator or radiation oncologist choice for each individual patient (total dose 54 Gy-74 Gy). Separately, it has been shown that RT is a double-edged sword regarding immune effects: it has both an immunostimulatory effect but also an immunosuppressive effect.<sup>2</sup> IO might reduce or overrule this RT-related immunosuppression.<sup>1,3</sup> Furthermore, lower doses to the heart, circulating blood pool, and lymphoid organs are associated with reduced immunosuppressive effect.<sup>3,4</sup> It can thus be hypothesised that an optimised RT protocol has the potential to decrease the immunosuppressive effects of RT, for example, by reducing RT-related lymphopenia (LP).

Several studies have shown that low blood lymphocyte count at baseline, across a range of cancer types, is a negative predictor of outcome.<sup>5-11</sup> Furthermore, the presence of CD8<sup>+</sup> tumour infiltrating lymphocytes on pathology review is a well-established predictor of better overall survival.<sup>12-14</sup> Additionally, preclinical experiments with lymphocyte depletion, i.e. decreased CD4<sup>+</sup> and CD8<sup>+</sup> counts, have clearly established a causal relationship with reduced efficacy of RT and (radio)-IO.<sup>15</sup>

The effect of RT on LP is well-documented and has been extensively described for several decades.<sup>16,17</sup> Typically, LP is a transient phenomenon with a recovery within 3 months after RT, but in certain cases it can continue to persist even years after treatment<sup>18</sup> which has been correlated to RT dose, RT sites, (hyper)fractionation, adjuvant chemotherapy, and irradiated volume.<sup>4,11-13,19-23</sup> A causal relationship between RT-induced LP and adverse locoregional control or survival has been speculated but not confirmed.<sup>24</sup>

## The Radiobiology of Lymphocytes

Lymphocytes are located in the blood (circulating lymphocytes), in reservoir lymphoid organs such as the spleen, and the thymus (in children and teenagers), in lymph nodes, and in the bone marrow, which is continuously producing new lymphocytes. As noted, some tumours are infiltrated by lymphocytes. It is important to appreciate that lymphocytes are a highly heterogeneous cell population comprised of subgroups with different roles in the crosstalk of tumours and the host immune system. The most prominent cell type in anti-tumour immune responses are CD8<sup>+</sup> effector T cells,<sup>25</sup> reflected in their prognostic significance<sup>26</sup> and their use in adoptive T cell therapy.<sup>27</sup> T<sub>H</sub>1 polarised (CD4<sup>+</sup>),<sup>28</sup> as well as CD4<sup>+</sup> cytolytic T cells, have also been shown to induce strong anti-tumour responses.<sup>29</sup> On the other hand, regulatory T cells<sup>30</sup> and T<sub>H</sub>2 polarised CD4<sup>+</sup> T cells<sup>31</sup> have mostly been linked to pro-tumour effects.<sup>29</sup> There is contradictory data on the role of T<sub>H</sub>17 T cells<sup>29</sup> and cancer in cancer immune responses.<sup>32,33</sup>

It has long been known that lymphocytes are the most radiosensitive cells of the hematopoietic system, as well as the entire body.<sup>34</sup> This radiosensitivity is surprising for a nondividing cell type, but may be related to robust apoptotic response pathways. The lethal dose required to reduce the surviving fraction of circulating lymphocytes by 90% (LD90) is only 3

Gy.<sup>35</sup> 0.5 Gy already leads to significant cell death induction in lymphocytes. Such a dose could easily be reached in standard RT schedules. Yovino et al found that with a standard treatment of 60 Gy in 30 fractions for glioblastoma (GBM) treatment, during every fraction of RT, 5% of circulating cells receive >0.5 Gy,<sup>36</sup> summing up to >95% of circulating cells being exposed to >0.5 Gy over the 6 week treatment. The induced cell death is predominantly apoptosis.<sup>37</sup>

Importantly, different lymphocyte subtypes show distinct radiosensitivity.<sup>38-40</sup> Naive CD8<sup>+</sup> effector T cells are more sensitive than memory T cells,<sup>37,40,41</sup> while regulatory T cells are relatively resistant.<sup>40,42,43</sup> Furthermore, the state of T cells, the solid organs and the different location containing CD8<sup>+</sup> T cells also influences radiosensitivity.<sup>44,45</sup> T cells that are proliferating are more radioresistant than T cells in other state.<sup>44</sup> With regard to the organs, the parenchymal CD8<sup>+</sup> T cells in the solid lymphoid organs (lymph nodes and spleen) are found most radiosensitive, followed by those residing in liver and gut. The CD8<sup>+</sup> T cells located intratumourally have a higher radioresistance, an increased motility and IFN- $\gamma$  secretion compared to circulating CD8<sup>+</sup> T cells and T cells in unirradiated tumours.<sup>45</sup> This may be due to changes in the tumour microenvironment wherein TGF- $\beta$  is a key regulator in making the intratumoural T cells more radioresistant.<sup>45</sup> Similar differential effects have been observed concerning radiation dose rate<sup>46</sup> with high dose rates leading to less lymphocyte death.<sup>47,48</sup> These findings are well in line with clinical observations of decreased naive T cells and enriched regulatory T cells in patients undergoing RT.<sup>14,49-51</sup>

## Analysis of the Clinical Literature

In many trials, the Common Terminology Criteria for Adverse Events (CTCAE) is used to differentiate between LP Grade 1 (<~1000-800/mm<sup>3</sup>), Grade 2 (<800-500/mm<sup>3</sup>), Grade 3 (<500-200 mm<sup>3</sup>), and Grade 4 (<200/mm<sup>3</sup>). Clinical factors that are associated with LP and key findings regarding LP for various cancers (GBM, head and neck squamous cell carcinoma, nasopharyngeal cancer, NSCLC, SCLC, breast cancer, esophageal cancer, pancreatic cancer, hepatocellular cancer, cervical cancer)<sup>24</sup> are summarised below.

### Factors That Influences LP

A disbalance in immunosurveillance due to tumour suppressor systems can contribute to LP that is present before treatment.<sup>14</sup> Also immunosuppressive medication or cancer-related treatment can lead to pre- and post-treatment LP, for example, corticosteroids, tyrosine-kinase inhibitors, and immune checkpoint inhibitors.<sup>11,14</sup> In addition, patients with immune-related conditions, such as multiple comorbidities, autoimmune diseases, genetic disorders in innate or adaptive defense, or patients with a poor WHO performance state are known to have worse PFS and overall survival (OS), probably related to a sub-optimally functioning immune system.

Also, treatment factors such as RT and chemotherapy have been shown to influence incidence and severity of LP. Firstly, RT in general results in a lymphocyte reduction.

More specifically, hypofractionation results in less reduction than normo- or hyperfractionation. Yuan et al and Saito et al have found in a breast and a palliative cohort, respectively, that LP was correlated with the number of fractions, independent of overall dose.<sup>52,53</sup> Secondly, irradiating larger Gross Tumour Volumes in NSCLC patients has been associated with lower lymphocyte count but not with lower total leukocyte, neutrophil, or monocyte counts during RT.<sup>24</sup> Thirdly, if lymphopoietic sites or organs containing large blood volumes are within the planning target volume, it will contribute to (longer duration of) LP.<sup>14</sup> Several authors have also found that higher spleen irradiation doses (total dose of 50-60 Gy) were significantly correlated with more patients experiencing LP during RT for hepatocellular cancer or palliative RT.<sup>53-56</sup> Based on these results, Liu et al recommend sparing of the spleen during abdominal irradiation.<sup>54</sup> Furthermore, a lower heart and lung dose resulted in less LP.<sup>57-60</sup> Increasing the heart and lung dose, severe loss of cardiopulmonary performance was seen in preclinical studies.<sup>61-66</sup> Lastly, another important factor is the use of concurrent chemotherapy. Concurrent chemotherapy has been shown to have an impact on the severity of LP,<sup>22</sup> whereas adjuvant chemotherapy may prolong the duration of LP.<sup>22</sup> Importantly, different chemotherapy agents differ in LP impact.<sup>14</sup>

### Predictive/Prognostic Factors for OS After Radiation Induced LP

Many possible prognostic factors for OS and PFS have been investigated, including the role of LP. Ladbury et al concluded that estimated dose of radiation to immune cells, Karnofsky performance status, not-otherwise-specified histology in NSCLC, lack of completion of chemotherapy,<sup>9,23</sup> and smoking history<sup>23</sup> are negative predictors for OS.

Disadvantageous prognostic factors for PFS and OS are baseline LP,<sup>5-11,14</sup> early LP after chemotherapy treatment (5 or 15 days),<sup>14</sup> LP after RT<sup>14</sup> or LP after IO.<sup>7</sup> Post-treatment LP has been negatively associated with poor tumour specific outcome in multiple cancer types for example, GBM, HNSCC, cervical, esophageal, NSCLC, and pancreatic.<sup>11</sup>

### Effect of Combination Treatment (RT + chemo, RT + chemo and/or IO)

As described previously, RT alone can induce or worsen LP. However, combining RT with systemic treatment has an even bigger impact on LP and treatment outcome. Cho et al found that RT + checkpoint inhibitor -treated NSCLC patients with LP pre-IO treatment had a significant poorer PFS (2.2 vs 5.9 months) and OS (5.7 vs 12.1 months)<sup>10</sup> compared to patients who had normal lymphocyte counts before IO treatment. Furthermore they found that RT significantly increased LP before start of IO, however irradiating with SABR, proton beam therapy, hypofractionation or radiosurgery reduced the risk on (increasing) RT-induced LP.<sup>10,14,60</sup> The combination of RT with immunocytokines like IL2, IL7, or IL15 could eliminate LP due to their stimulatory effect to let the T cells develop, proliferate and survive.<sup>14</sup>

Joseph et al found that after concurrent chemo-radiotherapy the absolute lymphocyte count dropped significantly compared to absolute lymphocyte count pretreatment,<sup>4</sup> but did not alter treatment outcome. In contrast, Grossman et al observed worse tumour control and shorter OS in GBM patients with depleted CD4<sup>+</sup> T cell counts pre- and post-chemo-radiotherapy treatment.<sup>67</sup> Furthermore, a prolonged duration of LP was also seen with RT. Similar results were found retrospectively by Wang et al, with almost 50% of SCLC patients experiencing severe LP and 70.4% prolonged LP of 3 months minimum after chemo-radiotherapy.<sup>21</sup> For reasons not currently well understood, LP following RT can last from several months up to several years, whereas LP seen after sepsis or even chemotherapy alone tends to resolve more quickly.<sup>68,69</sup>

It is reasonable to hypothesise that transient LP has a different effect on the outcome than persistent LP. Thus, the negative influence of RT on LP might be abolished by combinatorial approaches with IO, which could result in differences in the timing, the length and probably the grade of LP. This effect also depends on type of IO agent applied. On the other hand, it might indicate that the effect of adding IO to RT schedules lies primarily in a better functioning immune system, which in turn will be crucial to slow down the pace of microscopic disease spread in at least some patients.

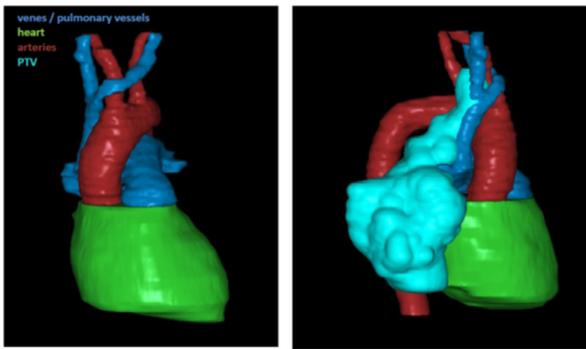
### Modelling Approaches to Predict the Incidence and Severity of LP

Taking into account the negative effect of LP on clinical outcomes, it is important to identify high-risk patients timely and possibly adapt the treatment. Models predicting grade 4 RT-induced LP during chemo (radio) therapy for esophageal cancer, or acute and late LP for prostate cancer have already been published,<sup>70,71</sup> although the prostate model is yet to be validated.<sup>19,72</sup> Also for NSCLC, a predictive risk model has been developed where clinical and genetic factors, for example, lung V5 > 48%, age >65 years, >40 pack-years, and XRCC1 rs25487 AA genotype, are associated with severe RT-induced LP.<sup>73</sup>

Several recent analyses have indicated that irradiation of cardiovascular structures may lead not just to heart-related morbidities but to unexplained reductions in OS following RT for NSCLC. A key question is whether this is mediated primarily through immune suppression. Contreras et al showed that adjuvant chemotherapy and heart V50 > 25% are associated with LP at 4 months post-RT.<sup>3</sup> Thor et al observed that out-of-treatment-field regional recurrence was statistically linked to LP at 2 months post-RT.<sup>74</sup> However, details of the relationship between patient/disease/treatment factors and LP, as well as the impact on disease progression remain elusive and need further study.

### Recommendations for Clinical Trials

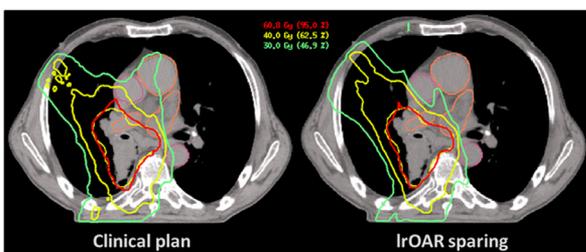
There is a large body of literature evidence showing that incidence and severity of LP are associated with patient and



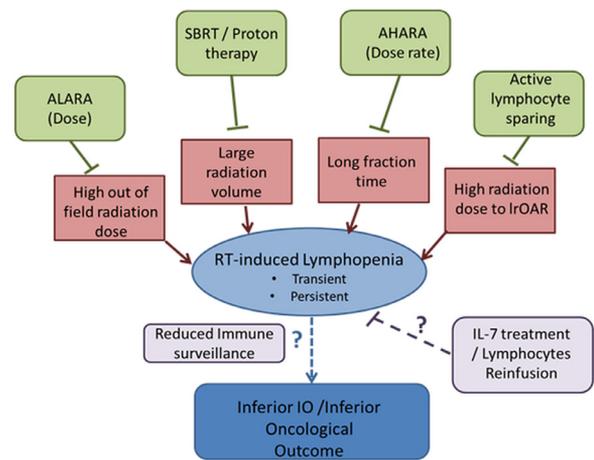
**Figure 1** Example of segmentation for lung cancer treatment: left: delineation of the Lymphocyte-related Organs At Risk (LOAR), right delineation of the LOAR and the planning target volume.

treatment characteristics, but also showing the importance for clinical outcomes. Moreover, we have identified 437 trials listed in clinicaltrial.gov combining IO with RT, September 2019, indicating that combining RT with IO is being increasingly adopted as treatment strategy. To improve clinical outcomes, but also to gain the most of RT-IO combination treatment, it is of utmost importance to establish recommendations for RT planning with regard to lymphocyte dose. However, as indicating absolute dose constraints is not (yet) possible, we propose to apply the As Low As Reasonably Achievable principle to Lymphocyte-related Organs At Risk (LOARs) without compromising irradiation of the planning target volume (see Figs. 1 and 2) and keeping the constraints for “conventional” organs at risk such as lung, heart and spinal cord, as recommended in clinical protocols (Fig. 3). Furthermore, systematic recording of dose-volume and dose-rate statistics for those LOARs, as well as longitudinal lymphocyte counts is recommended. These data, routinely available at most treatment centres, would allow the design of strategies to predict and to some extent prevent RT-induced LP. It would also help to answer the main remaining hypothesis whether maintaining and/or restoring optimal lymphocyte counts may improve treatment RT outcomes, or increase the efficacy of IO.

These data can only be obtained if relevant organs are systematically delineated. These include the large vessels, heart,



**Figure 2** A standard dose distribution of a clinically applied radiation treatment plan (left), and an example of an optimised radiation plan applying the As Low As Reasonably Achievable (ALARA) principle (right), demonstrating that sparing of LOAR is feasible without compromising dose coverage of the target volume or increasing dose to OARs important in clinical radiotherapy planning.



**Figure 3** Hypothetical model linking radiation to lymphopenia and to inferior oncological outcomes.

and any irradiated lymphoid organs such as bone marrow (eg, pelvic bones, vertebrae, large long bones), nodal regions not included in the clinical target volume, spleen, and thymus in children. To facilitate the segmentation of large vessels, we propose to explore the use of contrast-enhanced computed tomography, acquiring data during the early blood dominated phases. Automatic segmentation methods based on deep learning will certainly facilitate this process.<sup>75</sup> Dose, fractionation, dose rate, and mean doses to LOARs should be reported as a minimum. Blood can be seen as a “moving OAR”, therefore long irradiation times should be avoided. Instead, high-dose rate irradiation, following the principle of “As High As Reasonably Achievable” should be favoured, for example, using flattening filter-free irradiation.<sup>76,77</sup>

## Prospects

As it is clear that the role of the immune system is very important for clinical outcomes, much research currently focuses on unraveling the complex interplay between treatment characteristics and the immune system and how to influence this relationship. In an attempt to preserve the immune system from the effects of radiation and chemotherapy, lymphocytes were isolated before treatment, stored, and administered again to the patient upon treatment completion (NCT01653834).<sup>67</sup> Interestingly, the promising therapeutic effect of immunoadjuvant therapy with IL7 (essential for lymphocyte proliferation and survival) has been explored in for example, immunocompromised patient and in some cancer trials, however the data regarding IL7 and LP during, pre, and post cancer treatment are scarce.<sup>14,78-82</sup>

New imaging methods may also become important. New magnetic resonance (MR) sequences may enable the investigator to quantify blood volume in vessels and organs using non-contrast MR imaging such as a venography technique or velocity-selective pulse trains.<sup>83,84</sup> These new approaches will allow us not only to quantify blood volume without contrast in the vascular system but also in organs such as liver, brain and

spleen. New positron emission tomography tracers that can precisely track CD8<sup>+</sup> T cells are also under development.<sup>85</sup> Furthermore, the combination of new strategies and precise technological developments,<sup>20</sup> such as a MR linear accelerator (MR-linac),<sup>86</sup> will make it possible to not only more precisely identify and track LOARs, but also avoid or restrict radiation dose to these LOARs. To facilitate comparable analyses, new autosegmentation and artificial intelligence methods could be distributed using portable container technology to extract dosimetric characteristics of the LOARs.<sup>87</sup>

## Conclusion

The breakthrough improvement in outcomes by IO alone, or in combination with RT, has renewed the interest of the scientific community in strategies to predict and avoid RT-associated LP that may be immunosuppressive. There is a convergence of preclinical and clinical evidence correlating unintentional irradiation of LOARs with LP and poor outcomes. Preclinical studies definitively show an established causal relationship between lymphocyte depletion and the effectiveness of IO. However, accurate, individualised normal tissue complication probability models for LP are currently lacking. Therefore, we propose that the As Low As Reasonably Achievable principle should be applied to LOARs, and dose rates should be kept as high as practical possible to spare peripheral blood lymphocytes, in particular in the context of clinical trials combining RT with IO. Furthermore, we urge investigators of clinical RT trials with an immune component to systematically record the potentially-relevant dosimetric and hematopoietic parameters. Such unique data will hopefully lead to predictive models that will allow us to predict and prevent RT-induced LP in an individualised approach for each patient in order to answer the key unresolved question: whether maintaining and/or restoring optimal lymphocyte counts independently improves RT or IO outcomes.

## References

1. Antonia SJ, Villegas A, Daniel D, et al: Durvalumab after Chemoradiotherapy in Stage III Non-Small-Cell Lung Cancer. *N Engl J Med* 377:1919-1929, 2017
2. Formenti SC, Demaria S: Systemic effects of local radiotherapy. *Lancet Oncol* 10:718-726, 2009
3. Contreras JA, Lin AJ, Weiner A, et al: Cardiac dose is associated with immunosuppression and poor survival in locally advanced non-small cell lung cancer. *Radiother Oncol* 128:498-504, 2018
4. Joseph N, McWilliam A, Kennedy J, et al: Post-treatment lymphocytopenia, integral body dose and overall survival in lung cancer patients treated with radical radiotherapy. *Radiother Oncol* 135:115-119, 2019
5. Grassberger C, Hong TS, Hato T, et al: Differential Association Between Circulating Lymphocyte Populations With Outcome After Radiation Therapy in Subtypes of Liver Cancer. *Int J Radiat Oncol Biol Phys* 101:1222-1225, 2018
6. Karantanos T, Karanika S, Seth B, et al: The absolute lymphocyte count can predict the overall survival of patients with non-small cell lung cancer on nivolumab: a clinical study. *Clin Transl Oncol* 21:206-212, 2019
7. Diehl A, Yarchoan M, Hopkins A, et al: Relationships between lymphocyte counts and treatment-related toxicities and clinical responses in patients with solid tumors treated with PD-1 checkpoint inhibitors. *Oncotarget* 8:114268-114280, 2017
8. Ho WJ, Yarchoan M, Hopkins A, et al: Association between pretreatment lymphocyte count and response to PD1 inhibitors in head and neck squamous cell carcinomas. *J Immunother Cancer* 6:84, 2018
9. Suzuki R, Wei X, Allen PK, et al: Prognostic Significance of Total Lymphocyte Count, Neutrophil-to-lymphocyte Ratio, and Platelet-to-lymphocyte Ratio in Limited-stage Small-cell Lung Cancer. *Clin Lung Cancer* 20:117-123, 2019
10. Cho Y, Park S, Byun HK, et al: Impact of Treatment-Related Lymphopenia on Immunotherapy for Advanced Non-Small Cell Lung Cancer. *Int J Radiat Oncol Biol Phys* 105:1065-1073, 2019
11. Kleinberg L, Sloan L, Grossman S, et al: Radiotherapy, Lymphopenia, and Host Immune Capacity in Glioblastoma: A Potentially Actionable Toxicity Associated With Reduced Efficacy of Radiotherapy. *Neurosurgery* 85:441-453, 2019
12. Yu PC, Long D, Liao CC, et al: Association between density of tumor-infiltrating lymphocytes and prognoses of patients with gastric cancer. *Medicine (Baltimore)* 97:e11387, 2018
13. Ibrahim EM, Al-Foheidi ME, Al-Mansour MM, et al: The prognostic value of tumor-infiltrating lymphocytes in triple-negative breast cancer: a meta-analysis. *Breast Cancer Res Treat* 148:467-476, 2014
14. Menetrier-Caux C, Ray-Coquard I, Blay JY, et al: Lymphopenia in Cancer Patients and its Effects on Response to Immunotherapy: an opportunity for combination with Cytokines? *J Immunother Cancer* 7:85, 2019
15. Zegers CM, Rekers NH, Quaden DH, et al: Radiotherapy combined with the immunocytokine L19-IL2 provides long-lasting antitumor effects. *Clin Cancer Res* 21:1151-1160, 2015
16. Raben M, Walach N, Galili U, et al: The effect of radiation therapy on lymphocyte subpopulations in cancer patients. *Cancer* 37:1417-1421, 1976
17. Newman GH, Rees GJ, Jones RS, et al: Changes in helper and suppressor T lymphocytes following radiotherapy for breast cancer. *Clin Radiol* 38:191-193, 1987
18. Petrini B, Wasserman J, Blomgren H, et al: Blood lymphocyte subpopulations in breast cancer patients following radiotherapy. *Clin Exp Immunol* 29:36-42, 1977
19. Yang TJ, Oh JH, Apte A, et al: Clinical and dosimetric predictors of acute hematologic toxicity in rectal cancer patients undergoing chemoradiotherapy. *Radiother Oncol* 113:29-34, 2014
20. Suryadevara CM, Desai R, Abel ML, et al: Temozolomide lymphodepletion enhances CAR abundance and correlates with antitumor efficacy against established glioblastoma. *Oncoimmunology* 7, 2018:e1434464
21. Wang X, Lu J, Teng F, et al: Lymphopenia association with accelerated hyperfractionation and its effects on limited-stage small cell lung cancer patients' clinical outcomes. *Ann Transl Med* 7:385, 2019
22. Lin AJ, Campian JL, Hui C, et al: Impact of concurrent versus adjuvant chemotherapy on the severity and duration of lymphopenia in glioma patients treated with radiation therapy. *J Neurooncol* 136:403-411, 2018
23. Ladbury CJ, Rusthoven CG, Camidge DR, et al: Impact of Radiation Dose to the Host Immune System on Tumor Control and Survival for Stage III Non-Small Cell Lung Cancer Treated with Definitive Radiation Therapy. *Int J Radiat Oncol Biol Phys* 105:346-355, 2019
24. Venkatesulu BP, Mallick S, Lin SH, et al: A systematic review of the influence of radiation-induced lymphopenia on survival outcomes in solid tumors. *Crit Rev Oncol Hematol* 123:42-51, 2018
25. Reiser J, Banerjee A: Effector, Memory, and Dysfunctional CD8(+) T Cell Fates in the Antitumor Immune Response. *Journal of immunology research* 2016, 2016:8941260
26. Galon J, Costes A, Sanchez-Cabo F, et al: Type, density, and location of immune cells within human colorectal tumors predict clinical outcome. *Science (New York, NY)* 313:1960-1964, 2006
27. Yee C, Thompson JA, Byrd D, et al: Adoptive T cell therapy using antigen-specific CD8+ T cell clones for the treatment of patients with metastatic melanoma: in vivo persistence, migration, and antitumor effect of transferred T cells. *Proceedings of the National Academy of Sciences of the United States of America* 99:16168-16173, 2002
28. Corthay A, Skovseth DK, Lundin KU, et al: Primary antitumor immune response mediated by CD4+ T cells. *Immunity* 22:371-383, 2005
29. Kim HJ, Cantor H: CD4 T-cell subsets and tumor immunity: the helpful and the not-so-helpful. *Cancer immunology research* 2:91-98, 2014

30. Banerjee A, Vasanthakumar A, Grigoriadis G: Modulating T regulatory cells in cancer: how close are we? *Immunol Cell Biol* 91:340-349, 2013
31. Tatsumi T, Kierstead LS, Ranieri E, et al: Disease-associated bias in T helper type 1 (Th1)/Th2 CD4(+) T cell responses against MAGE-6 in HLA-DRB1\*0401(+) patients with renal cell carcinoma or melanoma. *J Exp Med* 196:619-628, 2002
32. Namm JP, Li Q, Lao X, et al: B lymphocytes as effector cells in the immunotherapy of cancer. *Journal of surgical oncology* 105:431-435, 2012
33. Zhang Y, Morgan R, Podack ER, et al: B cell regulation of anti-tumor immune response. *Immunologic research* 57:115-124, 2013
34. Trowell OA: The sensitivity of lymphocytes to ionising radiation. *J Pathol Bacteriol* 64:687-704, 1952
35. Nakamura N, Kusunoki Y, Akiyama M: Radiosensitivity of CD4 or CD8 positive human T-lymphocytes by an in vitro colony formation assay. *Radiat Res* 123:224-227, 1990
36. Yovino S, Kleinberg L, Grossman SA, et al: The etiology of treatment-related lymphopenia in patients with malignant gliomas: modeling radiation dose to circulating lymphocytes explains clinical observations and suggests methods of modifying the impact of radiation on immune cells. *Cancer Invest* 31:140-144, 2013
37. Grayson JM, Harrington LE, Lanier JG, et al: Differential sensitivity of naive and memory CD8+ T cells to apoptosis in vivo. *Journal of immunology* (Baltimore, Md. 1950) 169:3760-3770, 2002
38. Williams JL, Patchen ML, Darden JH, et al: Effects of radiation on survival and recovery of T lymphocyte subsets in C3H/HeN mice. *Experimental hematology* 22:510-516, 1994
39. Zarybnicka L, Vavrova J, Havelek R, et al: Lymphocyte subsets and their H2AX phosphorylation in response to in vivo irradiation in rats. *International journal of radiation biology* 89:110-117, 2013
40. Pugh JL, Sukhina AS, Seed TM, et al: Histone deacetylation critically determines T cell subset radiosensitivity. *Journal of immunology* (Baltimore, Md. 1950) 193:1451-1458, 2014
41. Tabi Z, Spary LK, Coleman S, et al: Resistance of CD45RA- T cells to apoptosis and functional impairment, and activation of tumor-antigen specific T cells during radiation therapy of prostate cancer. *Journal of immunology* 185:1330-1339, 2010
42. Persa E, Balogh A, Safrany G, et al: The effect of ionizing radiation on regulatory T cells in health and disease. *Cancer letters* 368:252-261, 2015
43. Gururangan S, Reap E, Schmittling R, et al: Regulatory T cell subsets in patients with medulloblastoma at diagnosis and during standard irradiation and chemotherapy (PBTC N-11). *Cancer Immunol Immunother* 66:1589-1595, 2017
44. Heylmann D, Badura J, Becker H, et al: Sensitivity of CD3/CD28-stimulated versus non-stimulated lymphocytes to ionizing radiation and genotoxic anticancer drugs: key role of ATM in the differential radiation response. *Cell Death Dis* 9:1053, 2018
45. Arina A, Beckett M, Fernandez C, et al: Tumor-reprogrammed resident T cells resist radiation to control tumors. *Nat Commun* 10:3959, 2019
46. Gridley DS, Peca MJ, Dutta-Roy R, et al: Dose and dose rate effects of whole-body proton irradiation on leukocyte populations and lymphoid organs: part I. *Immunology letters* 80:55-66, 2002
47. Sterzing F, Munter MW, Schafer M, et al: Radiobiological investigation of dose-rate effects in intensity-modulated radiation therapy. *Strahlentherapie und Onkologie* 181:42-48, 2005
48. Ware JH, Sanzari J, Avery S, et al: Effects of proton radiation dose, dose rate and dose fractionation on hematopoietic cells in mice. *Radiat Res* 174:325-330, 2010
49. Eckert F, Schaedle P, Zips D, et al: Impact of curative radiotherapy on the immune status of patients with localized prostate cancer. *Oncoimmunology* 7, 2018:E1496881
50. Sage EK, Schmid TE, Geinitz H, et al: Effects of definitive and salvage radiotherapy on the distribution of lymphocyte subpopulations in prostate cancer patients. *Strahlentherapie und Onkologie* 193:648-655, 2017
51. Schaeue D, Comin-Anduix B, Ribas A, et al: T-cell responses to survivin in cancer patients undergoing radiation therapy. *Clin Cancer Res* 14:4883-4890, 2008
52. Yuan C, Wang Q: Comparative analysis of the effect of different radiotherapy regimes on lymphocyte and its subpopulations in breast cancer patients. *Clin Transl Oncol* 20:1219-1225, 2018
53. Saito T, Toya R, Matsuyama T, et al: Dosimetric Predictors of Treatment-related Lymphopenia induced by Palliative Radiotherapy: Predictive Ability of Dose-volume Parameters based on Body Surface Contour. *Radiol Oncol* 51:228-234, 2017
54. Liu J, Zhao Q, Deng W, et al: Radiation-related lymphopenia is associated with spleen irradiation dose during radiotherapy in patients with hepatocellular carcinoma. *Radiat Oncol* 12:90, 2017
55. Chadha AS, Liu G, Chen HC, et al: Does Unintentional Splenic Radiation Predict Outcomes After Pancreatic Cancer Radiation Therapy? *Int J Radiat Oncol Biol Phys* 97:323-332, 2017
56. Chadha AS, Suh Y, Krishnan S: In Reply to Yazici et al. *Int J Radiat Oncol Biol Phys* 98:485-486, 2017
57. Shiraishi Y, Fang P, Xu C, et al: Severe lymphopenia during neoadjuvant chemoradiation for esophageal cancer: A propensity matched analysis of the relative risk of proton versus photon-based radiation therapy. *Radiother Oncol* 128:154-160, 2018
58. Fang P, Jiang W, Davuluri R, et al: High lymphocyte count during neoadjuvant chemoradiotherapy is associated with improved pathologic complete response in esophageal cancer. *Radiother Oncol* 128:584-590, 2018
59. Fang P, Shiraishi Y, Verma V, et al: Lymphocyte-Sparing Effect of Proton Therapy in Patients with Esophageal Cancer Treated with Definitive Chemoradiation. *Int J Part Ther* 4:23-32, 2018
60. Badiyan SN, Robinson CG, Bradley JD: Radiation Toxicity in Lung Cancer Patients: The Heart of the Problem? *Int J Radiat Oncol Biol Phys* 104:590-592, 2019
61. van Luijk P, Novakova-Jiresova A, Faber H, et al: Radiation damage to the heart enhances early radiation-induced lung function loss. *Cancer research* 65:6509-6511, 2005
62. van Luijk P, Faber H, Meertens H, et al: The impact of heart irradiation on dose-volume effects in the rat lung. *Int J Radiat Oncol Biol Phys* 69:552-559, 2007
63. van Rongen E, Tan CH, Durham SK: Late functional, biochemical and histological changes in the rat lung after fractionated irradiation to the whole thorax. *Radiother Oncol* 10:231-246, 1987
64. Ghobadi G, van der Veen S, Bartelds B, et al: Physiological interaction of heart and lung in thoracic irradiation. *Int J Radiat Oncol Biol Phys* 84:e639-e646, 2012
65. Peterson LM, Evans ML, Thomas KL, et al: Vascular response to fractionated irradiation in the rat lung. *Radiat Res* 131:224-226, 1992
66. Ghobadi G, Bartelds B, van der Veen SJ, et al: Lung irradiation induces pulmonary vascular remodelling resembling pulmonary arterial hypertension. *Thorax* 67:334-341, 2012
67. Feasibility of Lymphocyte Reinfusion in Newly Diagnosed High Grade Gliomas. *ClinicalTrials.gov*. [<https://clinicaltrials.gov/ct2/show/NCT01653834>]
68. Dean RM, Fry T, Mackall C, et al: Association of serum interleukin-7 levels with the development of acute graft-versus-host disease. *J Clin Oncol* 26:5735-5741, 2008
69. Llano A, Barretina J, Gutierrez A, et al: Interleukin-7 in plasma correlates with CD4 T-cell depletion and may be associated with emergence of syncytium-inducing variants in human immunodeficiency virus type 1-positive individuals. *J Virol* 75:10319-10325, 2001
70. Sini C, Fiorino C, Perna L, et al: Dose-volume effects for pelvic bone marrow in predicting hematological toxicity in prostate cancer radiotherapy with pelvic node irradiation. *Radiother Oncol* 118:79-84, 2016
71. van Rossum PSN, Deng W, Routman DM, et al: Prediction of Severe Lymphopenia During Chemoradiation Therapy for Esophageal Cancer: Development and Validation of a Pretreatment Nomogram. *Pract Radiat Oncol* 10:E16-E26, 2019
72. Julie DA, Oh JH, Apte AP, et al: Predictors of acute toxicities during definitive chemoradiation using intensity-modulated radiotherapy for anal squamous cell carcinoma. *Acta Oncol* 55:208-216, 2016
73. Xie X, Lin SH, Welsh JW, et al: Radiation-induced lymphopenia during chemoradiation therapy for non-small cell lung cancer is linked with

- age, smoking, lung V5, and XRCC1 rs25487 genotype in lymphocytes. *Int J Radiat Oncol Phys Biol* 105:E538-E539, 2019
74. Thor M, Montovano M, Hotca A, et al: Are unsatisfactory outcomes after concurrent chemoradiotherapy for locally advanced non-small cell lung cancer due to treatment-related immunosuppression? *Radiotherapy and Oncology* in press <https://doi.org/10.1016/j.radonc.2019.1007.1016>
  75. Huang Q, Sun J, Ding H, et al: Robust liver vessel extraction using 3D U-Net with variant dice loss function. *Comput Biol Med* 101:153-162, 2018
  76. Yan Y, Yadav P, Bassetti M, et al: Dosimetric differences in flattened and flattening filter-free beam treatment plans. *J Med Phys* 41:92-99, 2016
  77. Dubois L, Biemans R, Reniers B, et al: High dose rate and flattening filter free irradiation can be safely implemented in clinical practice. *International journal of radiation biology* 91:778-785, 2015
  78. Francois B, Jeannot R, Daix T, et al: Interleukin-7 restores lymphocytes in septic shock: the IRIS-7 randomized clinical trial. *JCI Insight* 3, 2018
  79. Levy Y, Lacabaratz C, Weiss L, et al: Enhanced T cell recovery in HIV-1-infected adults through IL-7 treatment. *J Clin Invest* 119:997-1007, 2009
  80. Rosenberg SA, Sportes C, Ahmadzadeh M, et al: IL-7 administration to humans leads to expansion of CD8+ and CD4+ cells but a relative decrease of CD4+ T-regulatory cells. *J Immunother* 29:313-319, 2006
  81. Sportes C, Babb RR, Krumlauf MC, et al: Phase I study of recombinant human interleukin-7 administration in subjects with refractory malignancy. *Clin Cancer Res* 16:727-735, 2010
  82. Tredan O, Menetrier-Caux C, Ray-Coquard I, et al: ELYPSE-7: a randomized placebo-controlled phase IIa trial with CYT107 exploring the restoration of CD4+ lymphocyte count in lymphopenic metastatic breast cancer patients. *Ann Oncol* 26:1353-1362, 2015
  83. Aguirre-Reyes DF, Sotelo JA, Arab JP, et al: Intrahepatic portal vein blood volume estimated by non-contrast magnetic resonance imaging for the assessment of portal hypertension. *Magn Reson Imaging* 33:970-977, 2015
  84. Liu D, Xu F, Lin DD, et al: Quantitative measurement of cerebral blood volume using velocity-selective pulse trains. *Magn Reson Med* 77:92-101, 2017
  85. Colevas AD, Bedi N, Chang S, et al: A study to evaluate immunological response to PD-1 inhibition in squamous cell carcinoma of the head and neck (SCCHN) using novel PET imaging with [18F]F-AraG. *Journal of Clinical Oncology* 36, 2018. 6050-6050
  86. Tijssen RHN, Philippens MEP, Paulson ES, et al: MRI commissioning of 1.5T MR-linac systems - a multi-institutional study. *Radiother Oncol* 132:114-120, 2019
  87. Apte AP, Iyer A, Thor M, et al: Library of model implementations for sharing deep-learning image segmentation and outcomes models. *bioRxiv* 773929, 2019