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Glucose kinetics during prolonged exercise in highly trained human subjects: effect of glucose ingestion


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1. The objectives of this study were (1) to investigate whether glucose ingestion during prolonged exercise reduces whole body muscle glycogen oxidation, (2) to determine the extent to which glucose disappearing from the plasma is oxidized during exercise with and without carbohydrate ingestion and (3) to obtain an estimate of gluconeogenesis.

2. After an overnight fast, six well-trained cyclists exercised on three occasions for 120 min on a bicycle ergometer at 50 % maximum velocity of $O_2$ uptake and ingested either water (Fast), or a 4 % glucose solution (Lo-Glu) or a 22 % glucose solution (Hi-Glu) during exercise.

3. Dual tracer infusion of [U-13C]-glucose and [6,6-H2]-glucose was given to measure the rate of appearance ($R_a$) of glucose, muscle glycogen oxidation, glucose carbon recycling, metabolic clearance rate ($MCR$) and non-oxidative disposal of glucose.

4. Glucose ingestion markedly increased total $R_a$ especially with Hi-Glu. After 120 min $R_a$ and rate of disappearance ($R_d$) of glucose were 51—52 μmol kg$^{-1}$ min$^{-1}$ during Fast, 73—74 μmol kg$^{-1}$ min$^{-1}$ during Lo-Glu and 117—119 μmol kg$^{-1}$ min$^{-1}$ during Hi-Glu. The percentage of $R_d$ oxidized was between 96 and 100 % in all trials.

5. Glycogen oxidation during exercise was not reduced by glucose ingestion. The vast majority of glucose disappearing from the plasma was oxidized and MCR increased markedly with glucose ingestion. Glucose carbon recycling was minimal suggesting that gluconeogenesis in these conditions is negligible.
we compared the rate of appearance of glucose ($R_a$) measured with a [6,6-$^3$H$_2$]-glucose tracer (GC-MS) and a [U-$^{13}$C]-glucose tracer (GC-IRMS) during exercise. With glucose ingestion, gluconeogenesis may be suppressed and we hypothesized that glucose carbon recycling would decrease with glucose ingestion.

In summary, the primary purpose of the present study was to investigate whether carbohydrate ingestion can reduce muscle glycogen oxidation measured at whole body level with the indirect stable isotope tracer method. The secondary purpose was to determine the percentage of glucose disappearing from the plasma that is oxidized during exercise and to quantify non-oxidative glucose disposal with or without carbohydrate ingestion. A third purpose was to investigate glucose carbon recycling as a measure of gluconeogenesis and to see whether glucose ingestion could suppress gluconeogenesis.

**METHODS**

**Subjects**

Six highly trained cyclists participated in this study. Their mean age and weight were 24 ± 2 years and 68 ± 1 kg. After explaining the nature and the risks of the experimental procedures to the subjects, their informed written consent was obtained. The study was approved by the local Medical Ethical Committee.

**Pre-testing**

Subjects' $V_{O_{2,max}}$ was measured on an electronically braked cycle ergometer (Lode Excalibur Sport, Groningen, The Netherlands) during an incremental exhaustive exercise test (Kuipers et al. 1995, 1996) one week before the first experimental trial and was found to be 76 ± 1 ml kg$^{-1}$ min$^{-1}$. Maximal work rate during the incremental exercise test was 397 ± 10 W. The results of this test were used to determine the 50% $V_{O_{2,max}}$ workload which was later employed in the experimental trials.

**Experimental trials**

Each subject performed six exercise trials, each separated by at least seven days. The order of the trials was determined by a Latin square design. Each trial consisted of 120 min cycling at ~50% $V_{O_{2,max}}$. Subjects ingested drinks containing no glucose (Fast), a 4.4% glucose solution (Lo-Glu) or a 22.0% (Hi-Glu) glucose solution. Each trial with one of these drinks was repeated, once with and once without infusion in order to allow correction for changes in breath $^{13}$CO$_2$ background enrichment during exercise.

**Protocol**

Subjects reported to the laboratory at 08.00 h after an overnight fast. A Teflon catheter (Baxter Quick Cath Dupont, Uden, The Netherlands) was inserted into an antecubital vein of one arm and at 08.30 h a resting blood sample of 10 ml was drawn. In the contralateral antecubital vein a second Teflon catheter was inserted for isotope infusion. Resting breath gases were collected over 5 min periods (Oxycon $\beta$, Minhardt, Mannheim, Germany) and vacutainer tubes were filled directly from the mixing chamber to determine the $^{13}$C/$^{12}$C ratio in expired CO$_2$. In all figures, time point 0 min refers to this resting sample. At 08.55 h subjects started a warming-up of 5 min at 100 W. Also, at 08.55 h a sodium bicarbonate prime was given (5.5 µmol kg$^{-1}$ Na $^{13}$CO$_3$; Cambridge Isotope Laboratories, USA) followed by a [6,6-$^3$H$_2$]-glucose (Cambridge Isotope Laboratories, USA) and a [U-$^{13}$C]-glucose (Cambridge Isotope Laboratories, USA) prime (dose equal to one hour infusion). Thereafter a continuous infusion of sterile pyrogen free [6,6-$^3$H$_2$]-glucose and [U-$^{13}$C]-glucose was started via a calibrated IVAC 560 pump (San Diego, CA). The concentration of isotope in the infusate was measured for each experiment so that the exact infusion rate could be determined. Infusion rates were 0.234 ± 0.01 and 0.032 ± 0.002 µmol kg$^{-1}$ min$^{-1}$ for [6,6-$^3$H$_2$]-glucose and [U-$^{13}$C]-glucose, respectively.

At 09.00 h the workload was increased to 50% $V_{O_{2,max}}$ for 120 min. During the first minute subjects drank an initial bolus (8 ml kg$^{-1}$) of one of the glucose solutions (24 g glucose during Lo-Glu and 120 g glucose during Hi-Glu). Thereafter every 15 min a beverage volume of 2 ml kg$^{-1}$ (6 g glucose during Lo-Glu; 30 g glucose during Hi-Glu) was provided. This feeding schedule has been shown to result in high rates of gastric emptying (Rehner et al. 1990). The mean amount of glucose provided during the 120 min of exercise was 72 ± 1 g in the Lo-Glu trial and 360 ± 7 g in the Hi-Glu trial. Blood samples were drawn at 15 min intervals until the end of exercise. Expiratory gases were collected every 15 min.

**Glucose solutions**

Subjects ingested glucose solutions prepared from potato-derived glucose (AWEBE, The Netherlands) containing either 44 or 220 g kg$^{-1}$ glucose. This glucose had a $^{14}$C-enrichment of −261 per thousand (‰ per mill) versus PDB (the international standard, Pee Dee Bellemnite; determined by on-line combustion-IRMS, Carlo Erba-Finnigan MAT 252, Bremen, Germany) which is similar to the $^{14}$C-enrichment of expired air of Europeans (Wagenmakers et al. 1993). To minimize shifts in background enrichment as a result of changes in endogenous substrate utilization, standard procedures were followed (Jeukendrup et al. 1995, 1996b). Subjects were instructed not to consume any products with a high natural abundance of $^{13}$C during the entire experimental period. Furthermore, subjects were instructed to keep their diet as constant as possible during the days before the trials.

**Analysis**

Blood (10 ml) was collected into tubes containing EDTA and was centrifuged for 4 min at 4 °C. Aliquots of plasma were frozen immediately in liquid nitrogen and stored at −40 °C until analysis of glucose (Roche, Uni Kit III, 0710670), lactate (Gutmann & Wahlefeld, 1974) and free fatty acids (FFA; Wako NEFA-C test kit, Wako Chemicals, Neuse, Germany) which were performed with the COBAS BIO semi automatic analyser. Insulin was analysed by radioimmuno assay (Nuchlab Ultra-sensitive Human Insulin RIA kit).

Breath samples were analysed for $^{13}$C/$^{12}$C ratio by gas chromatography continuous flow isotope ratio mass spectrometry (GC-IRMS; Finnigan MAT 252, Bremen, Germany).

For determination of $^{13}$C/$^{12}$C ratio of plasma glucose, glucose was derivatized to its trimethyloxil derivative using TriSil Z® (Pierce). Thereafter the derivative was measured by GC-IRMS (Finnigan MAT 252, Bremen, FRG). By establishing the relationship between the enrichment of a series of glucose standards of variable enrichment (by combustion-IRMS; Carlo Erba-Finnigan MAT 252, Bremen, Germany) and the enrichment of the glucose trimethyloxil derivative of these standards, the enrichment of plasma glucose sample was determined.

Plasma [H$^3$]glucose enrichment was determined by gas chromatography-mass spectrometry (GC-MS) analysis of the glucose
penta-acetate derivatives on a Finnigan INCOS-XL (Bremen, Germany). For \(^{[3]H}\)-glucose enrichment, ion mass of 200 and 202 were selectively monitored.

From indirect calorimetry (respiratory exchange ratio (RER), rate of oxygen consumption (\(V_{\text{O}}\)) and stable isotope measurements (\(^{13}\text{C}^{16}\text{O}_{2} / ^{13}\text{C}^{18}\text{O}_{2}\)) (IRMS, Finnigan MAT 252), total energy expenditure and oxidation rates of total fat, total carbohydrates and exogenous glucose were calculated.

**Calculations**

From the volume of CO\(_2\) production per unit time, (l min\(^{-1}\), \(V_{\text{CO}_2}\)) and \(V_{\text{O}_2}\), total carbohydrate and fat oxidation rates (g min\(^{-1}\)) were calculated using the non-protein respiratory quotient (Pieronnet & Maccio, 1991).

\[
\text{Glucose oxidation} = 4.585 V_{\text{CO}_2} - 3.226 V_{\text{O}_2}
\]

\[
\text{Fat oxidation} = 1.695 V_{\text{CO}_2} - 1.701 V_{\text{O}_2}
\]

The isotopic enrichment was expressed as a mil difference between the \(^{13}C/^{12}C\) ratio of the sample and a known laboratory reference standard according to the formula of Craig (1957):

\[
\delta^{13}C = \left( \frac{^{13}C/^{12}C_{\text{sample}}}{^{13}C/^{12}C_{\text{standard}}} - 1 \right) \times 10^3 \text{ per mil.}
\]

The \(\delta^{13}C\) will then be related to an international standard Pee Dee Bellemnitella (PDDB).

The total rate of appearance (i.e. total \(R_a\)) and rates of disappearance (i.e. total \(R_d\)) of glucose were calculated using the single-pool non-steady state equations of Steele (1959) adapted for use with stable isotope as described elsewhere (Wolfe, 1992). Total \(R_a\) represents the splanchic \(R_a\) from ingested CHO, hepatic glycogenolysis and gluconeogenesis.

\[
\text{Total } R_a = \frac{F - V(C_1 + C_2/2)[E_2 - E_1(V_2 - V_1)/2]}{(E_2 + E_1)/2},
\]

\[
\text{total } R_d = R_a V \left( \frac{C_2 - C_1}{V_2 - V_1} \right),
\]

where \(F\) is the infusion rate (\(\mu\)mol kg\(^{-1}\) min\(^{-1}\)), \(V\) is the volume of distribution (= 160 ml kg\(^{-1}\)), \(C_1\) and \(C_2\) are the glucose concentrations at times 2 and 1 (i.e. \(V_2, V_1\)), respectively, and \(E_1, E_2\) are the plasma glucose enrichments at times 2 and 1 (i.e. \(t_2, t_1\)), respectively.

The \(V_{\text{CO}_2}\) production from the tracer infusion was calculated as:

\[
V_{\text{CO}_2}\ (\mu\)mol kg\(^{-1}\) min\(^{-1}\)) = (E_{13} - E_{12}) V_{\text{CO}_2},
\]

where \(E_{13}\) is the breath \(^{13}C/^{12}C\) ratio at a given time and \(E_{12}\) is the breath \(^{13}C/^{12}C\) ratio at the same time during the trial with no tracer infusion. The \(C\) will then be related to an international standard Pee Dee Bellemnitella (PDDB).

The percentage of the infused \([U-^{13}\text{C}]\) tracer oxidized was calculated as:

\[
\% \text{ infused tracer oxidized} = \frac{(V_{\text{CO}_2} E_{13})}{[U-^{13}\text{C}]-\text{tracer infusion rate}} \times 100.
\]

**RESULTS**

**Plasma glucose, insulin, lactate, free fatty acids and glycerol**

In the Fast trial, plasma glucose concentrations were in the range of 4·2–4·6 mM at rest and throughout the exercise bout (Fig. 1). With glucose ingestion in the Hi-Glu and Lo-Glu trial plasma glucose concentrations peaked at 10–20 min at values of 5·5–5·8 mM. Plasma glucose concentrations were higher throughout exercise with glucose ingestion compared with Fast, although only Hi-Glu versus Fast reached statistical significance at all time points \((P < 0·05)\).

Plasma insulin was low at rest and during exercise when fasted (i.e. 5–7 \(\mu\)U ml\(^{-1}\); Fig. 1), but was significantly elevated by glucose ingestion during exercise. The highest insulin values were observed after 30 min (9 ± 2 \(\mu\)U ml\(^{-1}\) with Lo-Glu and 17 ± 4 \(\mu\)U ml\(^{-1}\) with Hi-Glu). Plasma insulin decreased in all trials but remained elevated in the Glu trials in comparison with Fast. In all conditions plasma insulin decreased to, or below, the resting fasting level at the end of the trials.

Plasma lactate at rest was 0·9 ± 0·1, 0·9 ± 0·1 and 0·8 ± 0·2 mM for Fast, Lo-Glu and Hi-Glu, respectively, and did not change during exercise (data not shown in Fig. 1). After 120 min of exercise the values were 0·9 ± 0·1, 0·8 ± 0·1 and 0·8 ± 0·1 mM, respectively.

During the Fast trial, plasma free fatty acid (FFA) concentration initially decreased during the first 10 min and thereafter gradually increased during exercise to about three times basal level (914 ± 99 \(\mu\)mol at 120 min; Fig. 1). Plasma FFA in both Glu conditions followed a similar
Plasma glycerol concentrations were increased during exercise from resting values in the range of 55–97 μM to 406 ± 39 μM at 120 min during Fast, 238 ± 35 μM during Lo-Glu and 152 ± 38 μM during Hi-Glu (Fig. 1; P < 0.05).

Whole body fat and carbohydrate oxidation

VO₂ was similar during each exercise session (38–39 ml kg⁻¹ min⁻¹) and elicited approximately 51 ± 2 % VO₂ max (Table 1). RER decreased in the Fast trial (P < 0.05) and remained stable with Hi-Glu (Fig. 2). After 120 min of exercise, total CHO oxidation rates were 130 ± 11, 154 ± 10 and 179 ± 6 μmol kg⁻¹ min⁻¹ for Fast, Lo-Glu and Hi-Glu, respectively (Fig. 2). Total fat oxidation was markedly suppressed by the Glu feedings (Fig. 2; P < 0.05). After 120 min of exercise total fat oxidation rates were 40 ± 2, 33 ± 3 and 26 ± 2 μmol kg⁻¹ min⁻¹ for Fast, Lo-Glu and Hi-Glu, respectively (P < 0.05).

Breath ratio of ¹³CO₂/¹²CO₂

In the exercise trials without tracer infusion, used to measure shifts in background or endogenous ¹³CO₂ production, the subjects displayed only a very slight non-significant elevation in ¹³CO₂/¹²CO₂ breath ratio both when fasted and when receiving glucose (Fig. 3). The glucose ingestion and oxidation did not alter ¹³CO₂/¹²CO₂ breath ratio during the background trial (i.e. without tracer infusion). The breath ratios during

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**Table 1. Whole body oxygen consumption (VO₂), respiratory exchange ratio, total carbohydrate oxidation, VCO₂ production and oxidation rates of the intravenously infused [U-¹³C]-glucose tracer during the 90–120 min period of exercise**

<table>
<thead>
<tr>
<th>Trial</th>
<th>Time (min)</th>
<th>VO₂ (ml kg⁻¹ min⁻¹)</th>
<th>Respiratory exchange ratio</th>
<th>Carbohydrate oxidation (μmol kg⁻¹ min⁻¹)</th>
<th>VCO₂ production (μmol kg⁻¹ min⁻¹)</th>
<th>Percentage of infused tracer oxidized</th>
</tr>
</thead>
<tbody>
<tr>
<td>Fast</td>
<td>90–105</td>
<td>38.9 ± 0.7</td>
<td>0.83 ± 0.01</td>
<td>130 ± 13</td>
<td>0.209 ± 0.024</td>
<td>98.6 ± 3.4</td>
</tr>
<tr>
<td></td>
<td>105–120</td>
<td>39.3 ± 0.7</td>
<td>0.83 ± 0.01</td>
<td>130 ± 11</td>
<td>0.210 ± 0.026</td>
<td>98.6 ± 3.4</td>
</tr>
<tr>
<td>Lo-Glu</td>
<td>90–105</td>
<td>38.2 ± 0.3</td>
<td>0.86 ± 0.01 *</td>
<td>156 ± 10 *</td>
<td>0.194 ± 0.007</td>
<td>98.7 ± 1.4</td>
</tr>
<tr>
<td></td>
<td>105–120</td>
<td>39.5 ± 0.3</td>
<td>0.85 ± 0.01 *</td>
<td>155 ± 10 *</td>
<td>0.192 ± 0.006</td>
<td>97.9 ± 1.0</td>
</tr>
<tr>
<td>Hi-Glu</td>
<td>90–105</td>
<td>37.7 ± 0.0</td>
<td>0.89 ± 0.01 *</td>
<td>182 ± 5 *</td>
<td>0.187 ± 0.003</td>
<td>98.3 ± 1.0</td>
</tr>
<tr>
<td></td>
<td>105–120</td>
<td>37.9 ± 0.0</td>
<td>0.89 ± 0.01 *</td>
<td>179 ± 6 *</td>
<td>0.183 ± 0.004</td>
<td>96.1 ± 1.3</td>
</tr>
</tbody>
</table>

Values are means ± s.e.m. VCO₂ production is the product of breath ¹³CO₂ enrichment above baseline and VCO₂ production. Percentage of infused tracer oxidized is calculated as (VCO₂ production/intravenous tracer infusion rate). * Glu trials significantly different from Fast at that time; P < 0.05. † Hi-Glu trial significantly different from the Lo-Glu trial at that time; P < 0.05.
the experimental trials with \([U-^{13}C]\)-glucose tracer infusion are shown in Fig. 3. The NaH\(^{13}CO_3\) prime, given just prior to beginning exercise during each trial, as expected, markedly raised breath \(^{13}CO_2/^{12}CO_2\) during the first 10 min. However, \(^{13}CO_2/^{12}CO_2\) breath ratios reached a plateau after \(\approx 60\) min (Fig. 3).

\([U-^{13}C]\)-tracer oxidation

No differences were observed between the \(^{13}CO_2\) production rates of the three trials (Table 1). During the 105–120 min period of exercise, the mean percentage oxidation of the infused tracer was \(99 \pm 3\%\) for Fast, \(98 \pm 1\%\) for Lo-Glu and \(96 \pm 1\%\) for Hi-Glu. The exact values for the percentage oxidation of the infused tracer for the 90–105 and 105–120 min period are displayed in Table 1.

Rate of appearance and disappearance of plasma glucose, glucose carbon recycling and metabolic clearance rate

The increase in plasma \(^{14}C/^{12}C\)-glucose ratio and \(^2\)H-enrichment as a result of the tracer infusion was stable during the 60–120 min period in all trials (Fig. 3). \(R_a\) and \(R_d\) glucose were calculated from both tracers ([6,6\(^2\)H\(_2\)]-glucose and \([U-^{13}C]\)-glucose) and were slightly lower when using the plasma \(^{14}C\)-glucose enrichment (Table 3, Fig. 4). Both \(R_a\) and \(R_d\) glucose were markedly elevated with

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**Table 2. Effects of ingestion of a low dose of glucose (Lo-Glu) or a high dose (Hi-Glu) during exercise compared with fasting (Fast) on various aspects of CHO metabolism during exercise including total CHO oxidation, the rate of appearance of plasma glucose (\(R_a\) glucose), the rate of disappearance of plasma glucose (\(R_d\) glucose), plasma glucose oxidation, glycogen oxidation and/or lactate oxidation**

<table>
<thead>
<tr>
<th>Trial</th>
<th>Time (min)</th>
<th>Total CHO oxidation ((\mu)mol kg(^{-1}) min(^{-1}))</th>
<th>(R_a) glucose(^a) ((\mu)mol kg(^{-1}) min(^{-1}))</th>
<th>(R_d) glucose(^a) ((\mu)mol kg(^{-1}) min(^{-1}))</th>
<th>Plasma glucose oxidation ((\mu)mol kg(^{-1}) min(^{-1}))</th>
<th>Percentage of (R_a) oxidized (%)</th>
<th>Muscle glycogen and/or lactate oxidation ((\mu)mol kg(^{-1}) min(^{-1}))</th>
</tr>
</thead>
<tbody>
<tr>
<td>Fast</td>
<td>90–105</td>
<td>130 ± 13</td>
<td>51 ± 3</td>
<td>53 ± 3</td>
<td>52 ± 3</td>
<td>99 ± 3</td>
<td>78 ± 12</td>
</tr>
<tr>
<td></td>
<td>105–120</td>
<td>130 ± 11</td>
<td>51 ± 3</td>
<td>52 ± 3</td>
<td>50 ± 2</td>
<td>99 ± 3</td>
<td>79 ± 11</td>
</tr>
<tr>
<td>Lo-Glu</td>
<td>90–105</td>
<td>156 ± 10</td>
<td>71 ± 1*</td>
<td>73 ± 1*</td>
<td>72 ± 2</td>
<td>99 ± 1</td>
<td>84 ± 11</td>
</tr>
<tr>
<td></td>
<td>105–120</td>
<td>155 ± 10</td>
<td>73 ± 3*</td>
<td>74 ± 5*</td>
<td>72 ± 4</td>
<td>98 ± 1</td>
<td>82 ± 13</td>
</tr>
<tr>
<td>Hi-Glu</td>
<td>90–105</td>
<td>182 ± 5</td>
<td>109 ± 8*†</td>
<td>111 ± 8*†</td>
<td>109 ± 8*†</td>
<td>98 ± 1</td>
<td>73 ± 9</td>
</tr>
<tr>
<td></td>
<td>105–120</td>
<td>179 ± 6</td>
<td>119 ± 10*†</td>
<td>117 ± 9*†</td>
<td>113 ± 9*†</td>
<td>97 ± 1</td>
<td>66 ± 6</td>
</tr>
</tbody>
</table>

Values are means ± s.e.m. for six subjects. Plasma glucose oxidation is calculated as the product of \(R_d\) glucose and the percentage of infused \([U-^{13}C]\)-glucose tracer oxidized. Glycogen and direct or indirect oxidation via lactate is calculated as the difference between total CHO oxidation and plasma glucose oxidation. \(^a\)\(R_a\) glucose and \(R_d\) glucose are calculated using the \([6,6-^{2}\)H\(_2\)]-glucose tracer. \(*\) Glu trials significantly different from the Fast trial; \(P<0.05.\) † Hi-Glu trial significantly different from the Lo-Glu trial; \(P<0.05.\)
glucose ingestion (44% with Lo-Glu and 125% with Hi-Glu; P < 0.05; Table 2).

Glucose carbon recycling was similar in all conditions. At 105–120 min, recycling rates were 5 ± 3, 6 ± 6 and 6 ± 8 \( \text{µmol kg}^{-1} \text{min}^{-1} \) during Fast, Lo-Glu and Hi-Glu, respectively, representing 4–10% of total \( R_a \text{glucose} \).

MCR was significantly higher with ingestion of glucose (Fast versus Lo-Glu) and was further increased when large amounts of glucose were ingested (Hi-Glu; Table 3). During Hi-Glu MCR was almost twice the MCR during Fast.

**Oxidation of plasma glucose**

Plasma glucose oxidation showed an identical pattern to that of the \( R_a \) and \( R_d \) glucose. Lo-Glu caused a 44% increase in plasma glucose oxidation compared with Fast and Hi-Glu produced a 126% increase. The rate of disappearance ([6,6-H\(_2\)]-glucose tracer) and the rate of oxidation ([U-\(^{13}\)C]-glucose tracer) were similar in all conditions. At 105–120 min, recycling rates were 5 ± 3, 6 ± 6 and 6 ± 8 \( \text{µmol kg}^{-1} \text{min}^{-1} \) during Fast, Lo-Glu and Hi-Glu, respectively, representing 4–10% of total \( R_a \text{glucose} \).

### Table 3. Comparison of tracers

<table>
<thead>
<tr>
<th>Trial</th>
<th>Time (min)</th>
<th>( R_a \text{glucose} ) ( \text{µmol kg}^{-1} \text{min}^{-1} )</th>
<th>( R_d \text{glucose} ) ( \text{µmol kg}^{-1} \text{min}^{-1} )</th>
<th>( R_d \text{glucose} ) ( \text{µmol kg}^{-1} \text{min}^{-1} )</th>
<th>( R_d \text{glucose} ) ( \text{µmol kg}^{-1} \text{min}^{-1} )</th>
</tr>
</thead>
<tbody>
<tr>
<td>Fast</td>
<td>90–105</td>
<td>51 ± 3</td>
<td>42 ± 4</td>
<td>53 ± 3</td>
<td>44 ± 4</td>
</tr>
<tr>
<td></td>
<td>105–120</td>
<td>51 ± 3</td>
<td>46 ± 5</td>
<td>52 ± 3</td>
<td>46 ± 5</td>
</tr>
<tr>
<td>Lo-Glu</td>
<td>90–105</td>
<td>71 ± 1*</td>
<td>65 ± 5*</td>
<td>73 ± 1*</td>
<td>67 ± 5*</td>
</tr>
<tr>
<td></td>
<td>105–120</td>
<td>73 ± 3*</td>
<td>67 ± 5*</td>
<td>74 ± 3*</td>
<td>68 ± 6*</td>
</tr>
<tr>
<td>Hi-Glu</td>
<td>90–105</td>
<td>109 ± 8†</td>
<td>105 ± 5†</td>
<td>111 ± 8†</td>
<td>107 ± 5†</td>
</tr>
<tr>
<td></td>
<td>105–120</td>
<td>119 ± 10*†</td>
<td>113 ± 6†</td>
<td>117 ± 9†</td>
<td>111 ± 6†</td>
</tr>
</tbody>
</table>

The rate of appearance of plasma glucose (\( R_a \text{glucose} \)) and the rate of disappearance of plasma glucose (\( R_d \text{glucose} \)) calculated from the [6,6-H\(_2\)]-glucose tracer compared with the [U-\(^{13}\)C]-glucose tracer. Values are means ± s.e.m. for 6 subjects. * Glu trials significantly different from the fast trial; P < 0.05. † Hi-Glu trial significantly different from the Lo-Glu trial; P < 0.05.

---

**Figure 3. Breath and plasma glucose enrichments**

Values are means ± s.e.m. Plateau in the breath \(^{13}\)CO\(_2/^{12}\)CO\(_2\) enrichment, the plasma glucose \(^{13}\)C-enrichment and the plasma \(^{13}\)C\(_2\)H\(_4\)-glucose enrichment during exercise at 50% \( V_o_{\text{max}} \) with water ingestion (Fast; □), a low dose of glucose (Lo-Glu; ●) or a high dose of glucose (Hi-Glu; ○). Breath \(^{13}\)CO\(_2/^{12}\)CO\(_2\) enrichment is also displayed for the background (BKG) correction trials (squares).
Muscle glycogen oxidation

Furthermore, whole body muscle glycogen oxidation (total CHO oxidation minus plasma glucose oxidation) was not reduced by CHO ingestion. Figure 6 describes the shifts in substrate utilization as a result of glucose ingestion. It should be noted that glucose feedings, markedly reduced fat oxidation and increased plasma glucose oxidation but did not alter the rate of muscle glycogen (and lactate) oxidation.

DISCUSSION

Effect of carbohydrate ingestion on muscle glycogen oxidation

Here we report that muscle glycogen oxidation during exercise, measured using an indirect tracer approach, was not reduced by glucose ingestion. Even when large amounts of glucose were ingested (180 g h⁻¹) and plasma glucose and insulin levels were slightly, but significantly elevated, glycogen oxidation was similar to the fasted condition. This is in agreement with several direct measurements of muscle glycogen concentrations in muscle biopsies before and after continuous cycling exercise with or without carbohydrate ingestion (Fielding et al. 1985; Coyle et al. 1986; Flynn et al. 1987; Hargreaves & Briggs, 1988; Bosch et al. 1994). However, studies in runners showed that carbohydrate ingestion reduced the net glycogen breakdown in type I fibres and this might have been responsible for the observed improvements in endurance capacity (Tsintzas et al. 1995, 1996). Tsintzas & Williams (1998) suggested that continuous cycling would cause less marked changes in plasma glucose concentration and insulin concentrations compared with running and that these differences might play an important role in the effect on glycogen utilization to be observed. However, in the present study we observed a large insulin response to the carbohydrate feedings and also plasma glucose concentration was increased. Yet, this did not result in a decrease in muscle glycogen oxidation.

Figure 4. $R_d$ and $R_a$ glucose

All values are means ± s.e.m. $A$, comparison of the rate of disappearance of glucose ($R_d$) measured by a [$^{1}$H]$\text{H}$ glucose tracer and the rate of plasma glucose oxidation ($R_{ox}$). $B$, comparison of two different tracers to measure the rate of appearance of glucose ($R_a$). The line represents the 1 : 1 relationship.

Figure 5. $R_d$ glucose versus plasma glucose oxidation

Values are means ± s.e.m. The rate of disappearance of glucose ($R_d$) and the rate of plasma glucose oxidation ($R_{ox}$) during the 60–120 min period with ingestion of water (Fast; open circles and squares), a low dose of glucose (Lo-Glu; shaded circles and squares) or a high dose of glucose (Hi-Glu; filled circles and squares). The $R_d$ presented is calculated from measurements of [6,6-$^{1}$H]$\text{H}$ glucose with GC-MS.
in 'glycogen sparing' as proposed by Taintzas & Williams (1998). Therefore, we feel that factors other than insulin are responsible for the discrepant findings in running (Taintzas et al. 1995, 1996) and cycling (Fielding et al. 1985; Coyle et al. 1986; Flynn et al. 1987; Hargreaves & Briggs, 1988; Bosch et al. 1994).

It could be argued the exercise intensity in the present study was too low to increase glycogen breakdown and therefore the effects of glucose ingestion on glycogenolysis were small and not measurable. However, other studies (Coyle et al. 1986; Bosch et al. 1994) showed that exercise intensities of 70–75% \( V_{\text{O}_2, \text{max}} \) did not result in glycogen sparing with glucose ingestion. In addition, higher exercise intensities might have abolished the insulin effect and, therefore, it would be more likely that glycogen sparing at lower exercise intensities was observed.

**Percentage of \( R_d \) oxidized**

Another important finding of this study was that the rate of disappearance (\( R_d \)) of glucose was closely matched by the plasma glucose oxidation rate (Figs 4 and 5). The percentages of \( R_d \) glucose oxidized were very high in the present study (96–100%), but are in agreement with studies by Coggan et al. (1991, 1992), who observed that 93% of \( R_d \) glucose was oxidized during exercise at 70% \( V_{\text{O}_2, \text{max}} \) (Coggan et al. 1991). Others have found that lower percentages of \( R_d \) glucose were oxidized during exercise at intensities comparable with the intensity in the present study (50% \( V_{\text{O}_2, \text{max}} \)) (Colberg et al. 1994; Roberts et al. 1996; Friedlander et al. 1997). For instance Roberts et al. (1996) observed that only 33–38% of \( R_d \) glucose was oxidized and Colberg et al. (1994) reported 33–50% of \( R_d \) was oxidized. However, these studies were performed at lower absolute exercise intensities and did not prime the bicarbonate pool as was done in the present study and it is thus possible that a fair amount of \(^{13}\text{CO}_2\) was temporarily trapped in the bicarbonate pool which may have caused a marked underestimation of the true plasma glucose oxidation. Other studies in which a bicarbonate prime was given showed values more comparable with those in the present study (88–94%) (Coggan et al. 1992).

It has been reported that when using a \(^{13}\text{C}\)-tracer for studying fatty acid metabolism part of the tracer may be trapped in exchange reactions with the tricarboxylic acid (TCA) cycle (Sidossis et al. 1995; Schrauwen et al. 1998). For example, some \(^{13}\text{C}\)-label may be incorporated into the glutamate–glutamine pool via \( \alpha \)-ketoglutarate, or into phosphoenolpyruvate via oxaloacetate. This label fixation results in a decreased recovery of label in the expired gases and in order to correct for this loss, the acetate correction factor has been proposed (Sidossis et al. 1995). The label loss is dependent on metabolic rate and at high oxygen uptakes (38–39 ml kg\(^{-1}\) in the present study) recovery of the \([1,13\text{C}]\)-acetate label has been shown to be 85–90% (Sidossis et al. 1995). Besides that, \([U-13\text{C}]\)-glucose has six labelled carbons of which two will appear directly in \(^{13}\text{CO}_2\) and, therefore, do not enter the TCA cycle. Only two-thirds (66%) of the glucose carbons is subject to label fixation. Therefore the recovery of carbons from \([U-13\text{C}]\)-glucose will be higher than the recovery of \([1,13\text{C}]\)-palmitate (Sidossis et al. 1995) or \([U-13\text{C}]\)-palmitate (Schrauwen et al. 1998), explaining why we find that 96–100% of the infused glucose tracer is oxidized. The small difference from 100% may be explained by the acetate correction factor implying that all glucose molecules disappearing from the plasma might have been oxidized both in the presence and absence of glucose ingestion.

**Plasma glucose turnover and the rate of ingestion**

Plasma glucose has been shown to be an important substrate during exercise, especially during the later stages when muscle glycogen levels are reduced (Coggan, 1991). Here we observed that plasma glucose contributed 19% to energy expenditure during fasting and this value increased to 42% during Hi-Glu ingestion (Fig. 6). During Hi-Glu ingestion, plasma glucose was the most important substrate from a quantitative point of view.

Glucose turnover was markedly increased by the glucose ingestion as evidenced by increased \( R_a \) and \( R_d \) glucose, increased plasma glucose oxidation and metabolic clearance rate (MCR). The Lo-Glu ingestion increased \( R_a \) glucose by 44% and Hi-Glu ingestion increased \( R_d \) glucose by 126%. However, whereas glucose ingestion during Hi-Glu was five times larger than during Lo-Glu, the \( R_d \) glucose increased only less than twofold (72 ìmol kg\(^{-1}\) min\(^{-1}\) during Lo-Glu and 109–113 ìmol kg\(^{-1}\) min\(^{-1}\) during Hi-Glu). Mean ingestion during the second hour of exercise was 0·3 g min\(^{-1}\) in the Lo-Glu and 1·5 g min\(^{-1}\) in the Hi-Glu trials. The total rates of appearance of glucose were 0·84 and 1·36 g min\(^{-1}\) in the Lo-Glu and 1·5 g min\(^{-1}\) during Hi-Glu ingestion).

Figure 6. The energetic contribution of substrates

Values are means ± s.e.m. The energetic contribution of substrates during the 60–120 min period of exercise at 50% \( V_{\text{O}_2, \text{max}} \) with water ingestion (Fast), a low dose of glucose (Lo-Glu) or a high dose of glucose (Hi-Glu).
are needed to distinguish between gut-derived $R_g$ and hepatic glucose output. Without glucose ingestion the hepatic glucose production was approximately 0.5 g min$^{-1}$ indicating that the liver during the second hour of exercise may have produced about 30 g glucose. This is a relatively large amount since it is believed that the liver contains approximately 80 g of glycogen (Hultman & Nilsson, 1971). Glucose ingestion during exercise may reduce hepatic glucose production (i.e. spare liver glycogen) at 70% $V_{O_2, max}$ as shown by Bosch et al. (1994) and McConell et al. (1994).

**Limitations of exogenous carbohydrate oxidation**

These results also indicate that, with large doses of ingested glucose (360 g in the 2 h exercise period), not all the glucose appears in the plasma. This is in agreement with previous suggestions that the rate of exogenous glucose oxidation may be limited by the rate of digestion, absorption, maximal hepatic glucose output and subsequent transport of glucose into the systemic blood supply rather than by glucose uptake and oxidation by the muscle (Hawley et al. 1994). Several studies showed that oral carbohydrate oxidation is limited to about 1 g min$^{-1}$ (Hawley et al. 1992). Even when large amounts of carbohydrate were ingested oxidation rates did not exceed 1 g min$^{-1}$ (Rehrer et al. 1992; Wagenmakers et al. 1993).

**Effects of carbohydrate ingestion on fat and carbohydrate oxidation**

Glucose ingestion also markedly suppressed fat oxidation compared with fasting and this effect was dose dependent: Hi-Glu suppressed fat oxidation more than Lo-Glu. The lower rates of fat oxidation may be at least in part insulin mediated (Fig. 1). Insulin has been shown to be a potent inhibitor of lipolysis and $R_g$ FFA (Montain et al. 1991; Campbell et al. 1992; Horowitz et al. 1997). Here, higher insulin levels after glucose ingestion reduced whole body lipolysis as indirectly indicated by lower plasma glycerol and FFA concentrations (Fig. 1). Decreased FFA availability and higher plasma insulin concentrations along with an increased glycolytic flux have been shown to reduce FFA oxidation at rest (Sidossis & Wolfe, 1996) and during exercise (Coyle et al. 1997). Although in this study we observed that glucose ingestion during exercise increased total CHO oxidation, McConell et al. (1994) and Bosch et al. (1994) did not observe increased CHO oxidation rates with CHO ingestion during exercise. This may, in part, be explained by the differences in the exercise intensity and possibly the difference in plasma insulin concentration, which may have been higher in the present study. Whereas in their studies the exercise intensities were 70% $V_{O_2, max}$, we employed an exercise intensity of 50% $V_{O_2, max}$. At low to moderate exercise intensities (50% $V_{O_2, max}$) glucose utilization may be enhanced by CHO ingestion whereas during more intense exercise (> 70% $V_{O_2, max}$) exogenous CHO simply seems to substitute for endogenously produced glucose, with no overall increase in CHO oxidation (Bosch et al. 1994; McConell et al. 1994).

**Gluconeogenesis (glucose carbon recycling)**

When measured by GC-IRMS [U-$^{13}$C]-$\text{glucose}$ carbon may recycle during the process of gluconeogenesis and this may lead to an underestimate of the $R_g$ of glucose. The reason for this is the following. One molecule of uniformly labelled glucose produces two molecules of uniformly labelled pyruvate. This pyruvate may be converted into lactate or alanine (predominantly lactate) which are precursors for gluconeogenesis and this may lead to the incorporation of label into newly synthesised glucose. Fifty percent of the carbon atoms will be $^{13}$C in the newly synthesised glucose.

In GC-combustion-IRMS analysis the whole molecule is oxidized to CO$_2$ prior to the enrichment measurement and therefore the increase in ratio $^{14}$C/$^{13}$C of all carbons of the glucose molecule is measured. Tracer dilution measurements by GC-combustion-IRMS analyses of this tracer therefore exclude gluconeogenesis and consequently represent a possible underestimation of the true $R_g$ of glucose. It is believed that a more appropriate method of measuring total $R_g$ glucose is the use of a [6,6-D$_2$]-glucose tracer which measures total glucose production when the enrichment is measured by GC-MS as the tracer atoms do not recycle during gluconeogenesis (Wolf & Wolfe, 1992). To compare the above mentioned methods we also infused a [6,6-D$_2$]-glucose tracer. The difference between the two methods theoretically gives an estimate of glucose carbon recycling and a rough estimate of gluconeogenesis (Jeukendrup & Wagenmakers, 1996). In this study we found that $R_g$ and $R_h$ glucose were similar, regardless of the tracer used (i.e. [U-$^{13}$C]-$\text{glucose}$ measured by GC-IRMS or [6,6-D$_2$]-glucose measured by GC-MS) (Table 3). In Fig. 4, $R_g$ glucose calculated from the [6,6-D$_2$]-glucose tracer is plotted against the $R_g$ glucose calculated from the [U-$^{13}$C]-$\text{glucose}$ tracer (total enrichment). This figure shows that, generally, $R_g$ glucose calculated from the [U-$^{13}$C]-$\text{glucose}$ tracer gave slightly lower numbers, which is in agreement with the expectations. The recycling rate of $^{13}$C was low (~4-10 % of total $R_g$ glucose) and was not reduced by the glucose feedings, suggesting that independently of the amount of glucose ingested, gluconeogenesis was minimal. These low rates of recycling may have resulted from low precursor availability (i.e. low lactate concentrations as observed in previous studies with similar designs (Jeukendrup et al. 1996a,c, 1997b). Gluconeogenesis may also be lower in trained individuals (Coggan, 1997) and since the training level of the subjects in the present study was very high, the rates of gluconeogenesis may have been very low. Friedlander et al. (1997) also suggested that retnocceptors were downregulated in response to endurance training and this may be a mechanism for the blunted gluconeogenesis. With the high training level of the athletes in the present study this could be an explanation for the low levels of glucose carbon recycling as well.

**General overview: effect of carbohydrate feedings on substrate metabolism**

The results of the present study indicate that glucose ingestion during exercise leads to a decreased fat oxidation,
partly because of an inhibition of lipolysis (Horowitz et al. 1997) and an increased carbohydrate oxidation. The increased carbohydrate oxidation was explained by an accelerated plasma glucose turnover. The appearance of glucose in the bloodstream increased with carbohydrate feedings and this increase was dose dependent. However, at very high ingestion rates the appearance of glucose in plasma levelled off.

The increased rate of appearance of glucose did not lead to a large increase in the plasma glucose concentration because the rate of disappearance is also increased. Higher glucose concentrations may have been partly responsible for the increased glucose uptake during Hi-Glu, simply by mass action. However, the augmented glucose uptake was mainly explained by an increased clearance of glucose from the bloodstream. This seems to suggest that glucose transport was activated by mechanisms involving GLUT4 translocation, activation of the oxidative enzymes (phosphofructokinase (PFK) and/or activation of the pyruvate dehydrogenase (PDH) complex).

The glucose that disappeared from the plasma (and most likely was taken up by active skeletal muscle) was oxidized and not used for glycogen synthesis. Muscle glycogen oxidation was not reduced by the glucose feedings. Glycogenneogenesis in the liver was negligible in all conditions.

In summary, a method was developed to measure $R_g$ glucose and plasma glucose oxidation by using small amounts of a [U-13C]-glucose tracer and measuring plasma tracer enrichment with GC-IRMS. The method seems to be valid to make estimates of glucose turnover and for indirect estimates of muscle glycogen consumption at whole body level. The results of this study indicate that carbohydrate feeding during cycling exercise at 50% $V_{0,\text{max}}$ in highly trained subjects decreased fat oxidation and increased the $R_g$ of glucose, the MCR and plasma glucose oxidation. Muscle glycogen oxidation was not reduced even when large amounts of glucose were ingested and non-oxidative glucose disposal was negligible. A large percentage (96–100%) of glucose disappearing from the plasma was oxidized. Comparison of $R_g$ glucose calculated from [6,6-2H2]- and [U-13C]-glucose tracers revealed that only a small quantity of glucose carbons is recycled in the process of glycogenneogenesis.


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