

# Validation of the doubly labeled water method using off-axis integrated cavity output spectroscopy and isotope ratio mass spectrometry

Citation for published version (APA):

Melanson, E. L., Swibas, T., Kohrt, W. M., Catenacci, V. A., Creasy, S. A., Plasqui, G., Wouters, L., Speakman, J. R., & Berman, E. S. F. (2018). Validation of the doubly labeled water method using off-axis integrated cavity output spectroscopy and isotope ratio mass spectrometry. *American Journal of Physiology : Endocrinology and Metabolism*, 314(2), E124-E130.  
<https://doi.org/10.1152/ajpendo.00241.2017>

## Document status and date:

Published: 01/02/2018

## DOI:

[10.1152/ajpendo.00241.2017](https://doi.org/10.1152/ajpendo.00241.2017)

## Document Version:

Publisher's PDF, also known as Version of record

## Document license:

Taverne

## Please check the document version of this publication:

- A submitted manuscript is the version of the article upon submission and before peer-review. There can be important differences between the submitted version and the official published version of record. People interested in the research are advised to contact the author for the final version of the publication, or visit the DOI to the publisher's website.
- The final author version and the galley proof are versions of the publication after peer review.
- The final published version features the final layout of the paper including the volume, issue and page numbers.

[Link to publication](#)

## General rights

Copyright and moral rights for the publications made accessible in the public portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

- Users may download and print one copy of any publication from the public portal for the purpose of private study or research.
- You may not further distribute the material or use it for any profit-making activity or commercial gain
- You may freely distribute the URL identifying the publication in the public portal.

If the publication is distributed under the terms of Article 25fa of the Dutch Copyright Act, indicated by the "Taverne" license above, please follow below link for the End User Agreement:

[www.umlib.nl/taverne-license](http://www.umlib.nl/taverne-license)

## Take down policy

If you believe that this document breaches copyright please contact us at:

[repository@maastrichtuniversity.nl](mailto:repository@maastrichtuniversity.nl)

providing details and we will investigate your claim.

Download date: 16 Jul. 2024

## RESEARCH ARTICLE

# Validation of the doubly labeled water method using off-axis integrated cavity output spectroscopy and isotope ratio mass spectrometry

Edward L. Melanson,<sup>1,2</sup> Tracy Swibas,<sup>1</sup> Wendy M. Kohrt,<sup>1,2</sup> Vicki A. Catenacci,<sup>1</sup> Seth A. Creasy,<sup>1</sup> Guy Plasqui,<sup>3</sup> Loek Wouters,<sup>3</sup> John R. Speakman,<sup>4,5</sup> and Elena S. F. Berman<sup>6</sup>

<sup>1</sup>Divisions of Endocrinology, Metabolism, and Diabetes and Geriatric Medicine, Department of Medicine, University of Colorado Anschutz Medical Campus, Aurora, Colorado; <sup>2</sup>Geriatric Research, Education, and Clinical Center, Veterans Affairs Eastern Colorado Health Care System, Denver, Colorado; <sup>3</sup>NUTRIM School of Nutrition and Translational Research in Metabolism, Maastricht University, Maastricht, The Netherlands; <sup>4</sup>Institute of Biological and Environmental Sciences, Aberdeen University, Aberdeen, United Kingdom; <sup>5</sup>State Key Laboratory of Molecular Developmental Biology, Institute of Genetics and Developmental Biology, Chinese Academy of Sciences, Beijing, China; and <sup>6</sup>Los Gatos Research/ABB, San Jose, California

Submitted 13 July 2017; accepted in final form 25 September 2017

**Melanson EL, Swibas T, Kohrt WM, Catenacci VA, Creasy SA, Plasqui G, Wouters L, Speakman JR, Berman ES.** Validation of the doubly labeled water method using off-axis integrated cavity output spectroscopy and isotope ratio mass spectrometry. *Am J Physiol Endocrinol Metab* 314: E124–E130, 2018. First published October 3, 2017; doi:10.1152/ajpendo.00241.2017.—When the doubly labeled water (DLW) method is used to measure total daily energy expenditure (TDEE), isotope measurements are typically performed using isotope ratio mass spectrometry (IRMS). New technologies, such as off-axis integrated cavity output spectroscopy (OA-ICOS) provide comparable isotopic measurements of standard waters and human urine samples, but the accuracy of carbon dioxide production ( $\dot{V}CO_2$ ) determined with OA-ICOS has not been demonstrated. We compared simultaneous measurement  $\dot{V}CO_2$  obtained using whole-room indirect calorimetry (IC) with DLW-based measurements from IRMS and OA-ICOS. Seventeen subjects (10 female; 22 to 63 yr) were studied for 7 consecutive days in the IC. Subjects consumed a dose of 0.25 g  $H_2^{18}O$  (98% APE) and 0.14 g  $^2H_2O$  (99.8% APE) per kilogram of total body water, and urine samples were obtained on days 1 and 8 to measure average daily  $\dot{V}CO_2$  using OA-ICOS and IRMS.  $\dot{V}CO_2$  was calculated using both the plateau and intercept methods. There were no differences in  $\dot{V}CO_2$  measured by OA-ICOS or IRMS compared with IC when the plateau method was used. When the intercept method was used,  $\dot{V}CO_2$  using OA-ICOS did not differ from IC, but  $\dot{V}CO_2$  measured using IRMS was significantly lower than IC. Accuracy (~1–5%), precision (~8%), intraclass correlation coefficients ( $R = 0.87$ – $0.90$ ), and root mean squared error (30–40 liters/day) of  $\dot{V}CO_2$  measured by OA-ICOS and IRMS were similar. Both OA-ICOS and IRMS produced measurements of  $\dot{V}CO_2$  with comparable accuracy and precision compared with IC.

adult; deuterium; humans; oxygen isotope; respiratory gas exchange

## INTRODUCTION

The gold standard for measuring total daily energy expenditure (TDEE) in free-living individuals is the doubly labeled water (DLW) method, which is based on the principle that different elimination rates of isotopic labels of hydrogen and

oxygen provide a measurement of carbon dioxide production ( $\dot{V}CO_2$ ), subject to certain limiting assumptions (10, 19). TDEE measured using the DLW method has been shown to have an accuracy in humans of  $\pm 1$ – $5\%$  against whole-room indirect calorimetry (IC) (5, 8, 15, 17–19, 23). Although the number of DLW studies in humans has increased over time (~100 per year), widespread adoption of the DLW method in humans has been limited by the costs of the isotopic labels and challenges related to sample collection, preparation, and analysis using isotope ratio mass spectrometry (IRMS).

An alternative approach to IRMS for water isotope analysis is laser absorption spectroscopy. These instruments are less expensive than IRMS (~\$100,000 vs. \$250,000), do not require highly trained technicians for their operation (1), and provide simultaneous measurement of multiple isotopes with less tedious sample preparation (20). There are two commercially available forms of laser absorption spectroscopy for water isotope analysis, cavity ring-down spectroscopy (CRDS) and off-axis integrated cavity output spectroscopy (OA-ICOS). With CRDS, a laser pulse is trapped in a highly reflective optical cavity. The exponential decay of the light intensity is measured (“ring-down” time) and used to calculate the concentration of the absorbing substance in the gas mixture in the cavity. Although CRDS water isotope analyzers provide accurate and precise measurements of total body water ( $0.5 \pm 1\%$ ) and TDEE ( $0.5 \pm 6\%$ ) compared with IRMS, commercial CRDS analyzers have substantial instrumental memory effects, necessitating both careful considerations for reducing isotopic disparity between measured samples and mathematical correction (21). Furthermore, in the above-referenced study, CRDS was validated against IRMS but not against the criterion measurement of near-continuous respiratory gas exchange.

The other commercially available form of laser absorption spectroscopy for water isotopes, OA-ICOS, uses a laser light source that is coupled to an optical cavity in an off-axis fashion. The laser light wavelength is scanned over absorption features of interest, providing a direct measurement of the absorbing substances in the gas mixture (1). As with IRMS and CRDS, OA-ICOS also suffers from memory issues between adjacent samples. However, because the time to measure each

Address for reprint requests and other correspondence: L. Melanson, MS 8106, 12801 East 17th Ave., RC1 South RM 7103, University of Colorado Anschutz Medical Campus, Aurora, CO 80045 (e-mail: Ed.melanson@ucdenver.edu).

sample (100 s) with OA-ICOS is relatively short and requires only a small volume of sample per injection (~1,000 nL), memory issues can be circumvented using a higher number of injections per sample, negating the need to perform mathematical corrections. We (1–3) have previously shown this approach to be accurate and precise compared with IRMS for both measuring isotopic measurements of pure water and of human urine samples at both enriched and natural abundances. However, the accuracy and precision of measuring daily  $\dot{V}CO_2$  using the DLW method with samples measured using OA-ICOS by comparison to whole room indirect calorimetry has not yet been determined. Thus the purpose of this study was to compare measurement of daily  $\dot{V}CO_2$  in liters/day in a whole-room indirect calorimeter, with  $\dot{V}CO_2$  measured simultaneously using the DLW method and with the resultant body water samples (urine) analyzed using OA-ICOS. We also compared the accuracy and precision of OA-ICOS to those of IRMS.

## METHODS

**Institutional Approval and Ethics.** Procedures followed were in accordance with the ethical standards of the Helsinki Declaration of 1975 as revised in 1983. The study was approved by the Colorado Multiple Institutional Review Board on May 2, 2013. The study was registered on ClinicalTrials.gov (NCT01938794) on September 5, 2013. Subject recruitment and enrollment commenced in September, 2013, and the last study visit occurred in February, 2017.

**Subjects and screening procedures.** Adult volunteers ( $\geq 18$  yr) were recruited from the University of Colorado Anschutz Medical Campus (CU-AMC) and local communities. After participants provided informed, written consent, a Health History and Physical Examination was performed to confirm that volunteers were in a good state of health and that they met criteria for inclusion or exclusion. Primary study exclusion criteria were self-reported smoking or use of smokeless tobacco products, self-reported chronic disease (e.g., heart disease, diabetes, or thyroid disease), or current pregnancy. Body composition was then assessed using whole body dual-energy X-ray absorptiometry (DXA Hologic Delphi-W; Hologic, Bedford, MA). Because of weight limitations of the DXA, volunteers with a body weight  $> 135$  kg were also excluded.

**Experimental design and study procedures.** Subjects were studied for 1 wk in the whole-room indirect calorimeter located at the University of Colorado Anschutz Medical Campus. Upon subject arrival on *day 1*, body weight was measured to  $\pm 0.1$  kg, and a baseline urine sample was obtained for determination of background abundances of  $\delta^2H$  and  $\delta^{18}O$ . Subjects were then given an oral dose of 0.25 g of 98 atom percent (98% APE)  $^{18}O$ -labeled water and 0.14 g 99.8% APE  $^2H$ -labeled water (Sigma-Aldrich) per kilogram of total body water (estimated as 73% of FFM derived from DXA). The dosing cup was twice rinsed with 30 ml of tap water and consumed to ensure complete dosing. After the dose was provided, subjects entered the room calorimeter to begin the 7-day study. Subjects were instructed to completely void  $\sim 1$  h after the dose was delivered. Post-dosing urine samples were obtained 4 h (PD4) and 5 h (PD5) after the DLW dosing. On *days 2–7*, subjects exited the calorimeter for 1 h each day (0700–0800), during which time body weight was measured, and then subjects were permitted to shower. For the entire 7-day study, ad libitum meals were provided each day at 9 AM, 1 PM, and 6 PM. Subjects were instructed to perform exercise (30 min of treadmill walking at a brisk walking pace) each day to increase TDEE above sedentary levels. On *day 8*, subjects exited the calorimeter and end-dose urine and blood samples were obtained at the same time of day as on *day 1* (ED4 and ED5). Approximately 20 ml of each urine sample was immediately pipetted into airtight cryotubes and stored at

approximately  $-10^\circ C$  until transferred to a  $-80^\circ C$  freezer. Duplicate samples remained frozen at  $-80^\circ C$  until analysis.

**Whole-room-indirect calorimetry.** Average daily  $\dot{V}CO_2$  and 24-h energy expenditure (EE) over the 7-day period were measured using the whole-room indirect calorimeter located at CU-AMC using a previously described indirect calorimetry system (Sable Systems, International, Las Vegas, NV) (13).  $O_2$  consumption ( $\dot{V}O_2$ ) and  $\dot{V}CO_2$  were calculated in 1-min intervals using the flow rate, and the differences in  $CO_2$  and  $O_2$  concentrations between entering and exiting air, and minute-by-minute EE were calculated using the equations of Jequier et al. (7). Daily 24-h  $\dot{V}CO_2$  and EE were obtained by summing minute values over the 23-h measurement period and extrapolating to 24-h values. The accuracy and precision of the system were tested monthly using propane combustion tests. The average  $O_2$  and  $CO_2$  recoveries during the study were  $\geq 97.0\%$ . While this study was being performed, we also performed several tests using infusions of nitrogen and  $CO_2$  using high-precision mass flow controllers, and those tests yielded an accuracy of the IC within 1% of the expected values (unpublished observations).

**OA-ICOS analysis of urine samples.** Previously frozen urine samples were prepared by centrifugation, as previously described (3); no distillation or decolorizing steps were undertaken. The OA-ICOS instrument was calibrated using deionized working standards that had been previously calibrated by OA-ICOS against the VSMOW2 and SLAP2 international standards, as previously described (1, 3). Briefly, centrifuged urine samples were injected into a heated ( $\sim 85^\circ C$ ) stainless steel injection block to produce water vapor, which was then introduced into the OA-ICOS optical cavity. Simultaneous measurements of  $\delta^2H$  and  $\delta^{18}O$  were performed on each individual injection. Isotope range within each run was minimized by grouping samples expected to have similar enrichments (e.g., PD4/PD5, ED4/ED5) and by using working standards that closely bracketed the expected isotope ratios. Samples, working standards, and internal controls were interleaved throughout each analysis to ensure high accuracy by frequent intrarun calibration. For every individual measurement within a run, samples, working standards, and internal controls were injected 8–12 times depending on the total isotope range of the run (e.g., runs with high enriched samples were injected 12 times.). We have previously shown this approach to produce accurate and precise measurements without memory correction compared with IRMS (1, 3). Three to five urine samples were typically included in an individual run, which took  $\sim 5$ –7 h to complete. At the conclusion of each OA-ICOS run, the syringe, injector block, tubes, and filters were cleaned as previously described (1). Each sample was analyzed in a duplicate run on a subsequent day (typically within the same week). If the difference between duplicate runs exceeded 2 per mil ( $\delta\text{‰}$ ) for  $^2H: ^1H$  or 1  $\delta\text{‰}$  for  $^{18}O: ^{16}O$  for a given sample, then that sample was run again and only duplicate values that fell within this range were used.

Isotopic data from the OA-ICOS analyzer were processed using commercially available Post Analysis Software (LGR, version 3.1.0.9) as previously described (1, 2). Within each run, working standard measurements were utilized with a cubic spline standardization to calibrate urine sample measurements. Specifically, a cubic spline was fitted to all measurements of a single standard throughout the run. For each sample injection, an individual calibration curve was constructed from the splined values of each of the working standards. This approach maximally corrects for any instrument drift over the course of the run. To mitigate the effects of sample-to-sample memory on the OA-ICOS measurements, several procedures were employed (1, 3). First, to account for memory effects between successive samples, the last four injections for each sample were averaged, ignoring the first four to eight injections. Second, to monitor instrument performance, including memory effects between successive samples, an internal control water of known isotopic composition within the range of the isotope ratios of the working standards was measured periodically within each run. Internal controls were checked

against the known values. Runs where the internal controls differed from known values by more than  $\pm 1.0$ – $2.0$  ‰ (for low- and high-enriched samples) for  $\delta^2\text{H}$ , or  $\pm 0.3$  ‰ for  $\delta^{18}\text{O}$  from the known value were repeated. Precision of the urine samples was assessed using these same parameters. Finally, an injection volume (linearity) correction was employed to reduce the effects of different water concentrations (due to syringe volume fluctuations) on the measured isotope ratios. The postanalysis software also identified any individual injections that were outliers (isotope ratio  $\pm 3.0$  SD within an injection set) and for the presence of any organic contamination, using the integrated Spectral Contamination Identifier feature (9). The presence of any outliers also identified samples where memory effects had not been eliminated.

**IRMS analysis of urine samples.** Frozen urine samples were shipped from UC-AMC to Maastricht University in air-tight sealed glass vials and kept frozen using dry ice. Samples were transferred to a  $-80^\circ$  freezer and remained frozen until analyzed. For the analysis of  $^2\text{H}:^1\text{H}$ , a 2-ml glass vial containing 300  $\mu\text{l}$  of urine was filled with hydrogen gas, and equilibration occurred for 1 day at room temperature with a catalyst (5% platinum on alumina, 325 mesh; Aldrich Chemical) placed in an insert in the vial. For the analysis of  $^{18}\text{O}$ , 300  $\mu\text{l}$  of urine was put in a glass vial, which was then filled with  $\text{CO}_2$  gas. Equilibration then took place for 4 h at  $40^\circ\text{C}$ . The relative amounts of  $^2\text{H}:^1\text{H}$  in hydrogen gas and  $^{18}\text{O}:^{16}\text{O}$  in  $\text{CO}_2$  were then determined using IRMS (Micromass Optima Dual Inlet mass spectrometer with a Multiprep; Manchester, UK, 1998). Each run contained a total of 60 samples of which 12 were working standards with isotope concentrations that bracketed the expected isotope ratios of the urine samples. Each sample was analyzed in a duplicate run on a subsequent day (typically within the same week).

**Calculation of  $\dot{V}\text{CO}_2$  and TDEE.** For both OA-ICOS and IRMS, TBW was calculated as the average of the dilution spaces of  $^2\text{H}$  and  $^{18}\text{O}$  after correction for isotopic exchange with other body pools (14). Deuterium ( $k_{\text{D}}$ ) and oxygen ( $k_{\text{O}}$ ) turnover rates were calculated by linear regression of the natural logarithm of isotope enrichment as a function of time. All four time points were used in the calculation of  $k_{\text{D}}$  and  $k_{\text{O}}$ . TBW and  $\dot{V}\text{CO}_2$  were calculated using the plateau and intercept methods (using the average of the PD4 and PD5 enrichments) and Eq. A6 of Schoeller et al. (15):

$$\text{rCO}_2 \text{ (mol/day)} = (\text{N}/2.078) \times (1.01k_{\text{O}} - 1.041k_{\text{D}}) - 0.0246 \times \text{rGF}$$

where 1.01 and 1.04 represent the dilution spaces for deuterium and  $^{18}\text{O}$ , respectively, N is the body water dilution space, and rGF is the rate of gas fractionation estimated as  $1.05\text{N}(k_{\text{O}} - k_{\text{D}})$  (5). TDEE from OA-ICOS and IRMS was calculated using the calculated  $\dot{V}\text{CO}_2$  and the equation of Weir (TDEE =  $3.94 \times \dot{V}\text{O}_2 + 1.1 \dot{V}\text{CO}_2$ ), where  $\dot{V}\text{O}_2 = \dot{V}\text{CO}_2/\text{RQ}$  (22), assuming a respiratory quotient (RQ) of 0.86, and averaged over 7 days.

**Sample size justification.** Samples size estimates were based on repeated measures on 15 individuals studied in the room calorimeter located at the UC-AMC (unpublished data). The difference between the two 24-h  $\dot{V}\text{CO}_2$  measurements was  $\sim 12.7 \pm 7.5$  liters/day ( $\sim 3\%$  of mean values). A total sample of 16 paired measurements was estimated to achieve  $\sim 80\%$  power to detect equivalence in 24-h  $\dot{V}\text{CO}_2$  between IC and either IRMS or OA-ICOS when the margin of equivalence is  $\pm 7.7$  liters/day with a 0.05 significance level.

**Statistics.** Prior to analysis, all data were tested for normality. Differences between IC, OA-ICOS, and IRMS were determined using a repeated-measures ANOVA. Post hoc comparisons were performed using Tukey's multiple comparison test. Because our primary objective was to compare each instrument type to the criterion measure IC, we report only the comparisons between IC and OA-ICOS and IC and IRMS. Level of agreement was evaluated using the difference between the criterion and observed values (percent error, a measure of accuracy), the variance around the accuracy (a measure of precision), intraclass correlation coefficient (a measure of level of agreement), root mean squared error (rMSE, a measure of the magnitude of errors resulting from both bias and variability), and Bland-Altman plots (which provide a measure of bias and limits of agreement, as well as determining whether the error is associated with the magnitude of the criterion measure). The Bland-Altman analyses were performed using the IC as the criterion measure. Associations between subject characteristics and measurement error were determined using Pearson's correlation coefficient. Significance for all tests was set at  $P = 0.05$ . Analyses were performed using GraphPad Prism (v. 5.03, La Jolla, CA). Data are reported as means  $\pm$  SD.

## RESULTS

Nineteen subjects participated in the study. One subject withdrew after 1 day in the calorimeter. Due to technical issues, 2 days of data were lost on another subject, and that

Table 1. Subject characteristics and individual average total daily  $\dot{V}\text{CO}_2$  measured by IC and by OA-ICOS and IRMS using the plateau method

Subject	Sex	Age (yr)	Weight (kg)	BMI (kg/m <sup>2</sup> )	$\dot{V}\text{CO}_2$ (liters/day)		
					IC	OA-ICOS	IRMS
1	F	46	63.0	24.0	310.6	307.1	267.3
2	M	32	82.8	23.9	457.4	456.2	440.2
3	M	43	74.8	25.1	455.8	484.1	487.8
4	M	28	61.0	22.4	346.8	374.9	367.8
5	F	24	93.8	31.9	471.3	474.7	476.9
6	F	60	48.9	19.4	293.4	339.0	334.7
7	F	62	53.3	21.8	349.9	372.8	351.3
8	M	34	91.5	32.2	444.2	458.3	436.9
9	M	40	71.6	23.0	442.8	448.3	390.5
10	F	27	111.6	46.4	514.4	568.2	529.5
11	F	60	95.0	34.8	437.1	560.5	421.6
12	F	63	115.0	42.8	423.1	474.7	453.7
13	F	34	101.4	36.1	433.7	449.7	453.9
14	M	24	73.9	23.0	473.4	450.7	545.9
15	F	30	72.0	28.5	394.0	367.3	391.7
16	F	22	61.7	24.5	353.8	358.4	336.6
17	M	43	69.6	20.8	387.9	415.9	424.4
Mean (SD)		39 (14)	78.8 (19.7)	28.3 (7.9)	411.2 (62.1)	433.0 (72.7)	418.3 (73.0)

IC, indirect calorimetry; OA-ICOS, off-axis integrated cavity output spectroscopy; IRMS, isotope ratio mass spectrometry.

Table 2. TBW, FFM, FM, and %Fat measured by DXA, OA-ICOS, and IRMS. OA-ICOS and IRMS

	Intercept Method			Plateau Method	
	DXA	OA-ICOS	IRMS	OA-ICOS	IRMS
TBW, kg	38.3 (7.3)	38.3 (6.7)	35.6 (6.5) <sup>a</sup>	38.3 (6.7)	39.0 (6.7)
FFM, kg	52.5 (10.0)	52.2 (9.4)	48.8 (8.9) <sup>a</sup>	52.5 (10.0)	53.4 (9.2)
FM, kg	25.9 (15.8)	26.6 (15.8)	29.9 (16.0) <sup>a</sup>	26.3 (16.0)	25.3 (15.8)
%Fat	31.0 (12.5)	31.9 (11.9)	36.8 (11.3) <sup>a</sup>	31.5 (12.6)	30.2 (12.0)

Results are expressed as means (SD). TBW, total body water; FFM, fat-free mass; FM, fat mass; DXA, dual-energy X-ray absorptiometry. <sup>a</sup>Significantly different from DXA.

subject was excluded from the analysis. Thus, the final study sample consisted of 17 participants (Table 1).

Average daily turnover rates of deuterium ( $k_D$ /day) and oxygen ( $k_O$ /day) determined using OA-ICOS ( $0.118 \pm 0.031$ /day and  $0.142 \pm 0.034$ /day, respectively) were nearly identical to those determined using IRMS ( $0.118 \pm 0.032$ /day,  $0.141 \pm 0.033$ /day). The individual  $k_O$ ,  $k_D$ ,  $N_O$ , and  $N_D$  data used to perform these calculations is contained in the supplementary data file.

**Results using the plateau method.** TBW, fat-free mass (FFM), fat mass (FM), and body fat percentage (%Fat) measured by DXA, OA-ICOS, and IRMS are shown in Table 2. There were no differences in TBW, FFM, FM, or %Fat measured by OA-ICOS or IRMS when compared with DXA. Regardless of approach  $N_D$  and  $N_O$  were similar (Table 3), and the average dilution space ratios were close to the empirically derived value in adult humans of 1.031 (15).

There were no significant differences in average  $\dot{V}CO_2$  measured by OA-ICOS ( $433.0 \pm 72.7$  liters/day) or IRMS ( $418.3 \pm 73.0$  liters/day) compared with IC ( $411.2 \pm 62.1$  liters/day) (Fig. 1, Table 1). To demonstrate the effect on calculated TDEE, 24-h EE from IC (calculated using the measured RQ) was compared with TDEE calculated from OA-ICOS and IRMS using the assumed RQ of 0.86, as would be done in a standard DLW study. Mean TDEE measured by OA-ICOS ( $10.16 \pm 1.70$  MJ/day) and IRMS ( $9.91 \pm 1.70$  MJ/day) did not significantly differ from IC ( $9.88 \pm 1.56$  MJ/day).

The accuracy of  $\dot{V}CO_2$  measured by OA-ICOS (mean %error) and IRMS was 5.4 and 1.7%, respectively (Table 4). The accuracy of OA-ICOS was significantly different from zero (95% CI does not cross zero). However, the size of the 95% CIs around the percent error were similar for OA-ICOS (+1.1 to +9.6 liters/day) and IRMS (-2.5 to +5.8 liters/day), indicating a similar level of precision. The ICC between OA-ICOS and IC [0.87 (95% CI = 0.67 - 0.95)] was similar to the ICC between IRMS and IC [0.89 (0.72 - 0.96)]. The RMSE was

Table 3. Deuterium ( $N_D$ ) and oxygen ( $N_O$ ) dilution spaces and dilution space ratio ( $N_D:N_O$ ) measured by OA-ICOS and IRMS

	Intercept Method		Plateau Method	
	OA-ICOS	IRMS	OA-ICOS	IRMS
$N_D$ , kg	38.0 (6.7)	37.9 (6.9)	38.9 (6.8)	40.4 (6.7)
$N_O$ , kg	36.8 (6.6)	36.8 (6.7)	37.8 (6.6)	39.0 (6.8)
$N_D:N_O$	1.033 (0.005)	1.030 (0.006)	1.029 (0.0068)	1.037 (0.013)

Results are expressed as means (SD).

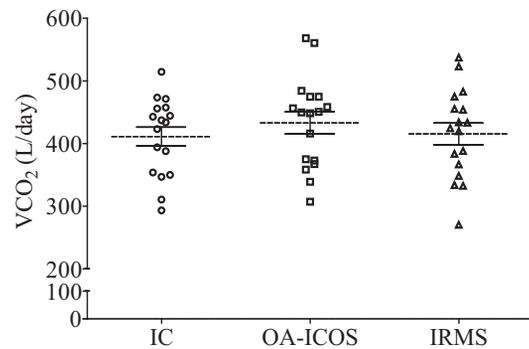


Fig. 1.  $\dot{V}CO_2$  (mean  $\pm$  SE) measured by indirect calorimetry (IC) and by off-axis integrated cavity output spectroscopy (OA-ICOS) and isotope ratio mass spectrometry (IRMS), using the plateau method.

40.2 liters/day for OA-ICOS and 31.5 liters/day for IRMS. Results of the Bland-Altman analysis are presented in Fig. 2. There was a significant bias for OA-ICOS (+21.8 liters/day, 95% CI = +3.9 to +39.8 liters/day) compared with IC, but not for IRMS (+7.1 liters/day, 95% CI = -9.1 to +23.4 liters/day). The reduced accuracy and significant bias for OA-ICOS was driven by a single outlier. The Bland-Altman correlations for OA-ICOS and IRMS were not significant, indicating no bias with absolute level of  $\dot{V}CO_2$ .  $\dot{V}CO_2$  for each individual measured by IC, OA-ICOS, and IRMS is shown in Table 1. For most individuals, all three methods produced similar results.

**Results using the intercept method.** When the intercept method was used, TBW and FFM estimated using IRMS were significantly lower, and FM and %fat significantly higher compared with DXA ( $P < 0.001$ ) (Table 2). There were no differences in TBW, FFM, FM, and %Fat measured by DXA compared with OA-ICOS.  $N_D$  and  $N_O$  were similar, and the average dilution space ratios were close to the theoretical value in adult humans of 1.031 (15) (Table 3). There was no difference in average  $\dot{V}CO_2$  measured by OA-ICOS ( $422.9 \pm 70.7$  liters/day) when compared with IC ( $411.2 \pm 62.1$  liters/day), but  $\dot{V}CO_2$  measured by IRMS ( $381.9 \pm 69.2$  liters/day) was significantly different compared with IC (Fig. 3). Similarly, mean TDEE measured by OA-ICOS ( $10.40 \pm 1.70$  MJ/day) was not different than 24 h EE. However, mean TDEE measured by IRMS using the intercept method ( $9.05 \pm 1.62$  MJ/day) was significantly lower than 24 h EE. Individual subject  $\dot{V}CO_2$  results calculated using the intercept method are presented in Supplemental Table S1 (all supplemental material for this article is accessible on the journal web site).

As with the plateau method, there was a similar level of agreement when  $\dot{V}CO_2$  measured using OA-ICOS and IRMS was compared with IC (Table 4). Interestingly, accuracy be-

Table 4. Limits of agreement for  $\dot{V}CO_2$  measured by OA-ICOS and IRMS

	Error (%) Mean (95% CI)	ICC (95% CI)	RMSE (liters/day)
OA-ICOS-plateau	5.4 (+1.1, +9.6)	0.87 (0.67, 0.95)	40.2
IRMS-plateau	1.7 (-2.5, +5.8)	0.89 (0.72, 0.96)	31.5
OA-ICOS-intercept	2.9 (-1.1, +6.9)	0.88 (0.70, 0.90)	33.8
IRMS-intercept	-7.2 (-11.2, -3.3)	0.90 (0.74, 0.96)	35.9

Results are presented for both plateau and intercept methods, ICC, interclass correlation; RMSE, root mean square error.

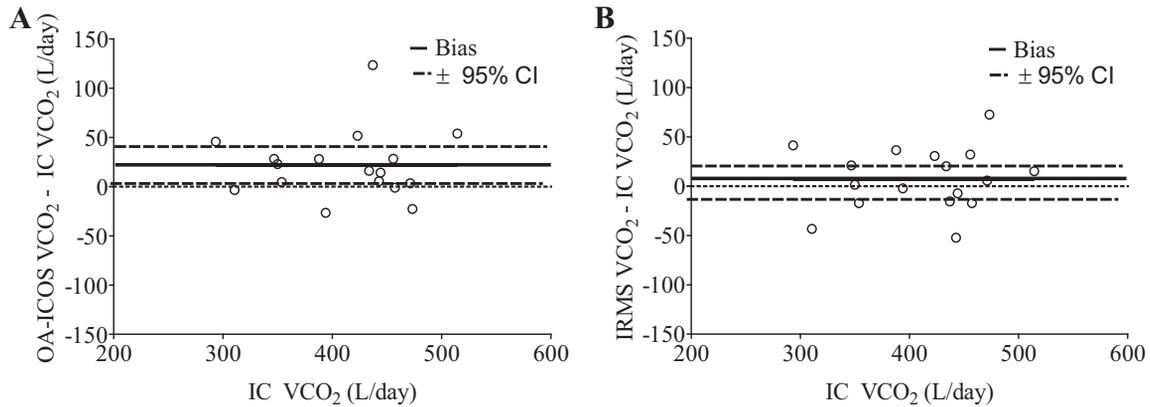


Fig. 2. Bland-Altman plots of OA-ICOS (A) and IRMS (B), using the plateau method vs. criterion measure IC.

tween OA-ICOS and IC tended to be better using the intercept method, whereas accuracy between IRMS and IC tended to be better using the plateau method. Precision, ICC, and RMSE were similar for OA-ICOS and IRMS using the intercept method. Results of the Bland-Altman analysis are presented in Fig. 4. There was a significant bias for IRMS ( $-29.2$  liters/day, 95% CI =  $-44.6$  to  $-13.9$  liters/day) compared with IC but not for OA-ICOS ( $+11.7$  liters/day, 95% CI =  $-5.1$  to  $+28.5$  liters/day). The Bland-Altman correlations between average  $\dot{V}_{CO_2}$  from IC and both IRMS and OA-ICOS were not significant indicating no bias with absolute level of EE.

**Additional analyses.** To determine whether %Fat, BMI, or age were contributing factors to differences between IC and IRMS or OA-ICOS, correlations between those variables and the differences in  $\dot{V}_{CO_2}$  between IC and OA-ICOS and IC and IRMS were determined (using the plateau data). The differences in  $\dot{V}_{CO_2}$  between IC and OA-ICOS were not significantly correlated with %Fat ( $r = 0.41$ ) or BMI ( $r = 0.42$ ) but were positively and significantly ( $P < 0.05$ ) associated with age ( $r = 0.59$ ). However, this significant correlation was driven solely by one subject (S12, a 60-yr-old female), where OA-ICOS substantially overestimated IC ( $+54$  liters/day). The differences between IC and IRMS were not significantly correlated with %Fat ( $r = -0.07$ ), BMI ( $r = -0.20$ ), or age ( $r = 0.04$ ). We also examined the association between the differences in  $\dot{V}_{CO_2}$  (IC - OA-ICOS, IC - IRMS) with measured RQ. The differences (TDEE - 24-h EE) between IC and OA-ICOS ( $r = 0.19$ ) and IRMS ( $r = 0.46$ ) were positively but weakly ( $P > 0.05$ ) correlated with average daily 24-h RQ. We

performed these same analyses using the intercept data, and results were similar (data not shown).

## DISCUSSION

Because of the high costs of operation and technical expertise required for operation of IRMS, only a few specialized laboratories are equipped to perform DLW measurements of TDEE. Although new approaches such as OA-ICOS are available, they have not yet been validated against room calorimetry. We compared  $\dot{V}_{CO_2}$  calculated using isotopic measurements obtained using OA-ICOS against 24-h  $\dot{V}_{CO_2}$  measured using whole-room indirect calorimetry as the criterion measurement. We also compared  $\dot{V}_{CO_2}$  calculated using isotopic measurements obtained using IRMS on the same samples to then evaluate whether the techniques provide comparable accuracy and precision compared with IC. Mean  $\dot{V}_{CO_2}$  measured using OA-ICOS did not differ significantly from IC, whether a plateau or intercept calculation approach was used. Mean  $\dot{V}_{CO_2}$  measured using IRMS did not differ from IC when the plateau method was used, but it was significantly lower than IC when the intercept method was used. Nonetheless, measurements of accuracy (%error), precision (SD of mean %error), ICC, RMSE, and Bland-Altman analyses suggested that level of agreement with IC was similar for both IRMS and OA-ICOS. Thus, results of this study demonstrate that OA-ICOS provides estimates of  $\dot{V}_{CO_2}$  from DLW studies in humans that are as accurate and precise as estimates derived from IRMS.

Initial validation work of the DLW method performed in the 1950s in several small animal species showed that  $\dot{V}_{CO_2}$  was within  $\sim 3\%$  of that measured simultaneously by indirect calorimetry (11, 12). Schoeller and van Santen (16) performed the first validation studies in humans in 1982 and reported that TDEE from the DLW method differed from measured energy intake (adjusted for changes in body composition) by an average of 2%. Subsequent validation studies against near-continuous respiratory gas exchange measured over 4–7 days reported precisions of  $\sim 1$ – $8\%$  for measuring  $\dot{V}_{CO_2}$  and TDEE (5, 8, 15, 17, 18, 23). The range of accuracies for both OA-ICOS and IRMS in the present study (Table 4), using both the plateau and intercept methods, were similar to those previous studies. Surprisingly, when using the intercept method, we observed a significant difference between mean  $\dot{V}_{CO_2}$  mea-

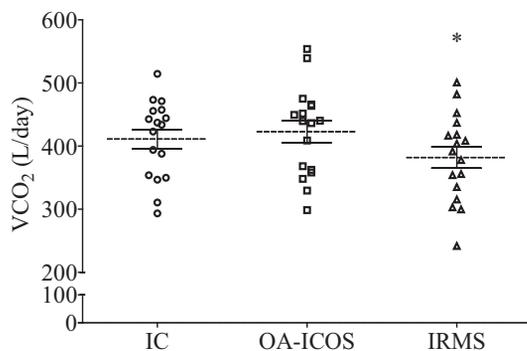


Fig. 3.  $\dot{V}_{CO_2}$  (mean  $\pm$  SE) measured by IC and by OA-ICOS and IRMS, using the intercept method. \*Significantly different from IC.

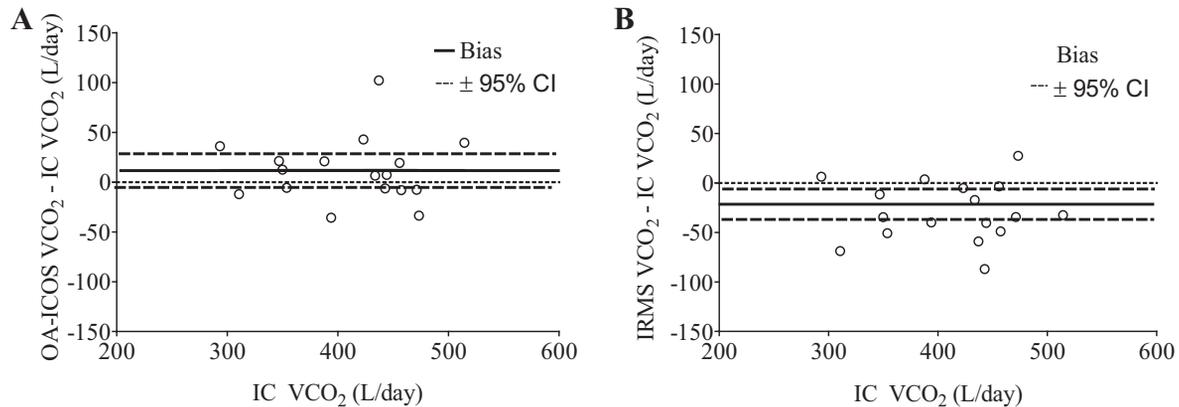


Fig. 4. Bland-Altman plots of OA-ICOS (A) and IRMS (B), using the intercept method vs. criterion measure IC.

sured by IC and IRMS, which is not consistent with previous validation studies.

To more thoroughly compare the IC to OA-ICOS (and IC to IRMS), we performed several statistical tests to assess the levels of agreement between instruments, some of which are more reflective of individual errors. Specifically, both the ICC and RMSE describe how concentrated the data are around the line of best fit (in this case, the line of identity), whereas the Bland-Altman allows identification of systematic differences between two measurements (4). Both the RMSE and Bland-Altman can be also used to identify where measurement errors are driven by the presence of outliers. Because DLW studies are performed on groups of individuals (e.g., to compare differences between groups to determine the effect of some intervention), more weight should be given to tests that are based on mean differences. For example, even though the Bland-Altman test indicated a significant, positive bias in measuring  $\dot{V}CO_2$  using the plateau method with OA-ICOS (+21.8 liters/day), there was no difference in mean  $\dot{V}CO_2$  measured by OA-ICOS and IC. On the basis of the current analyses, we conclude that OA-ICOS provides a measurement of average daily  $\dot{V}CO_2$  that is accurate (1–5%) and precise (8%) without systematic bias. We also conclude that accuracy, precision, and bias are similar to those observed with IRMS.

It has been suggested that adiposity and nutritional status affect the dilution space ratio ( $N_d/N_o$ ) between  $^2H$  and  $^{18}O$ , causing potential errors in  $\dot{V}CO_2$  when the DLW method is used (6). In that study, it was reported that there was an overestimation of  $\dot{V}CO_2$  by the DLW method in high-fat (HF) diet-fed mice compared with measured  $\dot{V}CO_2$  using continuous measurements with IC. This overestimation occurred in both diet-induced obesity-prone (DIO) and diet-induced obesity-resistant (DR) groups, suggesting that the overestimation is independent of body fat gain during a HF diet. In the present study, we found no association between either %Fat or BMI and the difference in  $\dot{V}CO_2$  measured with IC and DLW. We also explored the association between measured RQ and the difference in  $\dot{V}CO_2$  measured with IC and DLW. These associations were also nonsignificant with both OA-ICOS and IRMS. Although we did not measure energy intake (subjects consumed an ad libitum diet), our subjects were weight stable throughout the 7-day study ( $-0.5 \pm 0.8$  kg, mean  $\pm$  SD), suggesting that individual differences in av-

erage 24-h RQ reflected differences in habitual energy macronutrient intake rather than energy balance. Under this assumption, if  $\dot{V}CO_2$  is overestimated during consumption of a HF diet, a negative correlation would be expected when the differences between the DLW and IC  $\dot{V}CO_2$  are plotted against RQ (with a lower RQ indicative of a higher fat intake). Thus, results of the present study do not support the conclusion that  $\dot{V}CO_2$  from the DLW method is overestimated during a HF diet, but we concede that this can be determined only during studies in which energy and macronutrient intake are highly controlled.

**Strengths and limitations.** One strength of the current study is the sample size, which is larger ( $n = 17$ ) than previous validation studies performed using near-continuous measurements of respiratory gas exchange ( $n < 10$ ) (5, 8, 15, 17, 18, 24). A limitation of the current study, as in all validation studies, is the validity of the criterion measure (IC). However, as described in METHODS, the room calorimeter system at UC-AMC consistently measures within 1–3% of expected values using gas infusion and propane combustion tests. In addition to costs, OA-ICOS offers several advantages over IRMS, including easier sample preparation and reducing the need for highly trained technicians. However, it should be noted that the sample measurement configuration used in the present study (e.g., 8–12 injections per sample, with multiple interleaved measurements of working standards and internal controls) does not increase the throughput compared with IRMS and CRDS. The advantage of this approach is that it negates the need for mathematical correction due to memory effects. Throughput could be increased by reducing the number of injections per sample, but the tradeoff would then be the need to apply mathematical correction for memory effects.

In conclusion, mean  $\dot{V}CO_2$  measured using OA-ICOS did not differ significantly from concurrently measured 24-h  $\dot{V}CO_2$  using whole-room indirect calorimetry, whether using the plateau or the intercept calculation approach. Furthermore, both OA-ICOS and IRMS produced measurements of  $\dot{V}CO_2$  with comparable accuracy and precision compared with whole-room indirect calorimetry. On the basis of these results, we conclude that off-axis integrated cavity output spectroscopy provides a valid and viable alternative to IRMS for measuring TDEE using DLW in humans.

## ACKNOWLEDGMENTS

This work was supported with resources and use of facilities from the Geriatric Research, Education, and Clinical Center at the Denver Veterans Affairs Medical Center.

## GRANTS

This work was supported by an NIH Small Business Innovation (SBIR) research grant (R44 DK-093362), as well as support from the Colorado Nutrition and Obesity Research Center (P30 DK-048520) and the Colorado Clinical and Translational Science Institute (UL1 RR-025780). E. L. Melanson is also supported by resources from the Geriatric Research, Education, and Clinical Center at the Denver Veterans Affairs Medical Center Clinical Trial Registry: The study was registered on ClinicalTrials.gov (NCT01938794).

## DISCLAIMERS

The contents do not represent the views of the US Department of Veterans Affairs or the US Government.

## DISCLOSURES

E. Berman is employed by ABB/Los Gatos Research, the company that manufactures the OA-ICOS analyzer.

## AUTHOR CONTRIBUTIONS

E.L.M., J.R.S., and E.S.B. conceived and designed research; E.L.M., T.S., V.A.C., S.A.C., G.P., L.W., and E.S.B. performed experiments; E.L.M., G.P., L.W., J.R.S., and E.S.B. analyzed data; E.L.M., W.M.K., G.P., J.R.S., and E.S.B. interpreted results of experiments; E.L.M. prepared figures; E.L.M. drafted manuscript; E.L.M., T.S., W.M.K., V.A.C., S.A.C., G.P., L.W., J.R.S., and E.S.B. edited and revised manuscript; E.L.M., T.S., W.M.K., V.A.C., S.A.C., G.P., L.W., J.R.S., and E.S.B. approved final version of manuscript.

## REFERENCES

- Berman ES, Fortson SL, Snaith SP, Gupta M, Baer DS, Chery I, Blanc S, Melanson EL, Thomson PJ, Speakman JR. Direct analysis of  $\delta^2\text{H}$  and  $\delta^{18}\text{O}$  in natural and enriched human urine using laser-based, off-axis integrated cavity output spectroscopy. *Anal Chem* 84: 9768–9773, 2012. doi:10.1021/ac3016642.
- Berman ES, Levin NE, Landais A, Li S, Owano T. Measurement of  $\delta^{18}\text{O}$ ,  $\delta^{17}\text{O}$ , and  $^{17}\text{O}$ -excess in water by off-axis integrated cavity output spectroscopy and isotope ratio mass spectrometry. *Anal Chem* 85: 10392–10398, 2013. doi:10.1021/ac402366t.
- Berman ES, Melanson EL, Swibas T, Snaith SP, Speakman JR. Inter- and intraindividual correlations of background abundances of ( $^2\text{H}$ ), ( $^{18}\text{O}$ ) and ( $^{17}\text{O}$ ) in human urine and implications for DLW measurements. *Eur J Clin Nutr* 69: 1091–1098, 2015. doi:10.1038/ejcn.2015.10.
- Bland JM, Altman DG. Statistical methods for assessing agreement between two methods of clinical measurement. *Lancet* 327: 307–310, 1986. doi:10.1016/S0140-6736(86)90837-8.
- Coward WA, Prentice AM. Isotope method for the measurement of carbon dioxide production rate in man. *Am J Clin Nutr* 41: 659–663, 1985.
- Guidotti S, Meijer HA, van Dijk G. Validity of the doubly labeled water method for estimating  $\text{CO}_2$  production in mice under different nutritional conditions. *Am J Physiol Endocrinol Metab* 305: E317–E324, 2013. doi:10.1152/ajpendo.00192.2013.
- Jéquier E, Acheson K, Schutz Y. Assessment of energy expenditure and fuel utilization in man. *Annu Rev Nutr* 7: 187–208, 1987. doi:10.1146/annurev.nu.07.070187.001155.
- Klein PD, James WP, Wong WW, Irving CS, Murgatroyd PR, Cabrera M, Dallosso HM, Klein ER, Nichols BL. Calorimetric validation of the doubly-labelled water method for determination of energy expenditure in man. *Hum Nutr Clin Nutr* 38: 95–106, 1984.
- Brian Leen J, Berman ESF, Liebson L, Gupta M. Spectral contaminant identifier for off-axis integrated cavity output spectroscopy measurements of liquid water isotopes. *Rev Sci Instrum* 83: 044305, 2012. doi:10.1063/1.4704843.
- Lifson N, McClintock R. Theory of use of the turnover rates of body water for measuring energy and material balance. *J Theor Biol* 12: 46–74, 1966. doi:10.1016/0022-5193(66)90185-8.
- Lifson N, Gordon GB, McClintock R. Measurement of total carbon dioxide production by means of  $\text{D}_2\text{O}$ . *J Appl Physiol* 7: 704–710, 1955.
- McClintock R, Lifson N. Measurement of basal and total metabolism in hereditarily obese-hyperglycemic mice. *Am J Physiol* 193: 495–498, 1958.
- Melanson EL, Ingebrigtsen JP, Bergouignan A, Ohkawara K, Kohrt WM, Lighton JR. A new approach for flow-through respirometry measurements in humans. *Am J Physiol Regul Integr Comp Physiol* 298: R1571–R1579, 2010. doi:10.1152/ajpregu.00055.2010.
- Racette SB, Schoeller DA, Luke AH, Shay K, Hnilicka J, Kushner RF. Relative dilution spaces of  $^2\text{H}$ - and  $^{18}\text{O}$ -labeled water in humans. *Am J Physiol* 267: E585–E590, 1994.
- Schoeller DA, Ravussin E, Schutz Y, Acheson KJ, Baertschi P, Jéquier E. Energy expenditure by doubly labeled water: validation in humans and proposed calculation. *Am J Physiol* 250: R823–R830, 1986.
- Schoeller DA, van Santen E. Measurement of energy expenditure in humans by doubly labeled water method. *J Appl Physiol Respir Environ Exerc Physiol* 53: 955–959, 1982.
- Schoeller DA, Webb P. Five-day comparison of the doubly labeled water method with respiratory gas exchange. *Am J Clin Nutr* 40: 153–158, 1984.
- Seale JL, Conway JM, Canary JJ. Seven-day validation of doubly labeled water method using indirect room calorimetry. *J Appl Physiol* (1985) 74: 402–409, 1993.
- Speakman JR. *Doubly Labelled Water: Theory and Practice*. London: Chapman Press, 1997.
- Steig EJ, Gkinis V, Schauer AJ, Schoenemann SW, Samek K, Hoffnagle J, Dennis KJ, Tan SM. Calibrated high-precision  $\text{O}-17$ -excess measurements using cavity ring-down spectroscopy with laser-current-tuned cavity resonance. *Atmos Meas Tech* 7: 2421–2435, 2014. doi:10.5194/amt-7-2421-2014.
- Thorsen T, Shriver T, Racine N, Richman BA, Schoeller DA. Doubly labeled water analysis using cavity ring-down spectroscopy. *Rapid Commun Mass Spectrom* 25: 3–8, 2011. doi:10.1002/rcm.4795.
- Weir JB. New methods for calculating metabolic rate with special reference to protein metabolism. 1949. *Nutrition* 6: 213–221, 1990.
- Westerterp KR, Brouns F, Saris WH, ten Hoor F. Comparison of doubly labeled water with respirometry at low- and high-activity levels. *J Appl Physiol* (1985) 65: 53–56, 1988.
- Westerterp KR, Lefebvre HN, Sulkers EJ, Sauer PJ. Comparison of short term indirect calorimetry and doubly labeled water method for the assessment of energy expenditure in preterm infants. *Biol Neonate* 60: 75–82, 1991. doi:10.1159/000243391.