Strategies for optimization of Aminoglycoside and Vancomycin therapies : a Pharmacokinetic approach

Citation for published version (APA):

van Maarseveen, E. M. (2014). *Strategies for optimization of Aminoglycoside and Vancomycin therapies : a Pharmacokinetic approach*. [Doctoral Thesis, Maastricht University]. Maastricht University. <https://doi.org/10.26481/dis.20141128em>

Document status and date:

Published: 01/01/2014

DOI:

[10.26481/dis.20141128em](https://doi.org/10.26481/dis.20141128em)

Document Version:

Publisher's PDF, also known as Version of record

Please check the document version of this publication:

- A submitted manuscript is the version of the article upon submission and before peer-review. There can be important differences between the submitted version and the official published version of record. People interested in the research are advised to contact the author for the final version of the publication, or visit the DOI to the publisher's website.
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Strategies for Optimization of Aminoglycoside and Vancomycin Therapies

A **p**harmaco**K**inetic
Approach

Strategies for Optimization of Aminoglycoside and
Vancomycin Therapies

© 2014 E.M. van Maarseveen, Utrecht The Netherlands
Lay-out and cover design by Marieke Velthuis

Printed by: Gildeprint – The Netherlands

Printed on FSC certified paper
ISBN/EAN 9789461087966

Strategies for Optimization of Aminoglycoside and Vancomycin Therapies

A Pharmacokinetic Approach

PROEFSCHRIFT

ter verkrijging van
de graad van doctor aan de Universiteit Maastricht,
op gezag van de Rector Magnificus, Prof dr. L.L.G. Soete
volgens het besluit van het College van Decanen,
in het openbaar te verdedigen
op vrijdag 28 november 2014 om 14.00 uur

door

Erik Michiel van Maarseveen

Geboren op 17 mei 1979 te Tiel

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Chapter 1

General introduction

1.1 Aminoglycosides

Aminoglycosides are potent bactericidal antibiotics and used to treat severe infections primarily caused by Gram-negative bacteria, staphylococci as well as certain mycobacteria. Most common indications for the parenteral formulation are urinary tract, intra-abdominal and surgical infections, infective endocarditis, (neonatal) sepsis, and pulmonary exacerbation of cystic fibrosis (CF) (1, 2). Aminoglycosides are a cornerstone of the current antibiotic armamentarium because of their activity against multidrug-resistant Gram negative pathogens such as *Pseudomonas aeruginosa* and *Acinetobacter* spp (2, 3). Furthermore, to kill host pathogens combinations of aminoglycosides and beta-lactam antibiotics act synergistically and combining both antibiotic classes lowers the risk of bacterial resistance (4). Although very effective, aminoglycosides have a narrow therapeutic range and can induce both severe nephrotoxicity and ototoxicity (3, 4).

Mechanism of action

Aminoglycosides inhibit the protein synthesis of microorganisms and as a result alter the integrity of the bacterial cell membrane. Aminoglycosides bind to negatively charged residues in the outer membrane of Gram-negative bacteria (5). When the antibiotic diffuses through the outer membrane, low affinity binding of aminoglycosides to an energy-dependent transport system occurs. The transport across the cytosolic membrane is the rate-limiting step and can be blocked by calcium and magnesium ions, hyperosmolarity, low pH and anaerobic conditions. In the cytosol, aminoglycosides bind to 30S subunit ribosomes and block the translation process resulting in the disruption of chain elongation of proteins. Aberrant proteins are then inserted into the cell membrane, resulting in altered permeability and further stimulation of transmembrane aminoglycoside transport (6). Finally, disruption of the structure of the cell membrane leads to a leakage of intracellular contents, which precedes cell death (7).

Efficacy

The bactericidal activity of aminoglycosides is concentration-dependent (8). Importantly, aminoglycosides have demonstrated a post-antibiotic effect (PAE) against gram-negative bacteria (9). PAE refers to continued suppression of bacterial growth after antibiotic concentrations have dropped below the Minimal Inhibitory Concentration (MIC) (10). This allows a longer dose interval than predicted based on the MIC alone (11). The pharmacokinetic/pharmacodynamic predictors associated with efficacy are ratio of peak plasma drug concentration over the MIC of the causative pathogen (C_{max}/MIC) and ratio of the area under the concentration-time curve (AUC) over MIC (AUC/MIC) (12). Target C_{max} and AUC values of respectively 8-10 mg/L and 74-100 mg.h.L⁻¹ are recommended (1, 2). For microorganisms with an MIC of 2 mg/L, a C_{max} of at least 20 mg/L is recommended (1).

Toxicity

Nephro- and ototoxicity are the most frequently observed side effects of aminoglycosides (13). Aminoglycosides induce nephrotoxicity in an estimated 0-35% of treated patients based on reports from literature (14). Albeit aminoglycoside-associated nephrotoxicity is generally defined by an increase in serum creatinine by more than 0.5 mg/dL (44 nmol/mL) or increase more than 50% in serum creatinine from baseline, clinical studies report divergent definitions of nephrotoxicity which may partially explain this broad range of reported incidences of nephrotoxicity. Without intervention aminoglycoside-induced nephrotoxicity eventually manifests as nonoliguric renal failure (15). It is characterized by a decrease in glomerular filtration rate, tubular necrosis and as a consequence a rise in serum creatinine (14, 15).

After glomerular filtration, approximately 15% of the amount of filtered aminoglycoside is reabsorbed into the renal tissue (16) and retained in the epithelial lining cells of the proximal tubules. Aminoglycosides enter the endosomal compartment through endocytosis via the megalin-cubilin complex (17). When the concentrations of aminoglycosides exceed an undetermined threshold, the endosomal membrane destabilizes and aminoglycosides are released into the cytosol (17, 18). Aminoglycosides bind to phospholipids membranes to cause phospholipidosis (17). Cytosolic aminoglycosides and phospholipidosis promote cell apoptosis

(15, 17, 18). Aminoglycoside-associated nephrotoxicity is directly correlated to accumulation of the drug in the renal proximal tubules. This accumulation depends on several factors such as the number of amino groups and the distribution of these groups within the molecule and thus can differ within the aminoglycoside drug class. Aminoglycosides with a high number of positively charged amino groups at physiological pH are more likely to cause nephrotoxicity, perhaps due to enhanced interactions with anionic membrane phospholipids and megalin (15, 17, 18). Drug steric hindrance can also influence the affinity of the drug to the receptor binding domain. These factors are responsible for the cationic structure of the molecule and determine the potency of the binding affinity of aminoglycosides to the brush border's megalin receptor (15, 17, 18). Figure 1 displays potential cellular mechanisms of toxicity of the aminoglycoside gentamicin.

Ototoxic side effects of aminoglycosides usually occur within days or weeks after initiation of systemic application (19). Aminoglycosides can induce irreversible damage both to the auditory and vestibular organs. Gentamicin and tobramycin are predominantly vestibulotoxic, while neomycin, kanamycin and amikacin are mainly cochleotoxic (20). The prevalence of vestibulotoxicity and cochleotoxicity are 15% and 2-25%, respectively (21, 22). Symptoms of cochleotoxicity are hearing loss and/or tinnitus, while those of vestibulotoxicity are dizziness and light-headedness. Cochleotoxicity initially affects the high auditive frequencies and subsequently may extend towards the lower frequencies (19). Threshold monitoring by pure-tone audiograms in the conventional <8kHz and high-frequency regions 8 – 16 or 20 kHz is recommended (23). Aminoglycoside ototoxicity is characterized by degeneration of hair cells located in the organ of Corti, particularly in the basal turn which is required to sense high-frequency sounds. In rare cases aminoglycosides can also cause damage to nervus VIII (nervus vestibulocochlearis) resulting in complete deafness (24).

Figure 1. Potential cellular mechanism of gentamicin toxicity (printed with permission and at the courtesy of Dr. Andrew Prayle, Nottingham, UK)

This diagram summarizes the major mechanisms of nephrotoxicity from the literature, mainly from in vitro research and as such has to be interpreted as a model. Gentamicin enters the tubular cell via the multiligand receptor megalin and subsequently uptake is via a clathrin-coated pit. Gentamicin retrogradely passes through the lysosomes, Golgi and endoplasmic reticulum into the cytoplasm where it forms complexes with iron which catalyse the formation of reactive oxygen species (ROS). In the lysosomes, gentamicin causes release of the pro-apoptotic cathepsins and, in the endoplasmic reticulum, gentamicin causes release of the pro-apoptotic stress protein von Hippel–Lindau binding protein 1 (VBP1). Cytosolic gentamicin either directly or indirectly (via ROS) causes the release of cytochrome C from the mitochondria, a key step in apoptosis. ROS cause an increase in the gene expression of oxidative stress-inducible genes, stress-inducible chaperones and oxido-reductive enzymes. Gentamicin may have intracellular effects without entry to the cell. It can cause a rapid increase in intracellular Ca^{2+} and an early increase in the anti-apoptosis and proliferative signals Akt and Erk; this may be mediated by the membrane Ca^{2+} receptor CaR.

Aminoglycoside pharmacology and monitoring in highly exposed populations

Patients suffering from cystic fibrosis (CF), severe infections in the intensive care unit (ICU) or neonatal sepsis caused by Gram-negative bacilli are frequently treated with aminoglycosides. In the following paragraphs aminoglycoside therapy in these specific patient populations is addressed.

Cystic fibrosis patients

During childhood CF lungs are predominately colonized with *Staphylococcus aureus* and/or *Haemophilus influenzae*, and at adolescent and adult ages with *Pseudomonas aeruginosa* as the disease progresses (25). Intravenous aminoglycosides in combination with beta-lactam antibiotics are commonly used to treat moderate to severe acute pulmonary exacerbations of CF caused by the latter pathogen (26, 27). Aminoglycoside pharmacokinetics in CF patients show specific differences from other aminoglycoside treated populations. Malnourished CF patients due to the reduced adipose tissue mass, will have an increase in lean tissue per kg bodyweight (28, 29). As the volume of distribution is related to the lean body mass (LBM), a higher dose is required to attain predefined C_{max}/MIC targets. Additionally, AUC/MIC ratios are affected by a relatively high renal clearance in CF patients compared to other clinically infected patient populations and is associated with their relatively young age (12). Once-daily aminoglycoside treatment confers equivalent efficacy to a multiple-daily dosing, however has shown to be less nephrotoxic in children with CF (30-32). The current standard duration for intravenous aminoglycoside therapy is 14 days or longer in case of severe exacerbations or incomplete recovery (27). The present recommended intravenous tobramycin dose for adult CF patients is 10 mg/kg/day and 30-35 mg/kg/day for amikacin (33). Since CF patients usually are exposed to multiple courses of prolonged aminoglycoside exposure during their life, they are at high risk of developing aminoglycoside side effects such as nephrotoxicity (34). Not only monitoring of the renal function, but also therapeutic drug monitoring (TDM) is essential in these patients to optimize the efficacy and limit acute and cumulative toxicity.

The C_{max} targets of tobramycin and amikacin are 20–40 mg/L and 80–120 mg/L, respectively, with a C_{min} target of <1 mg/L for both aminoglycosides (33).

Critically ill patients

Aminoglycosides play an important role in the treatment of critically ill patients with severe sepsis, hospital-acquired pneumonia, ventilator-associated pneumonia, health care-associated pneumonia, urinary tract infections, endocarditis, bacterial meningitis and febrile neutropenia (35). In critically ill patients a large variability in aminoglycoside distribution and clearance due to pathophysiological changes and concomitant therapies compromises the attainment of predefined exposure targets. For instance, fluid resuscitation and total parenteral nutrition expand the extracellular fluid content and dilute serum and extracellular aminoglycoside concentrations. Meanwhile, patients with renal dysfunction and decreased cardiac output have a decreased clearance and therefore are prone to develop aminoglycoside-associated nephrotoxicity (36, 37). Achievement of target peak serum concentration is related to the volume of distribution. Thus, dose modification of aminoglycosides will be required more often in populations with high inter- and intra-patient variability in volume of distribution and/or clearance (35). Several nomograms recommend that the dosages of aminoglycosides should be higher compared to other adult non-CF populations; gentamicin or tobramycin 7 mg/kg and amikacin 15 mg/kg.

Although these dosing nomograms produce good results in clinical practice, many ICU patients do not achieve the target peak or trough serum concentration independent of the selection of a particular ‘a priori’ dosing algorithm (38–40). Hence, TDM of aminoglycosides is essential in critically ill patients to guarantee efficacy and prevent toxicity in this vulnerable population.

Neonates

Neonatal early onset infections are usually of maternal-foetal (perinatal) origin and typically occur in the first 5 days of life caused by contaminated amniotic fluid or blood. The predominant pathogens in term neonates include group B streptococci, *Escherichia coli*, enterococci and *Listeria monocytogenes*. In preterm neonates, group B streptococci, *Escherichia coli*, *Haemophilus influenzae* and Gram-negative bacilli

are predominant (41). Late-onset infections, on the other hand, occur in the first month of life, particularly from contaminated hands or venous access. Coagulase-negative staphylococci are the main pathogens, especially *Staphylococcus epidermis*. *Pseudomonas* spp, *Klebsiella* spp and *Serratia* spp are also common pathogens. Neonates demonstrate aberrant pharmacokinetics compared to adult patients. First, neonates have a relatively low fat tissue mass compared to adults. Water comprises about 60-70% of a neonate's body weight, which affects the volume of distribution of aminoglycosides (42). Therefore, a higher dose of aminoglycoside per kilogram of bodyweight is needed to achieve the target drug peak concentration (43). Furthermore, renal clearance of aminoglycosides is correlated to gestational age and accelerates directly postpartum. Because of these developmental influences, there is great inter- and intra-individual variability in the volume of distribution and clearance of aminoglycosides in neonates, (44). Currently, a minimum drug plasma concentration of <1-2 mg/L is targeted to prevent accumulation of aminoglycosides, which may lead to nephrotoxicity and ototoxicity (45). Gentamicin is the most widely used aminoglycoside in neonates. Tobramycin and netilmicin are also frequently used. Numerous gentamicin dosing regimens have been suggested for neonates, based on the bodyweight, gestational age (GA) and postnatal age (PNA) (45).

For example, the widely practiced Christchurch protocol, which was designed using data obtained from a neonatal intensive care unit (NICU) population, consists of a complex dosing equation based on birth weight, with a relatively high mg/kg dose and extended dosing intervals up to 60 hours for infants with a relatively low body weight. Prospective evaluation in over four hundred neonates showed that the vast majority (84%) of the population reaches the predefined peak concentration target of >10 mg/L and 77% reached target trough plasma concentrations <2 mg/L (46). Another dosing regimen of gentamicin intended for preterm neonates with a GA < 35 weeks recommends an initial dose of 5 mg/kg every 36 hours. Prospective evaluation showed that 56% of all peak blood concentrations and 79% of all trough concentrations were within the target values of 8-10 mg/L for peak concentration and <1 mg/L for trough concentration (47, 48). A third proposed dosing scheme of tobramycin recommends 4 mg/kg every 48 hours in neonates with GA < 32 weeks, 4 mg/kg every 36 hours in neonates with GA between

32 and 37 weeks and 4 mg/kg every 24 hours in neonates with GA >37 weeks resulting in >90% of neonates reaching target blood concentrations of 5–10 mg/L for peak concentrations and <2 mg/L for trough concentrations (49, 50). These three examples illustrate that ‘a priori’ dosing nomograms of aminoglycosides in neonates are very diverse. This large divergence can be explained by a high inter-population variation in pharmacokinetics in neonates and the predefined target exposure.

A 2006 Cochrane Collaboration review of once-daily versus several times a day dosing of gentamicin in neonates found that both methods demonstrated adequate clearance of sepsis, but once-daily dosing was associated with fewer failures to achieve appropriate peak (aiming for 8–10 mg/L) and trough (aiming for <1 or 2 mg/L) concentrations (51). Ototoxicity and nephrotoxicity were not seen with either regimen, although probably they were not investigated in depth. Once-a-day and extended-interval dosing of gentamicin in neonates is now a widely accepted practice (45, 51). As the serum creatinine concentration is a poor surrogate for glomerular function in neonates, the assessment of kidney function remains challenging. Importantly, by measurement of sensitive biomarkers clinical studies have shown tubular damage can occur in more than 50% of neonates and glomerular damage in 10% of neonates. However, the clinical relevance of these findings is currently unknown. And albeit serious hearing loss has been associated with aminoglycoside-related toxicity (52), causality has not been proven to date in neonates (53). Irrespective of the selected dosing nomogram, monitoring of serum concentrations of aminoglycosides is widely recommended and adopted to optimize the antibiotic therapy in individual neonates (45).

In conclusion

Aminoglycosides are potent antimicrobial agents. Yet, their clinical use poses serious risks of side effects. Patient populations that show aberrant aminoglycoside pharmacokinetics and/or are exposed to high dosages are potentially susceptible to aminoglycosides-related toxicity. Moreover, therapeutic drug monitoring is recommended to optimize aminoglycoside therapy in individual patients. Finally, biomarker monitoring such as monitoring serum creatinine plasma levels may reflect

the condition of the kidneys and may help identify patients at risk of developing aminoglycoside-associated nephrotoxicity.

Chapter 1.2.

Continuous and intermittent infusion of vancomycin are equally effective: *a review of the literature.*

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Adapted from Nederlands Tijdschrift voor Geneeskunde. 155(42); A2667

1.2 Vancomycin

Vancomycin is a glycopeptide antibiotic used in (suspected or proven) bacteremia, peritonitis or osteomyelitis with gram-positive micro-organisms. The compound was industrially produced by fermentation and given the generic name vancomycin, derived from the term “vanquish.” Originally, vancomycin was indicated to treat serious infections caused by penicillin-resistant Gram-positive bacteria. Vancomycin acts by abrogating proper synthesis of the cell wall in Gram-positive bacteria. Due to the different mechanisms by which Gram-negative bacteria produce their cell walls and the various factors related to entering the outer membrane of these pathogens, vancomycin is not active against Gram-negative bacteria (54). Since vancomycin is poorly absorbed after oral intake, it is generally administered through intravenous infusion. The product label states a daily dose of 2000 mg, to be infused as 500 mg every 6 hours or as 1000 mg every 12 hours (55). As there is no gastrointestinal uptake of vancomycin it is administered by intravenous infusion. The initial product label states a daily dose of 2000 mg, to be infused as 500 mg every 6 hours or as 1000 mg every 12 hours (52), which was not founded on pharmacodynamic data but was chosen arbitrarily (56). Vancomycin is recommended to be slowly administered in a dilute solution, over at least 60 minutes (maximum infusion rate of 10 mg/minute for doses >500 mg) because of the high incidence of pain during infusion, the emergence of thrombophlebitis and to avoid infusion reactions known as the red man syndrome. This syndrome is characterized by flushing or an erythematous rash of face, neck, and torso and usually appears within minutes or hours after the commencement or the completion of a vancomycin infusion. Antimicrobial killing of vancomycin is time and/or AUC dependent and requires a minimal inhibitory concentration (MIC) of vancomycin at the site of infection (57, 58). Although resistance against vancomycin and other glycopeptides is emerging worldwide among methicillin-resistant *Staphylococcus aureus* and vancomycin-intermediate *S. aureus* (VISA) strains (59), vancomycin still is the antibiotic of choice for the therapy of methicillin-resistant *S. aureus* (MRSA) infections according to recent guidelines from the Infectious Diseases Society of America (IDSA) (60). To preserve the current therapeutic armamentarium for Gram-positive infections, it is of paramount importance that guidelines

for vancomycin dosing are based on patient outcomes and on (in vitro) pharmacokinetic/ pharmacodynamic (PK/PD) data to attain maximum efficacy with minimal toxicity. Moreover, this strategy may also prevent the emerge of resistance. Although there is in vitro evidence showing that the time above the MIC and AUC/MIC-values are the most important parameters associated with efficacy, most human studies do not show a clear PK/PD relationship (58). To date, only Moise-Broder et al. found a significant in vivo PK/PD efficacy relation. They showed that for intermittent infusion of vancomycin the AUC_{24h}/MIC-ratio of <350 was associated with a lower eradication rate and higher mortality ($P = 0.005$) in patients with methicillin-resistant *S. aureus* pneumonia (57). Data from recent American studies suggest a reduced susceptibility in case of (suspected) methicillin-resistant *Staphylococcus aureus* (MRSA) infections to vancomycin urging for higher dosages (55, 61). Susceptibility to vancomycin is predicted by determining the in vitro minimum inhibitory concentration. The MIC is the concentration of vancomycin that is needed to inhibit the growth of the strain of bacteria in vitro. The Clinical and Laboratory Standards Institution (CLSI) determines breakpoints for susceptibility and resistance to vancomycin. In 2006, the *S. aureus* MIC breakpoints for vancomycin were lowered (from ≤ 4 $\mu\text{g/mL}$ to ≤ 2 $\mu\text{g/mL}$ for “susceptible,” from 8–16 $\mu\text{g/mL}$ to 4–8 $\mu\text{g/mL}$ for “intermediate,” and from ≤ 32 $\mu\text{g/mL}$ to ≤ 16 $\mu\text{g/mL}$ for “resistant”) to increase detection of heterogeneously resistant isolates of *S. aureus* (Table 1) (62). The latter is a reflection of the increasing amount of data implying that isolates of *S. aureus* are less likely to respond to vancomycin therapy. With these new breakpoints, 99% of MRSA remain within the susceptibility range and should respond to vancomycin therapy. However, three meta-analyses on vancomycin treatment of *S. aureus* infections observed MICs in the upper limit of the susceptibility range ($>1\text{--}1.5$ mg/L) to be associated with an increased rate of treatment failure and mortality (63–65), leaving room for vancomycin treatment optimization. To optimize the pharmacodynamics of vancomycin some authors suggest that the use of higher intermittent dosages could be helpful (66–68). Nevertheless, based on linear pharmacokinetics the attainment of concentrations above MIC remains a challenge with intermittent infusion (InI) in patients with a rapid clearance. Hence, it seems rational to explore alternative dosing strategies of vancomycin, for example continuous infusion (CoI).

In the following two sections the currently available studies comparing efficacy and toxicity of continuously and intermittently dosed vancomycin will be reviewed and discussed.

Table 1. Susceptibility and resistance breakpoints for vancomycin against *S. aureus*

	Current breakpoints, MIC (mg/L)	Breakpoints before 2006, MIC (mg/L)
Susceptible <i>S. aureus</i>	≤2	≤4
VISA	4-8	8-16
VRSA	≤16	≤32

Abbreviations are MIC, minimum inhibitory concentration; VISA, vancomycin-intermediate *S. aureus*; VRSA, vancomycin-resistant *S. aureus*

Vancomycin efficacy: a comparison of continuous and intermittent infusion

Vancomycin is thought to inhibit bacterial growth in a time-dependent manner and microbial killing activity therefore depends on the time that vancomycin serum concentrations exceed the MIC. However, this time dependency (time >MIC) was not found to be associated with clinical outcomes (57). The PK/PD parameter that is assumed to be the best predictor of efficacy is the area under the concentration-time curve over the minimal inhibitory concentration AUC/MIC (57, 69). The consensus guidelines by the American Society of Health-System Pharmacists (ASHP), the Infectious Diseases Society of America (IDSA) and the Society of Infectious Diseases Pharmacists (SIDP) recommend an AUC/MIC ratio ≥400 (61). This is supported by Moise-Broder et al., who found that an AUC/MIC ratio ≥350 is associated with an increased rate of successful clinical outcomes in patients with lower respiratory tract infections caused by *S. aureus* (57). However, confirmative studies to support these findings have not been reported to date. Since AUCs are not easily obtained in the clinical setting, trough serum concentrations are monitored as a surrogate marker (68). To obtain an AUC/MIC ratio of ≥400 against bacteria with an MIC ≤1 mg/L, it is

a suggested that a trough serum concentration of at least 15 mg/L is required with vancomycin InI (70). The current dosing advice is to administer 15-20 mg per kg bodyweight every 8-12 hours and to monitor of trough serum concentration, which should be 15-20 mg/L for severe MRSA infections (68, 71). Moreover, higher doses are needed to achieve an AUC/MIC of ≥ 400 for pathogens with an MIC > 1 mg/L (72). The need to improve clinical outcomes in patients infected with pathogens that have an MIC > 1 mg/L for vancomycin, has contributed to the recent debate on the preferential mode of administration of vancomycin: InI or Col. From a theoretical perspective, Col would be preferred based on the assumption that vancomycin inhibits bacterial growth in a time-dependent manner, since Col is assumed to yield more stable vancomycin serum concentrations and maintain serum concentrations that exceed the MIC. Currently, there is no international consensus on continuous dosing schedules of vancomycin. Some dosing nomograms are based on bodyweight (73, 74), where other authors propose dosing schedules based on renal function (75, 76). Nevertheless, most protocols recommend monitoring of plateau concentrations, that should reach minimum concentrations of at least 15-25 mg/L corresponding with an AUC will be between 350 and 600 mg.h.L⁻¹. Consequently, an AUC/MIC of ≥ 400 cannot be obtained with Col for organisms with MIC ≥ 2 mg/L without exceeding the target 'plateau' level recommendations. Results from clinical studies may indicate that Col is equivalent to InI based on both mortality and treatment failure data. A recent meta-analysis comparing InI to Col, showed that the relative risk (RR) for overall mortality did not differ between treatment groups (RR 1.03; CI 0.7-1.6; $p = 0.9$) (77). Since the definition of treatment failure was inconsistent in the studies included in the meta-analysis, this outcome measure could not be pooled. Verrall et al. sought to explore treatment failure between InI and Col in a retrospective study including outpatients. Treatment failure with InI was found to be equivalent to Col (RR 0.701; CI 0.432-1.136; $p = 0.159$) (78). Vuagnat and co-workers conducted a prospective study in patients with osteomyelitis and found no difference in the percentage of patients cured while treated with either InI or Col (77.8% versus 94.4%, respectively; $p = 0.25$). However, patients treated with InI were more likely to terminate treatment due to adverse reactions than those with Col (42.9% versus 8.7%; $p = 0.03$) (73). In a retrospective cohort study by Boffi El Amari et al., high dose (HD; 40 mg/kg/day) was compared to standard dose (SD; 20 mg/kg/day) in patients with osteomyelitis. The HD regimen was either infused continuously or

intermittently, whereas SD was only administered as InI. SD-InI resulted in a higher risk of treatment failure than HD (Col and InI) treatment (OR 7.5; CI 1.4-38.6). However, HD resulted in more adverse events compared to SD-InI (25% versus 4.5%; $p = 0.007$). This was mainly due to HD-InI, since the rate of adverse events in HD-Col did not differ significantly from SD-InI ($p = 0.06$). Subsequently, patients in the HD-Col group had the best outcomes (lowest number of treatment failures and adverse events) compared to the InI regimens (log rank test $p = 0.02$) (79). Currently, the only randomized clinical trial that compares InI to Col found no difference in treatment failure (19% versus 21%, respectively). However, this study by Wysocki et al. was underpowered with respect to treatment failure outcome measures (74).

Vancomycin toxicity: a comparison of continuous and intermittent infusion

Although treatment failure warrants for higher dosing regimens of vancomycin InI, the use of higher dosages – to acquire a trough level of 15-20 mg/L or higher – is limited by the risk of nephrotoxicity. Nephrotoxicity occurs in 5-43% of patients who are treated with vancomycin, and can partially be explained by determinants reported by clinical studies (80). For example, vancomycin is often administered to critically ill patients admitted to the ICU. In those patients the risk of developing nephrotoxicity is generally higher than for general ward patients as critically ill patients have various other risk factors for acute kidney injury (81). Observational cohort studies have shown that during vancomycin InI treatment serum trough concentrations of ≥ 15 mg/L are associated with a higher incidence of nephrotoxicity (66, 82-84). Data from other studies suggest that a daily dosage of >4000 mg could increase the risk of nephrotoxicity (85). In a meta-analysis, Van Hal et al. found an increased incidence of nephrotoxicity with an odds ratio (OR) of 2.67 for high trough serum concentrations (≥ 15 mg/L) compared to low trough serum concentrations (< 15 mg/L) ($p < 0.01$) (86). Lodise et al. confirmed in a retrospective study that nephrotoxicity is associated with higher trough levels ($C_{min} = 14.6 \pm 8.3$ mg/L versus $C_{min} = 9.6 \pm 5.1$ mg/L in patients with or without nephrotoxicity, respectively). No difference in AUC was found ($p = 0.11$) (87). Whether high trough levels are causative or a consequence of nephrotoxicity remains to be determined. Since vancomycin is mainly excreted through glomerular filtration, a decrease in the glomerular filtration rate (GFR) would result in higher vancomycin serum concentrations.

Among other risk factors that appear to be associated with an increased rate of nephrotoxicity are longer duration of vancomycin therapy and use of concomitant nephrotoxic medications (22). Nephrotoxic drugs include non-steroidal anti-inflammatory drugs (NSAIDs), inhibitors of the renin-angiotensin-aldosterone system (RAAS), loop diuretics, aminoglycosides, amphotericin B, colistin, immunosuppressant agents and intravenous contrast dye. Although nephrotoxicity resolves in 44-75% of the cases (86, 88), it is associated with a longer hospital length of stay and a higher mortality rate (89, 90).

For Col, data on risk factors for toxicity are limited compared to vancomycin InI. Ingram et al. conducted a retrospective cohort study in 102 outpatients and found a nephrotoxicity rate of 15.7%. Risk factors as determined in a multivariate analysis were hypertension, nephrotoxic drugs (more specifically aminoglycosides and loop diuretics) and a plateau concentration (C_{ss}) ≥ 28 mg/L (91). Spapen et al. also found that higher plateau concentrations ($C_{ss} > 30$ mg/L) as well as longer duration of treatment increased the incidence of nephrotoxicity in ICU patients treated with Col. In addition, patients who developed nephrotoxicity were more likely to have additional risk factors associated with impaired renal function, such as diabetes and hypotensive shock (90). The risk of nephrotoxicity may be decreased with Col when compared to InI. A recent meta-analysis compared nephrotoxicity of vancomycin Col versus InI and showed a relative risk of 0.6 (CI 0.4-0.9) (77). In addition, time to onset of nephrotoxicity appeared to be longer with Col compared to InI ($p=0.036$) in a study by Ingram et al. (92). Hutschala et al. found no difference in nephrotoxicity between InI and Col. However, a rise of 1 mg/L in vancomycin concentration induced a rise in serum creatinine of 0.04 mg/dL with InI versus a rise of 0.006 mg/dL with Col ($p < 0.001$) (93). In a retrospective cohort study, Saugel et al. showed that patients with InI of vancomycin were more likely to need renal replacement therapy (23% versus 7% in Col; $p = 0.007$). Of note, the targeted trough level for InI in this study was 5-10 mg/L, which is lower than is currently advised. The targeted plateau concentration for Col was 15-25 mg/L (94).

In conclusion

The IDSA guidelines recommend higher dosages in order to overcome the reduced susceptibility of MRSA to vancomycin (68). With higher dosing the risk of nephrotoxicity is likely to increase. Vancomycin Col has been suggested as an alternative approach to optimize the PK/PD of vancomycin. Current IDSA guidelines discourage the use of Col, since no benefit in terms of efficacy may be expected (68). The available literature suggests that in terms of efficacy vancomycin Col is neither superior nor inferior to vancomycin InI. Treatment failure and mortality were comparable in both treatment regimens (73, 74, 78, 95). Furthermore, current literature suggests the risk of developing nephrotoxicity tends to be lower in vancomycin Col compared to InI (77). Finally, continuous dosing of vancomycin, along with many other intravenously administered antibiotics, has shown to save health care costs and is easier to apply in home-care settings compared to intermittent dosing (74, 96).

Table 1. Comparative studies on clinical outcomes between intermittent and continuous infusion of vancomycin.

Study ID	Design	No. of patients		Setting/Condition	Vancomycin administration		Efficacy	Nephrotoxicity	Pharmacokinetics	
		Inl	Col		Inl	Col			Inl	Col
Boffi EI Amari, E. (2004) (21)	Ret. cohort	HD: 44 SD: 45		Osteomyelitis	HD = 40 mg/kg/d (Col and Inl); SD = 20 mg/kg/d (Inl)		Treatment failure higher with SD (OR = 7.5; CI 1.4-38.6) The best outcomes with regard to treatment failure and nephrotoxicity were found in HD- Col (log rank p = 0.02)	Total rate = 6.7%	C _{min} : SD = 10.0 ± 5.3 mg/L; HD = 24.4 ± 7.8 mg/L. Mean daily dose: SD = 1404 ± 593 mg; HD = 2389 ± 902 mg.	
Di Filippo, A. (1998) (37)	Ret. cohort	14	11	Critically ill patients (ICU)	Dose = 500 mg every 6h Dose = 83 mg/h; loading dose = 500 mg		Treatment with ND Col reduces SOFA score and WBC (p <0.05) but not length of stay ND		ND	ND
Hutschala, D. (2009) (35)	Ret. cohort	30	119	Critically ill patients (ICU) after cardiac surgery	C _{min} = 15 mg/L C _{ss} = 20-25 mg/L			Col: rate = 27.7%; Inl: rate = 36.7% (p = 0.3).	C _{min} = 17.0 ± 4.7 mg/L AUC _{24h} = 612 ± 213 mg.h.L ⁻¹ Target serum concentrations attained faster with Col (p <0.001)	C _{ss} = 25.0 ± 4.0 mg/L AUC _{24h} = 529 ± 98 mg.h.L ⁻¹
Ingram, PR. (2009) (34)	Matched ret. cohort	40	40	Outpatients			ND	Rate = 15.6% Later onset of nephrotoxicity with Col (p = 0.036).	C _{min} = 9.7 ± 5.0 mg/L C _{ss} = 13.6 ± 6.2 mg/L	

Study ID	Design	No. of patients		Setting/Condition	Vancomycin administration		Efficacy	Nephrotoxicity	Pharmacokinetics	
		Inl	Col		Inl	Col			Inl	Col
James, JK. (1996) (38)	Pros. cross-over	10	10	Assumed or confirmed gram-positive infection	Dose = 1000 mg every 12 h $C_{min} = 5\text{--}10\text{ mg/L}$	Dose = 80 mg/h $C_{ss} = 15\text{ mg/L}$	ND	ND	$C_{min} = 8.4 \pm 5.9\text{ mg/L}$ AUC/MIC = 550.0 ± 265.7 $C_{ss} = 20.2 \pm 11.1\text{ mg/L}$ AUC/MIC = 552.6 ± 373.4	
Saugel, B. (2013) (36)	Ret. cohort	75	164	ICU	$C_{min} = 5\text{--}10\text{ mg/L}$	$C_{ss} = 15\text{--}25\text{ mg/L}$	ND	Higher need for RRT in Inl vs. Col (23 vs. 7%; p = 0.007)	Median $C_{min} = 9.1 (5.0\text{--}15.7)$ Higher daily dose required with Col to attain target (p < 0.001). Less sample determinations needed with Col (p = 0.041)	Median C_{ss} = 19.8 (9.8–29.4)
Verral, AJ. (2012) (20)	Ret. cohort	56	188	Outpatients	$C_{min} = 15\text{--}20\text{ mg/L}$	$C_{ss} = 15\text{--}25\text{ mg/L}$	No difference in clinical failure rate (RR = 0.701; CI 0.432–1.136; p = 0.159)	ND	ND	ND
Vuagnat, A. (2004) (15)	Pros. cohort	21	23	Osteomyelitis	Dose = 40 mg/kg/d $C_{min} = 20\text{--}25\text{ mg/L}$	Dose = 40 mg/kg/d $C_{ss} = 20\text{--}25\text{ mg/L}$	No difference in cure rate in Inl vs. Col (77.8% vs. 94.4%; p = 0.25)	More treatment termination due to adverse effects in Inl vs. Col (42.9 % vs. 8.7%; p = 0.03)	Dose = 31.9 ± 12.5 mg/kg/d $C_{min} = 21.7 \pm 9.3\text{ mg/L}$ Dose = 33.9 ± 12.9 mg/kg/d $C_{ss} = 26.0 \pm 6.1\text{ mg/L}$	Lower variability of vancomycin concentrations in Col (p = 0.001)

Study ID	Design	No. of patients		Setting/Condition	Vancomycin administration		Efficacy	Nephrotoxicity	Pharmacokinetics	
		Inl	Col		Inl	Col			Inl	Col
Wysocki, M. (2001) (14)	Pros. randomized trial	58	61	ICU patients with severe MRS infections	Dose = 30 mg/kg/d C_{min} = 10-15 mg/L	Dose = 30 mg/kg/d C_{ss} = 20-25 mg/L Loading dose = 15 mg/kg.	Treatment failure at end of treatment was comparable (Col = 21% vs. Inl = 19%; power = 23%)	Rate = 20%. Comparable rates in Inl and Col (p = 0.64)	Success: C_{min} = 14 ± 5 mg/L ^a Failure: C_{min} = 21 ± 9 mg/L Success: C_{ss} = 23 ± 4 mg/L Failure: C_{ss} = 25 ± 5 mg/L AUC_{24h} = 577 ± 120 mg.h.L ⁻¹ Lower variability of AUC_{24h} with Col (p = 0.026); Targeted concentrations attained faster with Col (p = 0.03).	

Abbreviations: AUC_{24h} , area under the concentration-time curve; CI, 95%-confidence interval; C_{min} , trough serum concentration; Col, continuous infusion; C_{ss} , plateau serum concentration; HD, high dose; ICU, intensive care unit; Inl, intermittent infusion; MIC, minimum inhibitory concentration; MRS, methicillin resistant *Staphylococcus*; ND, not determined; OR, odds ratio; pros., prospective; ret., retrospective; RR, relative risk; RRT, renal replacement therapy; SD, standard dose; SOFA, Sequential Organ Failure Assessment; WBC, white blood cell count.

^a p <0.05 for treatment success vs. failure; ^b p <0.0001 for Inl vs. Col

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Aim of this thesis

The aim of this thesis is to explore and establish new dosing strategies for intravenous aminoglycosides and vancomycin therapies with an optimal balance between efficacy and toxicity.

Outline of this thesis

Chapter 2. Aminoglycosides

Chapter 2.1 describes a randomized clinical trial investigating the effects of the time of administration on tobramycin pharmacokinetics and nephrotoxicity in adult cystic fibrosis patients.

Chapter 2.2 reports retrospective data on the chronopharmacology of aminoglycosides in general ward and critically ill patients.

Chapter 2.3 compares the nephrotoxicity of tobramycin and gentamicin in a population of clinically infected patients.

Chapter 2.4 provides information on a prospectively tested new gentamicin dosing algorithm in neonates admitted to a special care nursery.

Chapter 3. Vancomycin

Chapter 3.1 reviews and compares vancomycin exposure variabilities of continuous and intermittent dosing regimens in adult hospitalized patients.

In Chapter 3.2 the exposure variability of vancomycin in a population of critically ill patients on continuous or intermittent infusion is studied.

Chapter 3.3 shows a method to design and validate a dosing regimen of continuously dosed vancomycin in a mixed general ward and intensive care unit population using historical within-population data.

Chapter 4 summarizes and discusses the results and conclusions of the conducted studies. Moreover, their impact on clinical practice is addressed and suggestions for future studies are provided with respect to aminoglycosides and vancomycin therapies in clinically infected patient populations.

Chapter 2

Chapter 2.1

The Effect of the Circadian Rhythm on the Pharmacokinetics of Once Daily Tobramycin in Cystic Fibrosis Patients

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In the era of multiple daily dosing of systemic aminoglycosides a circadian rhythm in the clearance of these vital antibiotics has been demonstrated in animals and healthy volunteers. Over the past two decades, however, once daily dosing (ODD) regimens proved to be less nephrotoxic and therefore ODD protocols were adopted by centers worldwide for most indications requiring treatment with an aminoglycoside. In this study the effect of the time of administration on the pharmacokinetics of once-daily tobramycin in adult CF patients was investigated. In this open randomized study CF patients were administered intravenous tobramycin at 8:00 or at 22:00. Pharmacokinetic parameter values and biochemical parameters of renal function were compared between groups. Twenty-five patients were included. Mean weight-corrected clearances of tobramycin were 1.46 vs. 1.43 mL/min*kg ($p=0.50$) and mean volumes of distribution were 0.25 vs. 0.27 L/kg ($p=0.54$) for the 8:00 and 22:00 groups, respectively. Furthermore, no significant differences were detected in the changes in estimated creatinine clearance or tobramycin clearance from day 1 to day 8 in the 8:00 group or in the 22:00 group indicating there was no decline in clearance over time. At day 8 of therapy the rise in serum BUN in the 22:00 group was significantly higher compared to the 8:00 group (1.8 vs. 0.2 mmol/L, $p=0.015$). In conclusion, the time of administration did not affect tobramycin pharmacokinetics the adult CF population studied. The rise in serum BUN in the 22:00 group deserves further exploration.

Introduction

Cystic fibrosis is an incurable, genetically inherited disease that causes changes in the ion transport mechanisms in different parts of the body (lungs, skin). Although the disease affects several body systems it is respiratory decline that leads to fatality (1). Patients with CF are prone to acquire pulmonary infections with *Pseudomonas aeruginosa* and other micro organisms which are associated with irreversible pulmonary damage leading to a reduced life expectancy (2). Intravenous aminoglycosides, in combination with beta-lactam antibiotics, are a cornerstone for the treatment of exacerbations of chronic pulmonary infections (3) caused by *Pseudomonas aeruginosa*.

However, frequent use of intravenous aminoglycosides is associated with nephro- and ototoxicity and in some cases with vestibulotoxicity

(4, 5). Repeated or prolonged dosing may cause damage to the proximal tubuli, resulting in renal impairment, which affects up to 42 % of adult CF patients (6). As the life expectancy of CF patients is extending as a consequence of improved treatment, cumulative toxicity caused by aminoglycosides is nowadays more relevant than in the past. The TOPIC study demonstrated that once-daily dosing (ODD) of tobramycin is at least as effective and may be less toxic compared to multiple daily dosing (MDD) in CF patients (7). Hence, ODD of tobramycin has been adopted by a great number of cystic fibrosis treatment centers worldwide. Since aminoglycosides are almost completely excreted into the urine by glomerular filtration, their kinetics may fluctuate with diurnal changes in renal function as previously demonstrated in animal experiments (8, 9), and in healthy volunteers (8, 10). Consequently, if an effect of the administration on the pharmacokinetics and toxicity of tobramycin exists in CF patients, then clinically it will be more relevant in ODD compared to MDD regimens. Human data on a circadian rhythm in pharmacokinetics are however conflicting and whether or not an association is found primarily seem depend on dosing regimen and the specific population investigated (11-13). A post-hoc analysis of the TOPIC study performed denoted a peculiar difference of 30 % in renal elimination between the ODD group and the three times daily group (14). The authors relate this to the fact that the majority of patients in the ODD group (53 of 71) received their active dose in the evening. Therefore, most of the systemic drug exposure occurred during the resting period when renal function is relatively decreased. In this study a decreased renal elimination for the pediatric as well as the adult once daily group was observed. One could hypothesize this to be the result of a circadian rhythm in drug clearance caused by either active and resting periods or dietary protein intake or both. To date, the effect of diurnal changes on aminoglycoside pharmacokinetics in CF patients has not been studied prospectively. Ultimately, it is important that CF patients are exposed to aminoglycosides at the time period of highest clearance to prevent acute adverse events and events resulting from accumulated toxicity without compromising efficacy (15, 16). The present randomized clinical study was designed to evaluate diurnal changes in tobramycin pharmacokinetics and renal function.

Patients & Methods

In this open randomised clinical trial ((Dutch Trial Register number: NTR3309) patients with cystic fibrosis, who were treated with intravenous tobramycin for a pulmonary exacerbation at the Haga Teaching Hospital (The Hague, The Netherlands), were eligible for inclusion. The study was approved by the local Medical Ethics Committee. The study was powered to detect a 25% difference in tobramycin exposure quantified by the area-under-the-concentration-time-curve (AUC), since regulatory authorities consider 80-125 % limits to be a clinically relevant cut-offs for bioequivalence. An informed consent was obtained from all participants. The inclusion criteria were: age older than 18, a diagnosis of cystic fibrosis (i.e., sweat chloride > 60 mmol/l or a genotype known to cause the disease), chronic infection with *Pseudomonas aeruginosa* with the most recently isolated organism showing sensitivity to tobramycin and pulmonary exacerbation as defined by Fuchs and colleagues (17). Patients who were on other nephrotoxic drugs or were allergic to aminoglycosides, had granulocytopenia ($<1.0 \times 10^9/L$), were pregnant, had estimated GFR < 40 ml/min or pre-existing hearing impairment, were excluded. After inclusion participants were randomly assigned to receive tobramycin at either 8:00 or at 22:00. Total daily doses equivalent to those previously received by patients during routine treatment were used. If a patient had not previously been treated with tobramycin, a dose of 10 mg/kgTBW/day was administered as a 30 min infusion (TBW is total body weight). Tobramycin concentrations were drawn one hour after start of the infusion (peak level) and between 6-8 h after start of the infusion (mid level). Dosage regimes were altered, if serum concentrations were outside target values (peak: 25-30 mg/L; extrapolated trough <0.5 mg/L). Samples were analyzed using a fluorescence polarization immunoassay (AxSym, Abbott, Illinois, U.S.A.). An assay error of standard deviation = $0.25 + 0.15 \cdot C$ (where C is the concentration measured) was used in the pharmacokinetic analysis. Tobramycin pharmacokinetic parameters at days 1 and 8 were calculated for each patient using a previously described one-compartment model with linear pharmacokinetics (18) using the MW/Pharm pharmacokinetic software package version 3.60 (19).

Pharmacokinetic parameters that were determined included apparent volume of distribution (Vd), tobramycin clearance (Cl_{Tob}) and area-under-the-concentration-time-curve (AUC).

Serum creatinine concentrations, 24h-urine collections and blood urine nitrogen (BUN) were taken in the morning on days 1 and 8 of tobramycin treatment. Serum and urinary creatinine were analyzed using a colorimetric method and BUN by a kinetic enzymatic assay (DxC, Beckman Coulter Inc., Brea, CA, USA). Subsequently, renal function was quantified at baseline and at day 8 using serum creatinine concentrations to estimate the creatinine clearance using the formula proposed by Jelliffe (20), which approximates the glomerular filtration rate (eGFR). Glomerular filtration was also measured GFR (mGFR) using 24h-urine collections. The effect of the time period of administration of tobramycin on renal function was assessed by the change in serum BUN, eGFR and mGFR at day 8 compared to baseline.

Statistical analysis

A Kolmogorov-Smirnov test was used to identify normal distribution of data. Means and standard deviations of continuous variables are presented for normal distributions. Otherwise, medians and ranges were reported. A Student's T-test was used to determine statistical differences among normally distributed data. If data were not normally distributed, a Mann-Whitney-U test was performed. A Chi-square test was used to compare dichotomous variables between groups. Vd, and Cl_{Tob} were analyzed after correction for TBW and AUCs were normalized to an administered dose of 500 mg tobramycin (500mgTOB). Tests were two-tailed and a p-value of <0.05 was considered statistically significant. All Statistical analysis were performed using GraphPad Prism version 5.03 for Window (GraphPadSoftware, San Diego, CA, USA).

Results

Study participants

Twenty-five patients, who received at least one dose of tobramycin between July 2008 and May 2011, were enrolled in the study. No significant differences in baseline characteristics between the 8:00 and 22:00 groups were detected (Table 1).

Table 1. Patient characteristics

	8:00	22:00	p-value
No. patients	12	13	
age (yr)	30 (12)	30 (11)	0.42 ^a
male gender (%)	63.6	72.7	0.87 ^c
weight (kg)	60 (21)	61 (22)	1.0 ^a
serum creatinine (nmol/mL)	66 (55-97)	65 (33-91)	0.46 ^b
eGFR (mL/min)	118 (67-132)	112 (82-175)	0.60 ^b
cGFR (ml/min)	102 (75-145)	108 (55-185)	1.0 ^b
BUN (nmol/mL)	4.2 (2.2-7.7)	4.0 (2.3-6.3)	0.27 ^b

Baseline characteristics of 8:00 and 22:00 groups are shown. Dichotomous variables were expressed as percentages and continuous variables as means or medians with standard deviations or ranges abetween parentheses. Abbreviations are eGFR, estimated glomerular filtration rate; BUN, blood urea nitrogen. a Student's T-test, b Mann-Whitney test, c Chi-square test

Treatment was either stopped or continued at home after discharge for 2 patients during the first week of tobramycin therapy in both study arms. Tobramycin dose was adjusted at day 1 in 3 patients in the 8:00 group and in 5 patients in the 22:00 group. The median cumulative doses over 7 days of tobramycin therapy were 3980 and 3840 mg (p=0.90) for the 8:00 and 22:00 groups respectively. Patients did not receive any other nephrotoxic medication from day 1 to day 8.

Pharmacokinetics

No significant differences in pharmacokinetic parameters were detected between the 8:00 and 22:00 groups (Table 2 and Figure 1). Furthermore, no significant difference was detected in the change in CI_{Tob} between day 1 and day 8 in the 8:00 group (p=0.58) or in the 22:00 group (p=0.67) indicating there was no decline in clearance over time.

Parameters of renal function

No significant difference in the change in eGFR from baseline was found between the 8:00 and 22:00 groups (3.8 vs. 10.0 mL/h respectively, $p = 1.0$) (Figure 2). The rise in serum BUN in the 22:00 group was statistically significant compared to the 8:00 group (1.8 vs. 0.2 mmol/L respectively, $p=0.015$) and serum BUN at day 8 was significantly higher compared to baseline in the 22:00 group ($p=0.014$), but not in the 8:00 group ($p=0.79$). The change in mGFR could not be analyzed due to incomplete 24h-urine sampling in 7 of 25 participants at day 8.

Table 2. Dosing and pharmacokinetic parameters

	time of administration		Rate ratio	p-value
	Day 1			
	8:00	22:00		
Dose (mg)	525 (420-600)	500 (400-600)	n.a.	0.35 ^b
Cmax (mg/L)	35.0 (3.9)	30.4 (4.1)	1.15	0.53 ^a
AUC (mg*h/L) ^c	87 (26)	97 (29)	0.90	0.21 ^a
Vd (L/kg)	0.25 (0.08)	0.27 (0.09)	0.93	0.54 ^a
Cl_TOB (mL/min/kg)	1.46 (0.44)	1.43 (0.52)	1.02	0.50 ^a
	Day 7			
	8:00	22:00		p-value
Dose (mg)	500 (500-720)	550 (400-640)	n.a.	0.39 ^b
Cmax (mg/L)	32.3 (3.6)	33.4 (3.2)	0.97	0.68 ^a
AUC (mg*h/L) ^c	101 (32)	88 (26)	1.15	0.34 ^a
Cl_TOB (mL/min/kg)	1.42 (0.52)	1.54 (0.63)	0.92	0.87 ^a
Vd (L/kg)	0.25 (0.09)	0.27 (0.11)	0.93	0.19 ^a

Dosing and pharmacokinetic parameters are displayed as means or medians with standard deviations or ranges between parentheses, respectively. Abbreviations are: Cl_TOB, clearance of tobramycin; Vd, volume of distribution and AUC, area-under-the-curve.

a Student's T-test

b Mann-Whitney test

c Normalized to a tobramycin dose of 500 mg

The graphs are box-and-whisker plots of clearance of tobramycin (Cl_{TOB} , fig 1A), volume of distribution (V_d , fig 1B) and area-under-the-curve (AUC , fig 1C). The line in the middle of each box represents the median; the box extends from the 25th to 75th percentile (interquartile range); and whiskers extend to 1.5 times the interquartile range, rolled back to where data are present.

The graph is box-and-whisker plot of the glomerular filtration rate (eGFR) and serum blood urea nitrogen (BUN). The line in the middle of each box represents the median; the box extends from the 25th to 75th percentile (interquartile range); and whiskers extend to 1.5 times the interquartile range, rolled back to where data are present.

Discussion

No influence of the time period of administration on tobramycin pharmacokinetics in a hospitalized adult CF population was found in this study. However, we found a rise in serum BUN after one week of therapy in the 22:00 group which is statistically significant compared to the 8:00 group and deserves further exploration.

Since at our clinic only patients in a clinically poor condition suffering from a pulmonary exacerbation are admitted to hospital, the majority is treated at home. Perhaps, the circadian rhythm in ambulatory treated CF populations is preserved during a pulmonary exacerbation. This may select patients who have a less pronounced circadian rhythm as a result of an altered status of the immune system (23). In fact, Vinks et al. found that early-morning serum concentrations of ceftazidime were significantly higher than the afternoon levels in CF patients who were

treated at home with a continuous infusion (24). Unfortunately, the presence or absence of a normal circadian rhythm was not assessed by measuring melatonin levels for instance. Finally, the contribution of the effect of the circadian may be limited relative to the effect in non-CF patients, since renal clearance of tobramycin is already very rapid in CF patients compared to non-CF patients (18).

This is the first study to examine the effect of diurnal changes on the pharmacokinetics of aminoglycosides in adult CF patients. The pharmacokinetic results were not in line with the results from the posthoc analysis of the TOPIC study (16) and were unexpected as 3 out of 5 studies in hospitalized patients indicate there is an influence of the circadian rhythm on renal clearance of aminoglycosides (11-13). However, the previously mentioned three positive studies were performed in an era where aminoglycosides were multiple daily or even continuously dosed and investigated amikacin, kanamycin and gentamicin that are more likely to accumulate compared to tobramycin (7, 25). Furthermore, with an actual aminoglycoside blood concentration of 0.5 mg/mL, most assays report a value somewhere between 0 and 1.0 mg/mL. Use of an inaccurate value may have major consequences for the calculation of clearance. For optimal estimation of tobramycin pharmacokinetics a peak level and a second level at 1.44 half-lives after the end of the administration should be drawn (i.e. between 1.5-9 h for an adult CF patient) (26).

In spite of lack of an observed effect of the time of administration on the pharmacokinetics of tobramycin, we found a significant rise in serum BUN in the 22:00 group compared to the 8:00 group. Although a rise in serum BUN of 1.8 nmol/mL may not be considered clinically relevant, it is rather unexpected since eGFR and exposure in terms of AUC were similar in the 8:00 and 22:00 groups. Perhaps, the former is caused by a disturbance of the diurnal pattern of serum BUN by the nightly exposure to tobramycin, resulting serum BUN to peak in the morning instead of in the evening (27). Furthermore, polycationic aminoglycosides, like tobramycin, can alter ultrastructure and glomerular permeability, tubular reabsorption and intracellular digestion of proteins (27). Finally, a 'false' rise in serum BUN or any other biomarker for that matter caused by interference of tobramycin with the analytical assay of BUN should not be overlooked. Regarding the latter, no effect was detected of tobramycin on BUN concentrations (data not shown). As a result of the known circadian rhythm of biochemical parameters, ideally sampling of these parameters should be performed at several

time points during the day or as a 24h-urine collection. Unfortunately, 24h-urine sampling failed in too many patients in our study to perform a statistical analysis of change in mGFR. Future studies addressing this issue should preferably investigate longer periods of exposure to tobramycin (28), and focus on obtaining highly sensitive as well as thoroughly characterized biomarkers for renal toxicity, such as cystatin C, β -2-microglobulin and N-acetyl- β -D-glucosaminidase (29, 30).

Acknowledgements

The authors are grateful to Prof Dr J.H. Proost from the department of Pharmacokinetics, University of Groningen, The Netherlands for his help with the pharmacokinetic analysis. Furthermore, we would like to thank Prof Dr A. Smyth, Department of Child Health, School of Clinical Sciences & Nottingham Respiratory BRU, University of Nottingham, United Kingdom for his comments on an earlier version of the manuscript.

Declaration of interest statement

The authors report no conflicts of interest.

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Chapter 2.2

Chronopharmacokinetics of Once Daily Dosed Aminoglycosides in Clinically Infected Patients

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Abstract

Clinically infected patients treated with aminoglycosides are at risk of developing nephrotoxicity. It has been suggested that the pharmacokinetics of aminoglycosides in humans follows a circadian rhythm. Therefore, the time period of administration could have important clinical implications in once daily dosing regimens primarily with regard to the incidence of nephrotoxicity. In this retrospective cohort study, the effect of the clock time of administration on aminoglycoside exposure and the incidence of nephrotoxicity in a large population of clinically infected patients was investigated. Patients treated with intravenous tobramycin or gentamicin were eligible for inclusion. Patients were divided into three groups by time of administration: morning, afternoon and night. Pharmacokinetic parameters and the incidences of nephrotoxicity were compared between the morning, afternoon and evening groups. 310 general ward and 411 intensive care unit patients were included. No significant differences were found in patient characteristics between the morning, afternoon and night groups. The time period of administration did not affect aminoglycoside exposure ($p=0.97$) or the incidence of nephrotoxicity ($p=0.47$). This large retrospective study shows that the time period of administration had no effect on the pharmacokinetics or nephrotoxicity of ODD aminoglycosides in hospitalized patients. Consequently, we advise aminoglycosides to be administered as soon as possible in case of (suspected) severe hospital-acquired infections and subsequent dosages to be based on therapeutic drug monitoring.

Introduction

Aminoglycoside antibiotics remain an important group of antimicrobial agents in the treatment of serious infections by gram-negative organisms (1). Although they are highly effective in killing these organisms, exposure to intravenous aminoglycosides comes with an increased risk of auditory and renal toxicity (2). All hospitalized patients, especially critically ill patients with a low renal clearance of aminoglycosides are at high risk of developing nephrotoxicity (3, 4). In an attempt to reduce the nephrotoxic side effects without compromising efficacy, once-daily dosing regimens of aminoglycosides have shown to be superior to conventional multiple daily dosing regimens (5).

Nowadays, intravenous aminoglycosides are mainly dosed once daily worldwide (6). Since a circadian rhythm of aminoglycoside pharmacokinetics has been observed in animal experiments (7-11) as well as in human studies involving multiple daily dosing protocols (8, 12, 13), the clearance of aminoglycosides is expected to be higher in the active period versus the resting period caused by an increase in glomerular filtration rate. As a result, the time of administration in patients receiving their drug with 24-hour intervals could have important implications with regards to aminoglycoside clearance and thus exposure. Nevertheless, human data involving the effect of the circadian rhythm on pharmacokinetics and toxicity of once daily dosed (ODD) aminoglycosides are inconsistent (14,15). To date, the effect of diurnal changes on key pharmacokinetic parameters of aminoglycoside exposure has never been extensively studied in a large cohort of patients with serious infections.

Aim of the study

To examine the effect of time period of administration on aminoglycoside exposure and the incidence of nephrotoxicity in a large population of clinically infected patients.

Ethical approval

A waiver for consent was provided by the local medical ethics committee.

Methods

In this retrospective cohort study data were extracted from patients' clinical records and pharmacokinetic database. All general ward and intensive care unit patients of 18 years or older who were admitted hospital between January 2006 and December 2013 and were treated with intravenous tobramycin or gentamicin were included. Furthermore, an estimated glomerular filtration rate calculated with the MDRD formula (16) higher than 25 mL/min and a once daily dose of 4-6 mg/kg were required. Finally, two tobramycin concentrations drawn after the first or second infusion were mandatory for inclusion. For the analysis of nephrotoxicity in the general ward population, the treatment duration had to be two days or longer. Furthermore, a minimum of two serum

creatinine concentrations was required; one drawn between 48 hours before and 12 hours after initiation of therapy. If two or more concentrations were available during this time window the concentration drawn closest to the time of initiation of aminoglycoside therapy was selected as the baseline creatinine concentration. The second 'post-treatment' concentration was drawn within 48 hours after initiation and 48 hours after cessation of therapy was mandatory for inclusion. If two 'post-treatment' were available in this period the highest creatinine concentration was appointed as the 'post-treatment' concentration. The delta serum creatinine (Δ Scr) was derived by subtracting the 'post-treatment' with the baseline creatinine. Nephrotoxicity was defined as a Δ Scr of at least 45 nmol/mL (0.5 mg/dL) from baseline (15). For the analysis of nephrotoxicity in the general ward population, a minimal treatment duration of 2 days was required for inclusion. According to the monitoring protocol, a peak concentration and second concentration were drawn 0.5-1 h and 6-14 h after end of the first or second infusion, respectively. Aminoglycoside dosing was guided by drug concentration monitoring targeting peak concentrations of 15-20 mg/L, and estimated trough concentrations <0.5 mg/L. It was a priori decided to analyze the pharmacokinetics separately for general ward and intensive care unit populations and to analyze nephrotoxicity only in the general ward population because the kidney function of critically ill patients admitted to the intensive care unit has an inherently large variability and many artifacts occur due to cardiovascular management (17). Patients were divided into three groups by time of administration: 5:00-12:55 (morning), 13:00-20:55 (afternoon) and 21:00-4:55 (night). Pharmacokinetic parameters were calculated using pharmacokinetic software (MW\ Pharm 3.80, MediWare, Zuidhoorn, The Netherlands) and population pharmacokinetic models (18).

Statistical analysis

A Kolmogorov-Smirnov test was used to examine normal distribution of the data. Means and standard deviations of continuous variables are presented for normal distributions. Otherwise, medians and ranges were reported. One-way ANOVA with bonferroni post hoc tests were used to determine statistical differences among normally distributed groups. If data were not normally distributed, a Kruskal-Wallis non-parametric method was applied with Mann-Whitney-U post hoc tests

including correction for multiple testing. A Chi-square test was used to compare dichotomous variables between groups. All Statistical analysis were performed using GraphPad Prism version 5.03 for Windows (GraphPad Software, San Diego, CA, USA). A p-value of <0.05 was considered to be statistically significant and in each instance a 2-sided test was carried out.

Results

A total of 1452 general ward and 843 intensive care unit patients were selected from the database. Approximately half of the selected general ward and intensive care unit patients were excluded as a result of missing pharmacokinetic data. In the general ward population, subsequently again half of the remaining patients were excluded from the nephrotoxicity analysis because of missing creatinine values or a treatment duration shorter than two days. 310 general ward and 411 intensive care unit patients were eligible for inclusion in the morning, afternoon and night groups. No significant differences were found in patient characteristics between the three groups within both populations (Table 1).

Although a trend toward a higher volume of distribution in the afternoon group was seen in the general ward population compared to the morning or night groups (0.28, 0.25 and 0.24 L/kg respectively, $p=0.06$), tobramycin clearance and area-under-the-curve did not differ significantly between the morning, afternoon and night groups in both populations (Table 2). Finally, no significant differences in delta serum creatinine ($p=0.11$) or in percentages of nephrotoxicity ($p=0.47$) were found between the morning, afternoon and night groups of the general ward population (Table 2).

Table 1. Patient characteristics

general ward	Time of administration			p-value
	Morning (n=88)	Afternoon (n=108)	Night (n=114)	
age (years)	63 (34)	62 (27)	57 (31)	0.56 ^a
weight (kg)	70 (29)	71 (27)	70 (26)	0.33 ^a
% of males	42	48	56	0.10 ^c
% of patients on TOB	21	27	19	0.23 ^c
eGFR (ml/min)	80 (26-191)	88 (26-189)	89 (26-185)	0.27 ^b
intensive care unit	Morning (n=124)	Afternoon (n=145)	Night (n=142)	p-value
age (years)	68 (32)	65 (28)	68 (31)	0.14 ^a
weight (kg)	72 (28)	75 (26)	75 (24)	0.48 ^a
% of males	64	65	69	0.47 ^c
% of patients on TOB	48	43	39	0.35 ^c
eGFR (ml/min)	55 (25-167)	64 (25-183)	57 (25-167)	0.41 ^b

Dichotomous variables are displayed as percentages. Continuous variables are presented as means $\pm$ standard deviations or medians with ranges in parentheses (min-max). Abbreviations are: eGFR, estimated glomerular filtration rate; SAPS, simplified acute physiology score; TOB, tobramycin.

a ANOVA

b Kruskal-Wallis test

c Chi-square test

Table 2. Dosing and pharmacokinetic parameters

General Ward	Morning (n=88)	Afternoon (n=108)	Night (n=114)	p-value
Dose (mg)	350 (180-500)	360 (220-600)	350 (240-650)	0.48 ^b
Treatment duration (days)	5 (3-18)	5 (3-22)	5 (3-21)	0.98 ^b
CL (L/h)	4.4 (1.7)	4.6 (1.8)	4.7 (1.7)	0.41 ^a
Vd (L/kg)	0.25 (0.08)	0.28 (0.09)	0.24 (0.08)	0.06 ^a
AUC (mg*h/L)	135 (47)	139 (48)	145 (51)	0.97 ^a
Delta Scr (nmol/mL)	6 (-40-167)	0 (-11-323)	-1 (-56-467)	0.11 ^b
Nephrotoxicity (%)	18	12	10	0.47 ^c
ICU	Morning (n=124)	Afternoon (n=145)	Night (n=142)	p-value
Dose (mg)	350 (250-675)	360 (240-600)	360 (150-750)	0.36 ^b
Treatment duration (days)	4 (3-12)	4 (3-13)	5 (3-17)	0.72 ^b
CL (L/h)	1.9 (0.06)	2.1 (0.08)	2.1 (0.07)	0.19 ^a
Vd (L/kg)	0.33 (0.11)	0.32 (0.10)	0.33 (0.12)	0.85 ^a
AUC (mg*h/L)	195 (71)	183 (75)	173 (67)	0.86 ^a

Continuous variables are presented as means ± standard deviations or medians with ranges in parentheses (min-max). Abbreviations are: eGFR, estimated glomerular filtration rate; AUC, area under the concentration time curve; CL, clearance; Scr, serum creatinine; Vd, volume of distribution, ΔScr, median delta serum creatinine concentration

a ANOVA

b Kruskal-Wallis test

c Chi-square test

Discussion

Since most previous studies did find an effect of the circadian rhythm on the pharmacokinetics of aminoglycosides, our results intuitively seem unexpected. Nevertheless, an in-depth review of the available evidence demonstrates that most studies were performed in relatively small populations of animals or (young) healthy volunteers, primarily investigated multiple daily or even continuously dosed aminoglycosides, and had suboptimal pharmacokinetic sampling protocols with only peak and trough concentration monitoring and/or reported on amikacin and kanamycin that are more likely to accumulate (7-13, 19-22). In addition and perhaps most eminent, therapeutic drug monitoring in the majority of studies was performed after 3 days of therapy at 'steady state'. At that moment in time the signs of clinical infection will have resolved in a large proportion of patients and aminoglycosides are then withdrawn in current clinical practice. It is thus critical to collect pharmacokinetic data during the first two days of therapy when the patient still shows signs of a clinical infection, since sepsis and an altered status of the immune system disrupt the circadian rhythm (23, 24). Our study investigated a large population of clinically infected patients, who received their aminoglycosides ODD with optimal pharmacokinetic sampling during the first or the second administration.

Caution needs to be exercised when interpreting the results of any retrospective cohort study because of errors due to confounding and selection bias. However, baseline patient characteristics showed no significant differences between groups and the proportion of excluded patients per exclusion criterion did not differ significantly between patients who were administered aminoglycosides in the morning, afternoon and night (data not shown). Furthermore, coadministration of nephrotoxic medication was not included in this analysis, but a correlation between the use of e.g. furosemide or other nephrotoxic antibiotics and the time of administration of aminoglycosides is highly unlikely.

As mentioned, the results obtained from studies examining the effect of the circadian rhythm on the pharmacokinetics of aminoglycosides in patient populations are conflicting (15, 25-28), but can be explained by the same conveyable causes for the vast majority. The three 'positive' studies all involved multiple daily dosing protocols of 'earlier', more accumulating aminoglycosides (e.g. kanamycin and amikacin). In contrast, the two 'negative' studies reported on once or twice daily dosed

aminoglycosides that show relatively low tissue accumulation (e.g. tobramycin, netilmicin and gentamicin). A randomized controlled trial by Fauville et al., in which netilmicin (4.5 mg/kg of lean body weight) was administered every 24h at 10 a.m. or 10 p.m. to 23 ill febrile patients with severe infection, showed that the time of administration had no influence on netilmicin concentrations in serum nor on pharmacokinetic parameters in these patients (27). The second was a prospective cohort study by Prins et al., who investigated the influence of drug administration time on serum drug concentrations and the incidence of nephrotoxicity in 179 patients with serious infections treated with gentamicin or tobramycin once daily. The authors did not find statistically significant differences in trough or peak concentrations between the time periods of administration. Nevertheless, the incidence of nephrotoxicity was significantly higher when the aminoglycosides were administered to a combined general ward and intensive care unit population during the resting period (midnight to 7:30 AM; $p = 0.004$). In our opinion, data on pharmacokinetics and nephrotoxicity of aminoglycosides in general ward and intensive care unit populations should be analyzed separately since admittance to the intensive care unit is a known risk factor for acute kidney injury (29, 30). Moreover, the study by Prins et al. demonstrated that intensive care unit patients are more 'at risk' to receive their aminoglycosides during the night (77 vs. 44 %) (15). Admittance to the intensive care unit is thereby a confounder by definition, since it correlates with both the dependent (i.e. nephrotoxicity) and the independent variable (i.e. time of administration). Furthermore, the authors had to statistically correct for not only the higher prevalence of intensive care unit admittance but also for significantly poorer baseline creatinine clearance (84 vs. 100 ml/min, $p < 0.05$) in the resting compared to the active period. All of the above are established risk factors of acute kidney injury (29, 30). Altogether, it is debatable whether statistical correction is a proper tool to adjust for these imbalances in patient characteristics.

Our data show a trend of a higher volume of distribution in the afternoon group of the general ward population but not in the intensive care unit population. This is probably caused by the discrepancy in the daily fluid management of a general ward patient compared to an intensive care unit patient. Whereas the latter will receive intravenous fluids continuously contributing to a relatively stable volume of distribution, a general ward patient will gain fluids during the day causing the volume of distribution of hydrophilic aminoglycosides to peak in the afternoon.

The presented pharmacokinetic data are in accordance with the data reported by Fauvalle et al.(27) and are further supported by the unpublished results of two recently performed randomized clinical trials in adult and pediatric cystic fibrosis patients suffering from pulmonary exacerbation that also found no effect of the circadian rhythm on the pharmacokinetics of once daily dosed tobramycin in cystic fibrosis patients (submitted for publication).

Conclusion

Currently it is widely believed that administration of aminoglycosides in the morning will result in a reduction in nephrotoxicity and many (international) antibiotic guidelines advice to administer aminoglycosides in the morning. This large retrospective study, however, shows no administration-time-related differences in aminoglycoside exposure or nephrotoxicity in clinically infected patients. It is therefore unlikely that administration during the resting period will result in a higher incidence of renal toxicity in these patients. We recommend aminoglycosides to be administered as soon as possible in case of (suspected) severe hospital-acquired pneumonia or sepsis to reduce the pen-to-needle time in antibiotics, which has shown to be effective in preventing death (31). Subsequent dosages should be based on therapeutic drug monitoring to optimize the efficacy/toxicity balance.

Funding

No funding was received for this study.

Competing interests

The authors declare that they have no competing interests.

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Chapter 2.3

Once Daily Dosed Gentamicin is more Nephrotoxic than Once Daily Dosed Tobramycin in Clinically Infected Patients

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Adapted from *J Antimicrob Chemother.* 2014 Sep;69(9):2581-3.

Abstract

Randomized clinical trials and meta-analyses have shown that multiple daily dosed (MDD) tobramycin is less nephrotoxic than MDD gentamicin. However, no prior studies have compared nephrotoxicity of tobramycin versus gentamicin when administered as nowadays more common once daily dosing (ODD) regimens. Hence, the present study aimed to investigate the nephrotoxicity of ODD gentamicin and tobramycin. A retrospective cohort analysis was performed on data of adult patients, treated with a once daily dose of 5 mg/kg gentamicin or tobramycin for three days or longer. As local treatment protocol switched from gentamicin to tobramycin in February 2010, gentamicin treated patients were selected from January 2008 to February 2010, while tobramycin treated patients were selected from March 2010 to December 2012. Information on gender, age, kidney function, dose, treatment duration, and aminoglycoside serum concentrations was collected. A multivariate logistic regression analysis was performed to identify covariates predictive of nephrotoxicity. Of the 2889 patients who were eligible, 202 gentamicin and 180 tobramycin met the inclusion criteria and were evaluated. Baseline characteristics did not differ between both groups. Nephrotoxicity developed in 43 of 202 (21.2%) of gentamicin and 19 of 180 (10.6%) of tobramycin treated patients (odds ratio (OR) 2.4, 95% confidence interval (95% CI) 1.2-4.7). Furthermore, an area-under-the-curve (AUC) of the first administration >150 mg*h/L was independently predictive of developing nephrotoxicity (OR 2.2, 95% CI 1.1-5.0). ODD tobramycin was less nephrotoxic than ODD gentamicin. Moreover, early AUC monitoring seems warranted to identify patients at high risk of developing nephrotoxicity.

Introduction

During the late 1970s, the commercial launch of broadspectrum antibiotics with fewer side effects, such as fluoroquinolones, carbapenems, cephalosporins, and β -lactam/ β -lactamase inhibitor combinations, led to a declined clinical application of aminoglycosides (1). Unfortunately, over the past forty years, a number of resistance mechanisms have emerged in gram-negative pathogens that pose a serious threat to our common antibiotic armamentarium.

As the pharmaceutical pipeline wanes drugs that are active against multi-drug resistant gram-negative organisms aminoglycosides have regained interest (2).

Aminoglycosides are very potent bactericidal antimicrobals, but nephrotoxicity is an important factor limiting their clinical utility. Although this is a clinically relevant adverse drug event of all systemic aminoglycosides, the nephrotoxic potential appears to differ within the drug class. For instance, compared to tobramycin gentamicin has a relatively high tissue accumulation in the renal proximal tubules, explaining its increased nephrotoxic potential (3-8). Furthermore, animal studies have confirmed gentamicin is more nephrotoxic than tobramycin (9, 10). In the late seventies and early eighties, during the era of multi-daily dosed (MDD) aminoglycoside regimens, human studies also suggested that gentamicin is more nephrotoxic than tobramycin (11). Nevertheless, since none of these studies complied with the methodological standards of a bias-free comparative drug trial, Smith et al. conducted a double-blind randomized clinical trial with larger sample size, showing that MDD gentamicin was indeed more nephrotoxic than MDD tobramycin (12). Related to this matter it is worth mentioning, that no real preference exist for either gentamicin or tobramycin as first line aminoglycoside with regard to worldwide resistance patterns (13-17). Over the past two decades traditional MDD schedules of aminoglycosides have largely been replaced by once daily dosed (ODD) aminoglycoside regimens with relatively more favorable toxicity profile. Aminoglycosides accumulate in renal proximal tubular cells and inner ear tissue (1, 6-8). Once daily dosing extends the period of minimal or drug free serum concentration allowing the renal and ear compartment more time to void the accumulated drug. Clinical studies have shown that once daily dosing of aminoglycosides substantially lowers the risk of nephro- and ototoxicity without compromising efficacy (18-20). Strong evidence exists that gentamicin has a higher nephrotoxic potential compared to tobramycin based on animal studies and clinical studies using MDD schedules. However, to date this has never been demonstrated for nowadays more common ODD regimens. Therefore, we investigated the nephrotoxicity and pharmacokinetics of ODD gentamicin versus ODD tobramycin in hospitalized patients with suspected or proven infection with gram-negative micro-organisms.

Patients and Methods

In this retrospective cohort study adult patients with nosocomial infections who were administered systemic gentamicin or tobramycin at the Medical Center Haaglanden Westeinde (The Hague, The Netherlands), a 500-bed teaching hospital, between January 2008 and December 2012 were selected from the electronic medication records. Following synchronization of regional antibiotic guidelines in February 2010, the local antibiotic guidelines switched from gentamicin to tobramycin for all indications requiring treatment with an aminoglycoside, without any changes in treatment and monitoring protocol. The selected patients were eligible for inclusion if they were treated with a once daily dose of 4-6mg/kg/day of gentamicin or tobramycin for a minimum of 3 days. Gentamicin and tobramycin were administered as 30 min infusions. According to the monitoring protocol, a peak serum concentration (C_{max}) and second concentration were drawn 0.5-1 h and 6-14 h after end of the first or second infusion, respectively. Aminoglycoside dosing was guided by drug concentration monitoring targeting a C_{max} of 15-20 mg/L and estimated trough concentrations <0.5 mg/L. In case of trough concentrations >0.5 mg/L, extended interval dosing was applied. In all other cases subsequent doses were administered every 24 hours. Drug concentrations were monitored at least twice weekly according to protocol. Timings of blood sampling for therapeutic drug monitoring were accurately recorded. Two serum creatinine concentrations were required for inclusion: one drawn within 48 hours before therapy and a minimum of one concentration drawn within 48 hours after initiation and 48 hours after cessation of therapy. Exclusion criteria were pre-existing renal impairment defined as a serum creatinine concentration ≥ 250 μ mol/L or an estimated glomerular filtration rate (eGFR; calculated with modified diet in renal disease formula (21)) of ≤ 20 ml/min at baseline, admittance to the intensive care unit, and a diagnosis of endocarditis. A waiver was provided by the local medical ethics committee. Nephrotoxicity was identified by the acronym RIFLE (Risk, Injury, Failure, Loss of kidney function, and End-stage kidney disease) with the exclusion of urine production values, since these could not consistently be retrieved from patient records (22). The rise in serum creatinine was determined by subtracting the initial concentration from the highest measured concentration drawn between 48 hours after initiation and

48 hours after cessation of therapy.(12) The incidence of nephrotoxicity was compared between gentamicin- and tobramycin-treated patients. Analogous to the rise in serum creatinine, delta blood urine nitrogen (BUN) was calculated. Pharmacokinetic (PK) parameters of tobramycin and gentamicin were calculated for all individuals using one-compartment open models and PK software (MW\PHARM 3.60, Mediware, The Netherlands) with an iterative two-stage Bayesian fitting procedure.

Statistical analysis

A Kolmogorov-Smirnov test was used to identify normal distribution of data. Means and standard deviations of continuous variables are presented in normal distributions. Otherwise, medians and ranges were reported. A Student's T-test was used to determine statistical differences for normally distributed data. If data were not normally distributed, a Mann-Whitney-U test was performed. A Chi-square test was used to compare dichotomous variables between groups. Uni- and multivariate logistic regression analyses were performed to identify determinants that were available prior to the start of therapy, i.e. aminoglycoside, gender, age, weight, ward and pretreatment creatinine, BUN and eGFR, prior to the second administration, i.e. C_{max}, estimated 24h-concentration, clearance, volume of distribution and the area-under-the-curve of the first administration (AUC_{first}) and available at the end of therapy, i.e. cumulative dose and duration of therapy. A stepwise backward elimination procedure was carried out, retaining covariates only if their removal significantly changed the model. To account for multiple testing only those covariates were tested that significantly affected the risk of nephrotoxicity in univariate analyses and a stringent P value ($P < 0.005$) during multivariate analysis was selected. Finally, the predictive value of AUC_{first} cut-off values of 120, 150, 180 and 210 mg*h/L on nephrotoxicity defined as RIFLE stage R were tested with multivariate regression. All tests were performed in SPSS Statistics version 20 (SPSS Inc., Chicago, IL, USA) and a P-value of < 0.05 was considered to be statistically significant and in each instance.

Results

Of the 2889 courses that were selected, 382 unique patients met the inclusion criteria and nephrotoxicity was compared in 202 and 180 patients who received gentamicin or tobramycin, respectively. Similar numbers and percentages of patients were excluded per inclusion criterion in each treatment group (Figure 1).

Baseline characteristics of the included patients did not statistically differ between groups (Table 1).

Table 1. Patient characteristics

	gentamicin	tobramycin	p-value
No. of patients	202	180	
Age (23)	65 (16.2)	63 (16.6)	.43§
% of males	67	72	.23*
Weight (24)	70 (16.8)	69 (17.5)	.49§
Baseline serum			
creatinine (nmol/mL)	93 (37)	89 (43)	.33§
eGFR (mL/min)	75 (40)	82 (43)	.12§
Ward			.13*
<i>Surgical (%)</i>	42	34	
<i>Internal (%)</i>	47	49	
<i>Pulmonary</i>			
<i>diseases (%)</i>	6	12	
<i>Other (%)</i>	5	4	
Nephrotoxic co-medication			
Cefalosporines (%)	72.7	68.1	.34*
NSAIDs (%)	30.3	31.9	.32*
Diuretics (%)	23.4	28.7	.24*
Ciprofloxacin (%)	2.2	2.1	.80*
Vancomycin (%)	1.7	3.3	.78*
Erythromycin (%)	1.3	0	.87*
Azytromycin (%)	0.9	0	.95*
Cyclosporine (%)	0.4	0.6	.91*
Indometacin (%)	0.4	0	.96*

Differences in categorical variables are expressed as numbers and percentages, and continuous variables as means if normally distributed and if not normally distributed as median. Abbreviations are: eGFR, glomerular filtration rate estimated with MDRD formula.

§Student's t-test, ^Mann-Whitney test, *Chi-squared test

The incidence of progression to RIFLE stage R was greater in the gentamicin group with nephrotoxicity developing in 43 of 202 patients (21.2%) as compared with 19 of 180 (10.6%) given tobramycin (odds ratio [OR] 2.4, 95%CI 1.2-4.7, $P=0.012$). RIFLE stages I and F were also more frequently observed in patients on gentamicin compared to tobramycin, but the risk was not statistically significant (Table 2).

Table 2. Nephrotoxicity according to RIFLE criteria in patients treated with gentamicin or tobramycin

RIFLE stage	gentamicin (n=202)	tobramycin (n=186)	OR	95%-CI	p-value
R	43	19	2.4	1.2-4.7	.01
I	11	7	1.5	0.5-3.8	.48
F	5	2	2.3	0.4-12.0	.45

Odds ratios for nephrotoxicity defined by the RIFLE classifications using multivariate analysis are presented. Abbreviations are: OR, odds ratio; 95%-CI, 95 % confidence interval

Univariate analyses identified gentamicin use ($P=0.006$), aminoglycoside clearance ($P=0.012$) and AUCfirst ($P<0.001$) as independent predictors. During multivariate analysis only gentamicin use ($P=0.08$) and AUCfirst ($P=0.02$) were retained by the model as independent predictors of progression to RIFLE stage R. Finally, an AUCfirst over 150 mg*h/L was associated with an increased risk of developing nephrotoxicity (OR 2.2, 95%CI 1.1-5.0, $P=0.048$). These results did not alter after correction for difference in baseline eGFR.

Mean increases in serum creatinine were 13.3 nmol/mL for the gentamicin group and 0.1 nmol/mL for patients given tobramycin ($P=0.02$) (Figure 2).

Figure 2. Mean increases in serum creatinine in tobramycin and gentamicin treated patients

The relative decrease in eGFR between groups showed a trend of a higher loss of eGFR for the patients given gentamicin compared to tobramycin (4.8 vs. 1%, $p=0.07$). No significant differences in delta BUN values were noted ($P=0.66$).

With respect to the pharmacokinetic parameters, the mean AUCfirst of tobramycin was significantly lower compared to gentamicin (98.3 vs. 85.6 $\text{mg}\cdot\text{h}/\text{L}$ respectively, $p=0.02$). The mean peak and estimated trough serum concentrations were within the predefined target range, with higher concentrations in patients treated with gentamicin. Estimation of pharmacokinetic parameters showed a higher mean volume of distribution and clearance of tobramycin compared to mean parameters found in patients on gentamicin (Table 3).

Table 3. Dosing and Pharmacokinetic parameters

	gentamicin	tobramycin	p-value
No. of patients	175	152	
Initial dose(mg)	330 (150-680)	350 (140-685)	0.21 [^]
Cumulative dose (mg)	1600 (1040-7620)	1775 (900-6600)	0.20 [^]
Duration of therapy (days)	5 (3-24)	5 (3-22)	0.32 [^]
AUC of first administration (mg*h/L)	98.3 (45)	85.6 (43)	0.02 [§]
Volume of distribution (L)	20.7 (7.2)	22.8 (6.5)	0.01 [§]
Clearance (L/h)	4.3 (2.5)	4.9 (2.2)	0.02 [§]
C _{max} (mg/L)	17.8 (6.5)	16.6 (6.2)	0.01 [§]
Estimated 24h-concentration (mg/L)	0.18 (5.0)	0.09 (3.7)	0.01 [^]

Differences in categorical variables are expressed as numbers and percentages, and continuous variables as means or medians and standard deviation or range between parentheses. Abbreviations are: AUC, area-under-the-curve and C_{max}, peak serum concentration.

§Student's t-test

[^]Mann-Whitney test

Discussion

This large retrospective cohort study shows that ODD gentamicin is more nephrotoxic than ODD tobramycin in clinically infected patients who were treated with aminoglycoside therapy for 3 days or longer. The higher nephrotoxic potential of gentamicin was associated with lower clearance and higher AUC's compared to tobramycin offering a biological explanation for this important finding. Furthermore, a high AUC of the first administration was associated with an increased risk of nephrotoxicity. To our knowledge this is the first study directly comparing the nephrotoxic effect among the two most frequently prescribed ODD intravenous aminoglycosides.

Despite the weakness of a retrospective design with consecutive cohorts the risks of both selection- and time bias appear low, since per criterion, similar numbers of patients were excluded, and there were no

significant differences between patient characteristics of both included and eligible patients (data not shown for the latter). The validity of our outcomes is strengthened by the large cohort investigated, and the hospital-wide switch to tobramycin in all aminoglycoside-indications, explaining similar baseline characteristics in both populations. Nevertheless, future prospective studies remain warranted to confirm these results and are preferably of randomized controlled design.

Whereas the AUC of the first administration and aminoglycoside clearances were significant univariate predictors of nephrotoxicity, baseline serum creatinine and eGFR were not predictive of developing nephrotoxicity. An explanation for these important secondary findings may be the route of excretion of aminoglycosides. As opposed to creatinine, which is also partially excreted by proximal tubular secretion, aminoglycosides are almost exclusively excreted via glomerular filtration (25,26). Secondly, the 'serum' AUC is expected to be a stronger predictor of nephrotoxicity than clearance, since it correlates best with aminoglycoside exposure in the distal tubuli (27). In fact, the daily AUC has previously been reported as significant predictor of nephrotoxicity by Rybak et al. (28). And thirdly, high serum creatinine concentrations will trigger the treating physician to initiate close monitoring and to consider extended interval dosing anticipating the increased risk of nephrotoxicity (29). Consequently, the 'a priori' higher risk of nephrotoxicity in patients with poor creatinine clearance will be reduced by the physician's preventive and corrective actions. Additionally, patients with pre-existing renal insufficiency which was a contraindication according to the local treatment protocol received other antibiotic agents. This renal function-based 'triage' methodology is vital to prevent kidney damage by aminoglycosides, but will concomitantly lower the predictive value of creatinine serum concentrations. In our opinion the optimal triage to identify patients who are at high at risk of nephrotoxicity if treated with an aminoglycoside for several days, should consist of baseline eGFR assessment followed by early AUC monitoring.

The randomized trial performed by Smith et al. found no significant and clinically relevant difference in ototoxicity of MDD gentamicin and tobramycin.

Unfortunately, pure-tone-audiometry for measuring hearing loss was not standard of care at our hospital and the risk of ototoxicity could not be investigated. Nevertheless, the low incidences of hearing loss and vertigo in clinically infected patients exposed to aminoglycosides less

then 7 days demand studies with larger samples sizes and/or novel more sensitive techniques of detecting ototoxicity such as otoacoustic emissions to detect differences between individual aminoglycosides (30).

As mentioned, once daily dosing strategies of 5-7 mg/kg offer a favourable pharmacokinetic/pharmacodynamic profile and are associated with lower incidence of kidney dysfunction and failure compared to MDD aminoglycoside regimens, since efficacy is assured by higher Cmax values for minimal inhibitory concentrations (MIC's) <1-2 mg/L and renal toxicity is minimized by the longer dosing intervals (31). In the current study lower Cmax values for tobramycin were found. Nevertheless, the predefined Cmax targets were attained for both aminoglycosides assuring efficacy. Furthermore, we found that estimated 24 hour concentrations were not predictive of nephrotoxicity. Importantly, the majority of the estimated trough concentrations were far below the limit of detection of currently available aminoglycoside serum concentration assays and these assays show high coefficients of variation in the lower concentration range (<0.5-1.0 mg/L) (32). Trough concentration monitoring thereby provides only limited information on drug exposure and pharmacokinetics in individual patients (33).

In conclusion, multidrug resistant micro-organisms and a dried up antibiotic pipeline have led to the resurgence of aminoglycosides. As their clinical usage is expected to expand in the years to come, it is of crucial importance to rediscover this class of highly effective antibiotics, and learn how to dose them in both an efficacious and safe manner. Our data suggest that in the once daily dosing era, a further reduction of the incidence of nephrotoxicity may be obtained by preferring tobramycin over gentamicin as first-line aminoglycoside in clinically infected patients. Moreover, the predictive value of 'first administration-AUC' suggests that early therapeutic drug monitoring can aid in reducing aminoglycoside related nephrotoxicity in patients that require treatment for three days or longer.

Funding

This work was not supported by outside funding. All work was funded by our department research budget.

Transparency declarations

None of the authors has conflicts of interest to declare.

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Chapter 2.4

**Extended-interval dosing of gentamicin in neonates
targeting trough concentrations under 0.5 mg/L:**

how low can we go?

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Abstract

Gentamicin-based regimens are often used in neonatal sepsis but should be used with care because of the narrow therapeutic index and potentially serious toxicity of gentamicin. In the adult populations, it is common to aim for a high maximum concentration (C_{max}) and a drug-free period (concentration <0.5 mg/L) of at least 4 hours to prevent toxicity. Albeit similar data in neonates are unavailable, there is no evidence that extended interval administration with target serum concentrations of <0.5 mg/L cannot be applied in this population. Therefore, the primary aim of this study was to prospectively assess the target attainment of a simple and practical dosing regimen of 5 mg/kg/36h aiming for trough concentrations <0.5 mg/L. In this prospective observational study neonates admitted to a level II neonatal nursery diagnosed with (suspected) early onset sepsis and commencing intravenous gentamicin therapy were included. According to the monitoring protocol, a peak concentration and second concentration were drawn 0.5-1 h and 6-18 h after end of the first or second infusion, respectively. Gentamicin dosing was guided by drug concentration monitoring targeting C_{max} values >8 mg/L, and estimated trough concentrations <0.5 mg/L. After exclusion of thirty patients because of insufficient data ($N=11$), GA <32 weeks ($N=7$) and PNA >7 days ($N=12$) a total of 184 unique patients were included for analysis. 90.4% of patients ($N=166$) achieved a C_{max} value >8 mg/L and a C_{min} <0.5 mg/L. In conclusion, the dosing regimen showed good performance with respect to target attainment in neonates admitted to a level II unit. Clinical studies investigating the pharmacokinetics and risk/benefit ratio of dosing strategies aiming for drug-free periods ($C_{min}<0.5$ mg/L) in neonatal populations are warranted.

Introduction

Early-onset neonatal sepsis occurs in the first seven days of life predominantly due to contaminated amniotic fluid or blood with predominant pathogens such as group B streptococcal (GBS), *Escherichia coli*, enterococci, *Listeria monocytogenes*, or other gram negative bacilli

(1) and is one of the primary causes of mortality among neonates (2). Current guidelines for empirical therapy in neonates with (suspected) sepsis recommend gentamicin-based regimens in preference to cefotaxime-based treatments, because of higher concentrations of susceptibility to gentamicin and the necessity to circumvent exerting selective pressure for resistance (3). Although the combination of the aminoglycoside gentamicin and a penicillin antibiotic has proven to be an effective treatment for staphylococcal species, enterococci and GBS infections (4), it should be used with care because of the narrow therapeutic index and potentially serious toxicity of gentamicin (5). Neonates have aberrant pharmacokinetics (PK) with high inter-patient variability compared to adult patients. As their kidneys are still in development, aminoglycoside clearance and glomerular filtration rate are correlated to gestational age (GA) and postnatal age (PNA) (6). Furthermore, over 70% of their body weight consists of water, which increases the volume of distribution (L/kg) of aminoglycosides (7). In neonates target peak serum concentrations (C_{max}) range from 5 to 12 mg/L and trough concentrations (C_{min}) from 0.5 to 2 mg/L. In contrast, in the adult population, it is common to aim for a C_{max} of 20 mg/L and a drug-free period (concentration <0.5 mg/L) of at least 4 hours (8). Although similar data in neonates are unavailable, there is no evidence that extended interval administration with target trough serum concentrations of <0.5 mg/L cannot be applied in this population (5). Another approach advocated in literature is area under the concentration-time curve (AUC) guided dosing. The AUC is believed to be a strong predictor of (nephro)toxicity (9) and the AUC over minimal inhibitory concentration (MIC) ratio is the surrogate pharmacodynamic index that correlates best with resistance suppression and treatment success (10). Albeit there are limited clinical data on the optimal gentamicin exposure in terms of AUC, it has been suggested that aiming for a 90% probability of a treatment success and under 10% probability of toxicity, an AUC_{24h} value of 75 mg.h/L should be aimed for when pathogen's MIC's are between 0.5 or 1.0 mg/L (9). The majority of current dosing regimens of aminoglycosides in neonates are based on data obtained from neonates admitted to level III intensive care unit also known as a neonatal intensive care unit (NICU). As these critically ill sometime very premature newborns show a large inter-patient variability in PK parameters, most dosing regimens consist of complex algorithms with different dosages and intervals depending on GA, PNA and/or body weight (5). It is, however, debatable whether complex algorithms with known high risk of prescription

errors (11) are required to attain predefined targets in non-critically ill term neonates admitted to a level II special care unit. The inter-patient variability in PK parameters is expected to be lower in this population in comparison with neonates admitted to a NICU due to a generally better clinical condition of the former. Therefore, the two primary aims of this study were (1) to prospectively assess the target attainment of a simple and practical dosing regimen of 5 mg/kg/36h aiming for low trough concentrations (<0.5 mg/L) and (2) to evaluate inter-patient PK variability in neonates admitted to a level II neonatal special care unit.

Methods

Study design and patients

In this prospective observational study neonates admitted to the level II neonatal nursery at the Juliana Children's Hospital (The Hague, The Netherlands) diagnosed with (suspected) early onset sepsis and commencing intravenous gentamicin therapy between January 2010 and January 2013 were eligible for inclusion. Main exclusion criteria were GA <32 weeks and PNA >7 days. A waiver for consent was provided by the regional medical ethics committee.

Gentamicin dosing and monitoring

A priori gentamicin was dosed 5 mg/kg every 36 hours administered as a 30 min infusion in combination with amoxicillin 100 mg/kg twice daily. According to the monitoring protocol, a peak concentration and second concentration were drawn 0.5-1 h and 6-18 h after end of the first infusion, respectively. Gentamicin dosing was guided by drug concentration monitoring targeting C_{max} values >8 mg/L, and estimated trough concentrations <0.5 mg/L. Dose adjustments were communicated by a clinical pharmacist before administration of the second dose. In case of estimated trough concentrations >0.5 mg/L, the dosing interval was extended. In all other cases subsequent doses were administered every 36 hours. Drug concentrations were monitored at least once every other dose according to protocol. Antibiotic treatment was stopped if blood cultures showed no bacterial growth within 3 days. Concentrations of gentamicin were measured using an automated chemiluminescent immunoassay (Architect c8000, Abbott Laboratories, Amstelveen, The Netherlands). The coefficient of variation for the gentamicin assay was less than 5% in the range of 1 to 10 mg/L. The limit of quantificati-

on (LOQ) was 0.3 mg/L.

Data collection and exposure analysis

The following demographic and clinical parameters were collected and registered in patients' clinical records: date of birth, gender, body weight, GA, PNA, Apgar score, date, time and dose of gentamicin administration and blood culture results. Furthermore, individual C_{max}, C_{min}, AUC, clearance (Cl) and volume of distribution (V_d) values were estimated using a one-compartment open model and PK software (MWPHARM version 3.60, Mediware, The Netherlands) with an iterative two-stage Bayesian fitting procedure. The performance of the dosing table was assessed by calculation of the percentage of patients attaining the predefined target exposure.

Statistical analysis

Relationships between BW, GA, PNA and PK parameters were analyzed using the Pearson's correlation test. Furthermore, uni- and multivariate logistic regression analyses were performed to identify covariates predictive of C_{max} target attainment failure. A stepwise backward elimination procedure was carried out, retaining covariates only if their removal significantly changed the model ($P < 0.005$). To account for multiple testing only those covariates were tested that significantly affected the risk of target attainment failure in univariate analyses and a stringent P value of 0.005 during multivariate analysis was selected. All tests were performed in SPSS Statistics version 20 (SPSS Inc., Chicago, IL, USA) and a P-value of < 0.05 was considered to be statistically significant and in each instance.

Results

During the 3-year enrolment-period 214 unique patients were diagnosed with (suspected) early-onset sepsis and treated with gentamicin treatment episodes. After exclusion of thirty patients because of insufficient data (N=11), GA < 32 weeks (N=7) and PNA > 7 days (N=12) a total of 184 unique patients (53% of male gender) were included for analysis. Their GA and PNA ranged from 32 to 42 weeks and 0 to 6 days with a median of 39 weeks and 0 days, respectively. Median body weight was 3.15 kg ranging from 1.2 kg to 5.1 kg. Median Apgar score 10 minutes postpartum was 9 ranging from 3-10.

As shown in Table 1, 90.4% of patients (N=166) achieved both a C_{max} value >8 mg/L and a C_{min} <0.5 mg/L. Because of the high attainment rate with respect to the C_{min} target, it was decided to estimate C_{24h} values and assess the target attainment as previously described for the eC_{min} values at 36 hours. Furthermore, inter-patient variability in clearance, half life and volume of distribution were 30, 19.3 and 16.7%, respectively. Next, significant correlations were found between GA and C_{max} (r 0.44, P<0.001) and between GA and C_{min} (r 0.42, P<0.001), between BW and C_{max} (r 0.58, P<0.001) and between BW and C_{min} (r 0.50, P<0.001). Correlations between AUC_T and GA (r 0.028, P=0.7) and between AUC_T and BW (r 0.064, P=0.4) were not significant (Figure 1). Finally, univariate binary logistic regression showed that both low body weight (P<0.001) and low GA (P<0.001) were significant predictors of C_{max} target attainment failure. During multivariate analysis only GA was retained as an independent predictor of under-exposure, meaning that neonates in the lower GA range would require relatively higher dose per kg body weight to achieve a C_{max} value over 8 mg/L. The patients (N=16) who did not reach C_{max} values >8 mg/L had a C_{max} between 4.9 and 7.9 mg/L. Body weights in 11 of these patients were under 2.7 kg (the first quartile) and 12 had a GA <35 weeks (the first quartile).

Blood culture results were positive in 5 of 186 patients (2.7%); Coagulase-negative staphylococcus (CNS) (N=4) or *Corynebacterium* species (N=1), three of which were considered to be positive due to contamination. No cases of ot- or renal toxicity were reported.

Table 1. Dosing, pharmacokinetics and target attainment rates

Dosing& PK parameters	Mean (S.D.)
Gentamicin dose (mg)	15.6 (4.0)
eC _{max} (mg/L)	10.8 (1.75)
eC _{min} (mg/L)	0.18 (0.12)
eC _{24h} (mg/L)	0.64 (0.29)
AUC _T	99 (17.1)
Volume of distribution (L/kg)	0.45 (0.075)
Half life (h)	5.7 (1.1)
Clearance (L/h/kg)	0.17 (0.051)
Target attainment	Rate (%)
C _{max} >8 mg/L	168/184 (91.3)
C _{min} <0.5 mg/L	179/184 (97.3)
C _{max} >8 mg/L and C _{min} <0.5 mg/L	166/184 (90.4)
C _{min} <1 mg/L	184/184 (100)
C _{24h} <1 mg/L	158/184 (85.9)
C _{24h} <0.5 mg/L	95/184 (51.6)

Values are presented as means with standard deviations (S.D.) and ranges or as rates and percentages. Abbreviations are eC_{max}, maximum serum concentration; eC_{min}, minimum serum concentration, eC_{24h}, estimated serum concentration 24h after end of infusion; AUC_T, area under the concentration-time curve of one dosing interval and V_d, volume of distribution.

Figure 1. Correlations between body weight, gestational age and exposure parameters

Correlations between BW and Cmax (Figure 1A, r 0.58, $P < 0.001$) and between GA and Cmax (Figure 1B, r 0.44, $P < 0.001$), between BW and Cmin (Figure 1C, r 0.50, $P < 0.001$) and between GA and Cmin (Figure 1D, r 0.42, $P < 0.001$), between BW and AUC $_{\tau}$ (Figure 1E, r 0.064, $P = 0.4$) and between GA and AUC $_{\tau}$ (Figure 1F, r 0.028, $P = 0.7$) are depicted. Abbreviations are BW, body weight; GA, gestational age; Cmax, maximum serum concentration; Cmin, minimum serum concentration; and AUC $_{\tau}$, area under the concentration-time curve of one dosing interval.

Discussion

In this prospective clinical study over ninety percent of the included patients reached the combined predefined C_{max} and C_{min} targets using a simple uniform dose of 5 mg/kg gentamicin every 36 hours aiming for low C_{min} values (<0.5 mg/l) in all neonates admitted to our level II unit with (suspected) early-onset sepsis. The good performance of the gentamicin dosing regimen is associated to the low inter-patient variability in population PK parameters relatively other reports of gentamicin PK in neonates (5, 12, 13). It should be noted, that most of the latter reports included neonates admitted to a level III unit. They are inherently prone to have a high variability in PK due to severe (underlying) pathology and to a wider range in GA and PNA which in turn are associated with a larger range of birth weight and developmental stage compared to neonates admitted to a level II unit (5). This is confirmed by our findings that low GA and BW were predictive of C_{max} target attainment failure.

As neonates with a GA over 32 weeks and diagnosed with (suspected) early onset sepsis (PNA <7 days) were included in this study, patients suffering from (suspected) late onset neonatal sepsis were not included as well as preterm neonates with a low GA (<32 weeks). The main exclusion criteria could therefore be considered as the most important limitations of the study outcomes. Yet, only seven neonates were excluded due to a GA under 32 weeks and no more than twelve patients had a PNA over 7 days at start of gentamicin therapy, which is less than 10% of the total population. These patients should have been referred to a NICU according to international guidelines (14) and the studied gentamicin dosing regimen was not intended to be used in these patients. In clinical practice, we suggest using a dosing algorithm specifically designed in a population of neonates with a GA under 32 weeks. For the twelve neonates with a PNA >7 days, a post hoc analysis was performed, which indicated that 10 out of twelve patients (82%) attained predefined C_{max} and C_{min} targets (data not shown). Although the sample size was too small to statistically compare the outcomes to neonates with a PNA <7 days at the initiation of therapy, these preliminary results suggest the dosing regimen may also show good performance in neonates with a PNA >7 days admitted to a level II unit.

One of the major strengths of this study is the combination of a prospective observational design with the minimal exclusion and loss of

data reflecting real-life clinical practice. Secondly, two plasma samples were drawn at sampling times optimal for accurate estimation of PK parameters. Using this approach, C_{min} and AUC values can be accurately estimated instead of sampling actual trough concentrations that are unreliable due to a high assay error at low gentamicin concentrations (15). This limited AUC sampling approach offers the opportunity to identify individual clearance next to the gentamicin 'load' represented by the AUC. In this study a mean gentamicin 'load' expressed as the AUC_{36h} of 99 mg·h/L was found. After correction for the extended interval a 5 mg/kg dose every 36 hours is a gentamicin 'load' equivalent to an AUC of 75 mg·h/L every 24 hours proposed by Drusano and co-workers that gives high chances of success and safety (9). Interestingly, the high target attainment rates may call for a revision of the intensive drug concentration monitoring after the first dose in all neonates > 32 weeks GA and < 7 days PNA. Sampling after the second dose could significantly reduce patients' blood sampling burden, since gentamicin is frequently administered for a short period of 36 to 72 hours in neonates (16). A further reduction in gentamicin course duration may be anticipated, since the turn-around-times of blood cultures have significantly been reduced in recent years. Especially, in combination with extended dosing interval dosing regimens this could facilitate current practices of stopping gentamicin therapy earlier, perhaps even before the second dose (16-18). Patients will not needlessly be exposed to gentamicin for longer periods, but will also be refrained from both the pain and risk of infection associated to blood sampling in neonates. Two meta-analyses that comprise approximately 600 newborns have shown that irrespective of the dosing schedule in combination with routine aminoglycoside blood concentration monitoring, sepsis was clinically cured in almost every patient (19, 20). In this context, one should realize that antibacterial courses are started in newborns even with a low probability of sepsis. Although many studies did not find any aminoglycoside-related toxicity, (transient) hearing loss and renal tubular damage have been reported (19, 21). Unfortunately, adequate diagnostic instruments are currently unavailable to sensitively and specifically identify aminoglycoside-toxicity in neonates. Currently, gentamicin trough concentrations or AUC's can be considered the best clinical 'markers' for toxicity monitoring (5). Because of the pharmacokinetic/pharmacodynamic profile of gentamicin and the rapid dose-dependent killing after the first dose, followed by adaptive resistance and a post-MIC effect, low trough concentrations

do not necessarily lead to a poorer outcome. A relatively drug-free interval as common in adult populations may reduce aminoglycoside-related toxicity and adaptive resistance in newborns (8). Hence, future clinical studies investigating the pharmacokinetics and risk/benefit ratio of dosing strategies aiming for drug-free periods ($C_{min} < 0.5 \text{ mg/L}$) in neonatal populations are warranted.

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Chapter 3

Chapter 3.1

Vancomycin Exposure Variability of Intermittent versus Continuous Infusion: a systematic review

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Submitted

Abstract

This systematic review aims to compare exposure variability and target attainment of vancomycin intermittent infusion (InI) and continuous infusion (CoI). A literature search was performed and clinical studies reporting on vancomycin treated populations were selected. After exclusion of reviews, case-reports and articles not published in the English language, 505 articles were screened for data on vancomycin serum concentrations. A total of 34 studies were included for review. A comparison of relative standard deviations (RSD) of vancomycin serum concentrations was performed to quantify InI and CoI exposure variability. The variability in serum concentrations was significantly larger for InI than CoI (RSD 46.5 and 32.1%, respectively; $p=0.001$). Exposure variability appeared to be independent of the study population or design. Studies directly comparing target attainment between both modes of administration denoted higher and faster target attainment with CoI. In conclusion, CoI was associated with lower exposure variability and favourable target attainment rates compared to InI. It remains, however, to be elucidated whether vancomycin CoI results in better clinical outcomes compared to InI.

Introduction

The glycopeptide antibiotic vancomycin was developed in the 1950s, when penicillin-resistant *Staphylococcus aureus* called for new antibiotics. In the early years of its development, vancomycin was known to cause nephrotoxicity and ototoxicity as well as infusion-related toxicities. These toxicities were most likely caused by impurities in the formulation, which gave it a brownish appearance and vancomycin was therefore also dubbed 'Mississippi mud' (1). After the fermentation process was changed, yielding purer vancomycin, the rate of toxicity was markedly reduced (2). Primarily, the clinical use of vancomycin declined when semisynthetic penicillins such as methicillin, oxacillin and nafcillin became available, which were considered to be less toxic. However, in the 1970s vancomycin regained interest as the result of the rise of methicillin resistant *Staphylococcus aureus* (MRSA) infections (1, 3). Currently, vancomycin is the drug of choice in the treatment of severe MRSA infections such as sepsis, endocarditis, osteomyelitis

and health care-associated or community-acquired pneumonia (4), as well as infections caused by other gram-positive bacteria. Usually, vancomycin is administered intermittently and serum concentrations are monitored (4). However, it has been suggested that continuous infusion of vancomycin may reduce the rates of nephrotoxicity without compromising efficacy and target serum concentrations may be attained faster (5). The studies comparing clinical outcomes between intermittent (InI) and continuous (CoI) infusion of vancomycin were recently reviewed in a meta-analysis by Cataldo et al. (6). A significant lower rate of nephrotoxicity was observed with CoI compared to InI in this meta-analysis. However, no differences in mortality were noted and the definition of efficacy was inconsistent in the included studies to pool efficacy outcome measures (6). An alternative strategy to compare InI and CoI of vancomycin is to compare surrogate outcome parameters e.g. variability in drug exposure or target attainment rates. Minimal interpatient variability in exposure and rapid target attainment are important aims of any vancomycin dosing strategy striving for maximum efficacy and minimal toxicity. Therefore, the present review aims to compare the variability of exposure of InI and CoI in vancomycin treated populations using serum concentrations in published data. Furthermore, target attainment with either mean of administration will be assessed.

Methods

To assess the variability of exposure and target attainment of InI and CoI vancomycin treatment, a literature search in PubMed/MEDLINE (1950 to February 2014), Web of Science (1900 to February 2014) and Embase (1947 to February 2014) was conducted and the reference lists of included articles were screened manually. The index search terms in PubMed were 'vancomycin' AND (serum concentration OR level* OR concentration*) AND (dosing OR intravenous) and the search was limited to articles published in English and studies performed in humans. An adapted search was performed in Embase and Web of Science. Clinical studies, as well as pharmacodynamic and pharmacokinetic studies were eligible for inclusion. Studies with the following criteria were excluded: paediatric data, data on haemodialysis, and studies that did not report serum concentrations or area under the concentration-time curve (AUC) (mean $\pm$ SD) values.

From the included articles, data on target attainment rates and serum

concentrations or AUCs (mean $\pm$ SD) were extracted. Next, the variabilities in AUC, trough and 'plateau' serum concentrations was determined by the reported relative standard deviation (RSD). If multiple serum concentrations and AUC values were reported per treatment arm, they were included separately.

Mean RSDs of vancomycin exposure for InI and CoI were compared by an Independent samples T-test. Statistical analysis were performed using SPSS Statistics version 20 (IBM Corporation, Chicago Ill, USA, 2011). All tests were performed two-sided and a p-value of <0.05 was considered statistically significant.

Results

In total 34 articles were included (see Figure 1 for a flow chart of inclusion).

A summary of study design, dosing regimen and exposure target of the included studies is displayed in Table 1.

Figure 1 Flow chart of inclusion

Table 1

Study ID	Design	Sample size		Setting/Condi- tion	Vancomycin administration and target		Pharmacokinetics					
							Inl		Col		Inl	
		Inl	Col		Inl	Col	C _{min} (mg/L)	RSD (%)	C _{ss} (mg/L)	RSD (%)	AUC (mg.h.L ⁻¹)	RSD (%)
Akers, K.S. (2012) (8)	Ret. cohort	81	90	Critically ill burn patients	D = 1 g every 8h C _{min} = 15- 20 mg/L	D = 3 g/d; no LD. C _{ss} = 20- 25 mg/L	14.8 ± 4.4	29.7	20.0 ± 3.8	19.0	ND	ND
Al-Kofide, H. (2010) (9)	Ret. cohort	Cancer: 18 Control: 13	0	Cancer vs. control patients	ND	-	Cancer: 7.1 ± 5.5 Control: 8.5 ± 4.5	Cancer: 77.5 Control: 52.9	-	-	ND	-
Ampe, E. (2013) (13)	Pros. cohort	0	94	Gen. ward patients	-	LD = 20 mg/kg; C _{ss} = 25- 30 mg/L	-	-	27.8 ± 5.7	20.5	-	661 ± 60
Cianferoni, S. (2013) (17)	Ret. cohort	0	207	ICU patients	-	D = 20-30 mg/kg; LD = 15 mg/kg; C _{ss} = 20- 30 mg/L	-	-	22.3 ± 7.0	31.4	-	ND
Clemens, E.C. (2011) (35)	Ret. cohort	MIC ≤1.5 mg/L: 94 MIC = 2 mg/L: 24	0	Gen. ward and ICU patients	ND	-	MIC ≤1.5 mg/L: 17.7 ± 7.3 MIC = 2 mg/L: 16.0 ± 5.0	MIC ≤1.5 mg/L: 42.2 MIC = 2 mg/L: 31.3	-	-	-	-
Dailly, E. (2008) (44)	Ret. cohort	0	149	Burn patients	-	D = 0.5-6 g/d	-	-	17.1 ± 8.6	50.3	-	ND
De Waele, J.J. (2013) (18)	Ret. cohort	0	227	ICU patients	-	D = 2 g/d; LD = 1000 mg (<65 kg) or 1.5 g (≥65 kg); C _{ss} = 15-25 mg/L	-	-	19.32 ± 6.12	31.7	-	ND

Study ID	Design	Sample size		Setting/Condition	Vancomycin administration and target		Pharmacokinetics							
							Inl		Col		Inl		Col	
		Inl	Col		Inl	Col								
							C_{min} (mg/L)	RSD (%)	C_{ss} (mg/L)	RSD (%)	AUC (mg·h·L ⁻¹)	RSD (%)	AUC (mg·h·L ⁻¹)	RSD (%)
Di Filippo, A. (1998) (19)	Ret. cohort	14	11	ICU patients	D = 500 mg every 6h D = 83 mg/h; LD = 500 mg		ND	ND	24.3 ± 3.9	16.0	ND	ND	ND	ND
Healy, DP. (1987) (37)	Randomized cross over	11	0	Healthy volunteers	D: 500 mg every 6 h or 1 g every 12 h		500 mg: 11.2 ± 2.2 1 g: 7.9 ± 1.7	500 mg: 19.6 1 g: 21.5	-	-	500 mg: 116 ± 15.6 1 g: 227 ± 28.3	13.4 12.5	-	-
Hochart, C. (2011) (10)	Ret. cohort	0	54	Febrile neutropenic patients	-	D = 40 mg/kg/d, adjusted to CrCl; LD = 15 mg/kg; $C_{ss} > 20$ mg/L	-	-	14.4 ± 4.7	32.6	-	-	ND	ND
Horey, A. (2012) (31)	Ret. cohort	270	0	Gen. ward and ICU patients	ND	-	15.5 ± 7.8	50.3	-	-	ND	ND	-	-
Hutscha, D. (2009) (20)	Ret. cohort	30	119	ICU patients	$C_{min} = 15$ mg/L	$C_{ss} = 20$ -25 mg/L	17.0 ± 4.7	27.6	25.0 ± 4.0	16.0	612 ± 213	34.8	529 ± 98	18.5
Ingram, PR. (2008) (39)	Ret. cohort	0	102	Outpatients	-	$C_{ss} = 20$ -25 mg/L	-	-	15.5 ± 10.8	69.7	-	-	ND	ND
Ingram, PR. (2009) (38)	Matched ret. cohort	40	40	Outpatients	ND	ND	9.7 ± 5.0	51.5	13.6 ± 6.2	45.6	ND	ND	ND	ND
James, JK. (1996) (29)	Pros. cross-over	10	10	Gen. ward and ICU patients	D = 1 g every 12 h $C_{min} = 5$ -10 mg/L	D = 80 mg/h $C_{ss} = 15$ mg/L	8.4 ± 5.9	70.2	20.2 ± 11.1	55.0	550.0 ± 285.7	48.3	552.6 ± 373.4	67.6
Karam, CM. (1999) (30)	Ret. cohort Pros. cohort	Ret: 120 Pros: 120	0	Gen. ward and ICU patients	Ret: $C_{min} = 5$ -10 mg/L Pros: $C_{min} = 5$ -20 mg/L	-	Ret.: 10.8 ± 7.4 Pros: 13.9 ± 9.6	Ret: 68.5 Pros: 69.1	-	-	ND	ND	-	-
Klepser, ME. (1998) (36)	Randomized cross-over	12	12	Healthy volunteers	D = 1 g every 12 h	D = 1 or 2 g/d	5.5 ± 1.9	34.5	D: 1 g: 8.8 ± 1.6 D: 2 g: 16.9 ± 1.9	D: 1 g: 18.2 D: 2 g: 11.2	-	-	-	-

Study ID	Design	Sample size		Setting/Condition		Vancomycin administration and target		Pharmacokinetics							
								Inl				Col			
		Inl	Col			Inl	Col	C _{min} (mg/L)	RSD (%)	C _{ss} (mg/L)	RSD (%)	AUC (mg.h.L ⁻¹)	RSD (%)	AUC (mg.h.L ⁻¹)	RSD (%)
Lodise, TP, (2008) (32)	Ret. cohort	246	0	Gen. ward and ICU patients	ND	-	-	12.4 ± 5	40.3	-	-	ND	ND	-	-
Maki, N, (2012) (33)	Ret. cohort	106	0	Gen. ward and ICU patients	D = 0.5-2 g/d in 1-4 doses	-	-	12.6 ± 6.5	51.6	-	-	ND	ND	-	-
Michalets, EL, (2011) (14)	Ret. cohort	67 nom, 66 control group	0	Gen. ward patients	Control: D = 15-20 mg/kg/D; Nom: AUC ≥ 400 and C _{min} ≥ 15 mg/L.	-	-	Control: 13.6 ± 8.5 Nom: 20.7 ± 10.8	Control: 62.5 Nom: 52.2	-	-	Control: 383 ± 208 Nom: 504 ± 158	Control: 54.3 Nom: 31.3	-	-
Mohammed, I, (2006) (21)	Pros. cohort	0	20 LD = 500 mg (A), 20 LD = 15 mg/kg (B)	ICU patients	-	LD = 500 mg (A) or 15 mg/kg (B); C _{ss} = 20 mg/L	-	-	-	A: 14.9 ± 5.4 B: 18.5 ± 6.4	A: 36.2 B: 34.6	-	-	ND	ND
Niu, SC, (2008) (15)	Pros. cohort	43 conv. protocol, 42 modified protocol based on SCr.	0	Gen. ward patients	Conv.: D = 10-15 mg/kg every 12h; Modified: C _{min} = 5-15 mg/L	-	-	Conv.: 21.4 ± 12.7 Modified : 47.3 ± 5.3	Conv.: 59.3 Modified : 47.3	-	-	ND	ND	-	-
Ocampos-Martinez, E, (2012) (22)	Ret. cohort	0	261	ICU patients	-	D = 30 mg/kg; LD = 15 mg/kg; C _{ss} = 20-30 mg/L	-	-	-	20.4 ± 8.9	43.6	-	-	ND	ND
Omote, S, (2009) (41)	Ret. cohort	Cancer: 65 Control: 41	0	Cancer vs. control patients	ND	-	-	Cancer: 11.5 ± 5.4; Control: 48.1	Cancer: 47.0 Control: 48.1	-	-	ND	ND	-	-

Study ID	Design	Sample size		Setting/Condition		Vancomycin administration and target		Pharmacokinetics							
								Inl				Col			
		Inl	Col	Inl	Col	Inl	Col	C _{min} (mg/L)	RSD (%)	C _{ss} (mg/L)	RSD (%)	AUC (mg·h·L ⁻¹)	RSD (%)	AUC (mg·h·L ⁻¹)	RSD (%)
Pea, F. (2000) (23)	Ret. cohort	18	0	ICU patients	-	D = 500 mg every 6-12 h.	-	9.12 ± 1.26	13.8	-	-	ND	ND	-	-
Pea, F. (2009) (34)	Ret. cohort Pros. cohort	0	Ret: 70 Pros: 63	Gen. ward and ICU patients	-	-	C _{ss} = 15-20 mg/L	-	-	Ret: 15.5 ± 4.8 Pros: 15.8 ± 5.0	Ret: 31.0 Pros: 31.6	-	-	ND	ND
Ricard, J.D. (2007) (24)	Pros. cohort	0	14	ICU patients	-	-	D = 60 mg/kg LD = 15	-	-	25.5 ± 7.3	28.6	-	-	ND	ND
Schmelzer, T.M. (2013) (25)	Pros. randomized trial	36	37	ICU patients	-	D = 15 mg/kg every 12 h LD = 20 mg/kg	D = 21.6-57.6 mg/kg LD = 20 mg/kg	8.9 ± 3.9	43.8	19.8 ± 6.13	31.0	ND	ND	ND	ND
Shimamoto, Y. (2013) (26)	Ret. cohort	24	0	ICU patients	-	C _{min} = 15-20 mg/L	C _{min} = 15-20 mg/L	13.6 ± 6.9	50.7	-	-	477 ± 191	40.0	-	-
Suzuki, Y. (2012) (16)	Ret. cohort	31	0	Gen. ward	-	ND	-	16.3 ± 6.8	41.7	-	-	540.6 ± 165.0	30.5	-	-
Van Maarseveen, E.M. (2014) (E.M. van Maarseveen, S.G.H. Gijmans, E.C. Vastbinder, M. Pejjak and A.R. van Zanten, submitted for publication)	Ret. cohort	27	44	ICU patients	-	D = 15 mg/kg every 12-24 h C _{min} = 8-12 mg/L	LD = 1 g C _{ss} = 16-20 mg/L AUC ≥ 350 mg·L ⁻¹ ·h ⁻¹	9.25 ± 4.55	49.2	16.37 ± 4.71	28.8	288.6 ± 87.7	30.4	365.1 ± 91.2	25.0

Study ID	Design	Sample size		Setting/Condi- tion		Vancomycin administration and target		Pharmacokinetics					
								Inl		Col		Inl	
		Inl	Col			Inl	Col	C _{min} (mg/L)	RSD (%)	C _{ss} (mg/L)	RSD (%)	AUC (mg.h.L ⁻¹)	RSD (%)
Vuagnat, A. (2004) (40)	Pros. cohort	21	23	Osteomyelitis		D = 40 mg/kg/d C _{min} = 20- 25 mg/L	D = 40 mg/kg/d C _{ss} = 20- 25 mg/L	21.7 ± 9.3	42.9	26.0 ± 6.1	23.5	ND	ND
Wysocki, M. (2001) (27)	Pros. randomized trial	58	61			D = 30 mg/kg/d C _{min} = 10- 15 mg/L	D = 30 mg/kg/d; LD = 15 mg/kg; C _{ss} = 20- 25 mg/L	ND	ND	ND	ND	653 ± 232	577 ± 120
Zelenitsky, S. (2013) (26)	Ret. cohort	35	0	ICU patients		ND	-	16.6 ± 8.4	50.6	-	-	484 ± 235	48.6

Serum concentration and AUC's are displayed as means ± standard deviations (S.D.).

Abbreviations: AUC, area-under-the-concentration-time-curve; C_{min}, trough serum concentration; Col, continuous infusion; Conv., conventional dosing; CrCl, creatinine clearance; C_{ss}, plateau serum concentration; D, dose; Gen. ward, general ward; ICU, intensive care unit; Inl, intermittent infusion; LD, loading dose; ND, not determined; Nom., nomogram dosing; Pros., prospective; Ret., retrospective; RSD, relative standard deviation; SCr, serum creatinine.

Vancomycin treated populations

The vancomycin target concentration range depends on the indication and patient population. The consensus guidelines by the American Society of Health-System Pharmacists (ASHP), the Infectious Diseases Society of America (IDSA) and the Society of Infectious Diseases Pharmacists (SIDP) advise to maintain a serum trough concentration of at least 10 mg/L to prevent development of resistance. More severe infections caused by *S. aureus*, such as endocarditis, osteomyelitis, meningitis and hospital-acquired pneumonia, may require higher trough concentrations of 15-20 mg/L to achieve an AUC/MIC ≥ 400 when the pathogen's MIC ≤ 1 mg/L. A loading dose of 25-30 mg/kg bodyweight may be considered for critically ill patients to ensure rapid target attainment (4). Since Col is not supported by the IDSA guidelines (4), the targets for this mode of administration are less well defined. However, it may be assumed that the same pharmacokinetic/pharmacodynamic (PK/PD) target of AUC/MIC ≥ 400 applies to Col, which would require a plateau concentration of 15-25 mg/L for pathogens with an MIC ≤ 1 mg/L (5). Pathophysiological processes or concomitant therapies may affect vancomycin disposition and clearance and thereby introduce variability of exposure and target attainment. The volume of distribution and vancomycin clearance can vary significantly between populations (7-10). Due to the hydrophilic properties of vancomycin, its volume of distribution is mainly limited to the extracellular fluid content (7). Therefore, any change in this content will influence the serum concentration. Furthermore, vancomycin clearance occurs predominantly through glomerular filtration of the unchanged form (11). Changes in renal function are likely to alter vancomycin clearance and thus exposure. Larger variability in vancomycin concentrations and AUCs can thus be expected in populations with large interpatient variability in volume of distribution and clearance, such as critically ill patients (12). Since exposure variability is largely dependent of the specific population, vancomycin exposure and target attainment rates are reported and discussed in detail per study population: general ward patients, patients admitted to the intensive care unit (ICU), mixed general ward and ICU populations, healthy volunteers, and other populations.

General ward patients

Three studies evaluated nomogram dosing in general ward patients; two studies administered vancomycin by InI and one study by Col (13-15). The InI studies compared conventional dosing based on body weight to dosing according to a nomogram based on body weight as well as creatinine clearance. Both studies suggest that InI dosing yields better target attainment with a dosing schedule based on body weight and creatinine clearance (14, 15). Ampe et al. implemented a nomogram for Col of vancomycin in the general ward and found a variability in serum concentration of 20.5%. Variability in AUC was very low (RSD 9.1%), probably because the AUC_{24h} was determined from 48 hours after initiation up to the end of treatment (13). Variability in AUC is expected to be highest in the first 24 hours of treatment, which will be discussed later. To summarize, four studies reported means and standard deviations of serum concentrations or AUCs (mean $\pm$ SD) in general ward patients (13-16). Three studies used InI as mode of administration and in the remaining study (14-16), patients were treated with Col (13). Overall, variability appeared to be larger in patients that received vancomycin through InI compared with Col. The median RSDs (range) of trough versus plateau concentrations were 52.2% (41.7-62.5%) and 20.5% (-), for InI and Col respectively. Only one RSD was available for Col, compared to five for InI.

ICU patients

It is widely accepted that patients admitted to the ICU should be considered a pharmacokinetically heterogeneous population (12). Inflammation leads to extravasation of fluids to the interstitial space, thereby increasing the volume of distribution. Furthermore, inotropic drugs can significantly increase cardiac output in critically ill patients. As a result, renal blood flow and thus vancomycin clearance can be enhanced substantially. This condition is known as glomerular hyperfiltration. However, in case of cardiac depression, cardiac output may be decreased leading to lower renal blood flow. Acute kidney insufficiency may occur with concomitant decreased vancomycin clearance (12). Thirteen studies reported serum concentrations or AUCs (mean $\pm$ SD) with InI or Col in ICU patients (17-28) (E.M. van Maarseveen, S.G.H. Gipmans, E.C. Vasbinder, M. Petjak and A.R. van Zanten, submitted for publication). Four of these were direct comparisons of InI versus Col (20, 25,

27) (E.M. van Maarseveen, S.G.H. Gipmans, E.C. Vasbinder, M. Petjak and A.R. van Zanten, submitted for publication). In three studies (23, 26, 28), InI was the only mode of infusion used and in the remaining six studies patients were treated through Col (17-19, 21, 22, 24). Only two of these comparative studies in ICU patients were prospective randomized trials. In the most recent one, Schmelzer et al. compared target attainment in the first 48 hours in patients with ventilator-associated pneumonia (25). The target ranges for InI and Col were 15-20 mg/L and 15-25 mg/L, respectively. Only 7% of the patients treated with InI attained trough concentrations in the target range. An adequate plateau concentration was attained in 57% of the patients treated with Col ($p < 0.0001$). Furthermore, variability in serum concentrations seemed to be lower with Col (31.0% versus 43.8% with InI) and fewer dose adjustments were necessary with Col compared to InI ($p = 0.002$). The authors choose to power the study solely as a pharmacokinetic study and reported no clinical outcomes. In the second prospective trial, Wysocki and co-workers showed that variability in AUC and daily dose were lower with Col compared to InI ($p = 0.026$ and 0.057 , respectively) (27). The authors also reported faster target attainment with Col, which may be clinically relevant in critically ill patients. Targets were attained within 51 ± 39 hours and 36 ± 31 hours in InI and Col, respectively ($p = 0.03$). The remaining direct comparisons were retrospective cohort studies. Hutschala et al. confirmed target attainment was reached faster with Col (16 ± 8 hours) compared to InI (50 ± 21 hours; $p < 0.001$) in patients after cardiac surgery (20). The targets were 15 mg/L for InI trough and 20-25 mg/L for Col plateau concentrations. Van Maarseveen et al. compared variability of exposure in ICU patients treated with InI to Col treatment (E.M. van Maarseveen, S.G.H. Gipmans, E.C. Vasbinder, M. Petjak and A.R. van Zanten, submitted for publication). Prior to dose adjustment based on TDM, a higher variability in serum concentration (RSD 49.2 and 28.7% for InI and Col, respectively) and AUC (RSD 30.4 and 24.9% for InI and Col, respectively) was detected in patients treated with InI. After dose adjustment based on TDM, the variability in exposure did not change notably for both modes of administration. Furthermore, albeit correlations between serum concentrations and AUCs for both modes of administration were significant ($p < 0.01$), correlation of Col compared to InI was significantly higher (determinant of correlation (r^2) 0.932 and 0.720, respectively). De Waele et al. sought to explore target attainment, defined as a plateau concentration of 15-25 mg/L, within 48 hours in patients treated

with Col (18). Target attainment after 48 hours of therapy was 70.5% and increased to 84.1% on day 3. A loading dose of ≥ 15 mg/kg was significantly associated with attaining adequate vancomycin concentrations on day 2 ($p = 0.006$). Ocampos-Martinez et al. assigned a target range of 20-30 mg/L to septic patients on Col vancomycin treatment (22). Adequate concentrations were found in 36% of the patients on day 1. This increased to 48% on day 2, even though the dose was only adapted in 41% of the patients with subtherapeutic vancomycin concentrations. On day 3, target attainment was 68%. Male sex and lower creatinine clearance were risk factors associated with insufficient serum concentrations on day 1 and 2. Zelenitsky et al. retrospectively collected InI pharmacokinetic data of patients in septic shock (28). No predefined targets were reported, but the authors observed a main trough concentration ≥ 10 mg/L in 77.1% and ≥ 15 mg/L in 48.6% of the patients. In 60% of the patients, AUC was greater than 400 mg.h.L⁻¹. In conclusion, a relatively high variability in exposure is reported in this subset of patients due to a relatively large interpatient variability in PK parameters. Nevertheless, the relative difference between the median RSDs for InI trough concentrations and Col plateau concentrations were similar to other populations included in this review: 46.5% (range 13.8-50.7%) and 31.2% (range 16.0-43.6%), respectively. These findings are based on ten RSDs for Col and six for InI.

Mixed General ward and ICU patient population

James et al. conducted a prospective randomized cross-over study in which vancomycin was administered through InI or Col for two consecutive days in 10 patients (29). Even though variability in serum concentration appeared to be lower with Col (RSD = 55.0% with Col and RSD = 70.2% with InI), variability in AUC appeared to be higher (RSD = 67.6% and 48.3% with Col and InI, respectively). Of note, this is the only study directly comparing InI and Col in which variability in either serum concentration or AUC appeared to be higher in Col than InI. Only one study assessed target attainment. Karam et al. conducted a study in order to minimize monitoring and drug adjustments with InI vancomycin (30). Nomogram dosing led to dose adjustments in 20.3% of the patients and predefined target exposure was attained in 94%. However, a wide target range of 5-20 mg/L was chosen compared to other studies included in this review. To conclude, seven studies made no distinction between general ward and ICU patients and studied

vancomycin in hospitalized patients (29-35). As may be expected from such heterogeneous populations, variability in serum concentration was high in this subset of patients (median RSD 31.6, range 31.0-55.0 for Col based on three RSDs and median RSD 51.0, range 31.3-70.2 for InI based on eight RSDs).

Healthy volunteers

Klepser et al. conducted a randomised, open-label, cross-over trial in which volunteers received a three day course of InI (1000 mg/day) or Col (1000 or 2000 mg/day) vancomycin (36). Samples were obtained after two days of treatment. Variability in serum concentration was higher with InI (RSD = 34.5%) compared to Col (RSD = 18.2 and 11.2% for 1000 mg/day and 2000 mg/day, respectively). Thrombophlebitis was a frequently reported complaint, mainly in patients treated with Col. This was due to infusion via a peripheral vein as opposed to a central vein. Healy et al. compared InI twice daily to four times a day in a randomised cross-over study with eleven healthy volunteers (37). The variability in trough concentrations (RSD 19.6% for four times a day and 21.5% for twice daily) and AUC (RSD 13.4% and 12.5% for four and two times a day, respectively) were comparable between both groups. The variation in serum concentrations could mainly be explained by body weight according to the authors. In conclusion, variability of exposure in healthy volunteers was relatively low, probably due to strict inclusion and exclusion criteria, resulting in a population with limited variability in PK parameters. Two studies were conducted in healthy volunteers, yielding two RSD's for Col and three for InI (36, 37). Median (range) RSD's of 14.7% (11.2-18.2%) and 21.5% (19.6-34.5%) for serum concentrations were acquired with Col and InI, respectively.

Other populations

The remainder of the studies included burn patients, orthopaedic outpatients, oncologic patients and febrile neutropenic patients. In a retrospective cohort study, Akers et al. evaluated the use of InI and Col in critically ill burn patients (8). Burn patients are known to have a larger volume of distribution and clearance when compared to non-burn patients. Target trough and plateau concentration were 15-20 mg/L and 20-25 mg/L, respectively. These targets were attained in 22.6% of the patients treated with InI and 30.7% of the patients treated with Col ($p = 0.01$). Furthermore, variability in serum concentration was larger

with InI as opposed to Col (RSD = 29.7 and 19.0%, respectively). No differences in clinical outcomes were observed. However, the study was underpowered to assess outcomes such as mortality and nephrotoxicity.

Three studies were conducted in orthopaedic outpatients (38-40). It should be mentioned that these patients are at increased risk of suboptimal therapeutic drug monitoring due to inappropriate timed sampling, especially for InI. Vuagnat et al. conducted a prospective comparative study in patients with osteomyelitis and strived for high vancomycin trough concentrations (trough and plateau level of 20-25 mg/L) (40). Treatment was initiated in hospitalized patients who were discharged upon achieving steady-state concentrations. The target serum concentrations were attained after 2.5 ± 2.8 weeks for InI and 1.4 ± 0.7 weeks for Col ($p = 0.18$). Variability in serum concentration at steady-state was significantly lower with Col than with InI ($p = 0.001$). Ingram et al. conducted two retrospective studies that focused on nephrotoxicity (38, 39). Only one of these studies directly compared InI or Col and yielded comparable variability in serum concentration (RSD = 51.5% for InI and 45.6% for Col) (38).

Two retrospective cohort studies compared vancomycin pharmacokinetics in cancer patients to non-cancer patients (9, 41). Al-Kofide et al. found that the volume of distribution and vancomycin clearance were higher in cancer patients ($p = 0.002$ and $p = 0.005$, respectively) (9). This resulted in significantly higher peak but not trough concentrations in the control patients compared to the cancer patients ($p = 0.027$ and $p = 0.458$, respectively). Furthermore, variability in trough concentration was higher in cancer patients compared to controls (RSD = 77.5% versus 52.9%, respectively). In contrast, Omote et al. did not report enhanced vancomycin clearance in cancer patients when compared to control patients (41). Variability in serum concentration was comparable between the two groups (RSD 47.0 and 48.1% for cancer and control patients, respectively).

Hochart et al. conducted a retrospective cohort study in patients with febrile neutropenia to assess the attainment of adequate initial or follow-up serum concentrations (10). The volume of distribution is often increased in these patients. Vancomycin was administered through Col and the target concentration was 20 mg/L. This target was initially attained in 12% of the patients. In only 40% of the cases, serum monitoring was performed optimally. Eventually 32% had a plateau serum concentration of ≥ 20 mg/L after dose adjustment.

Overall comparison

The patient populations in the included studies were very heterogeneous: two studies were conducted in healthy volunteers; four in general ward patients; thirteen studies were performed in ICU patients and seven studies were performed in hospitalized patients (general ward and ICU). Furthermore, eight studies were conducted in specific populations such as outpatients, which were mainly orthopaedic patients; cancer versus non-cancer patients; febrile neutropenia and burn patients. Variability in serum concentration, expressed as RSD, ranged from 11.2 to 77.5% with a mean of 40.3%. Overall, variability in serum concentration as well as AUC appeared to be higher in vancomycin InI treated patients compared to Col treated patients. The mean RSDs in serum concentration were 46.5% (15.4%) and 32.1% (14.1%) for InI and Col, respectively and differed significantly between both treatment groups ($p = 0.001$) (Figure 2).

Figure 2. Boxplots of the relative standard deviations of serum concentration with intermittent or continuous infusion of vancomycin.

The influence of study design and number of patients included in the study was explored and reported in table 2.

Table 2. Distribution of relative standard deviations (RSDs) of serum concentrations in different study designs

Mode of administration	Study design	No. of RSDs (n)	No. patients per study (n), median	RSD (%) mean $\pm$ SD
InI	Total	30	38	46.5 $\pm$ 15.4
	Ret. cohort	20	41	47.0 $\pm$ 14.3
	Pros. cohort	4	43	54.6 $\pm$ 11.9
	Clinical trial	6	12	38.0 $\pm$ 20.6
Col	Total	23	54	32.1 $\pm$ 14.1
	Ret. cohort	12	96	34.6 $\pm$ 15.6
	Pros. cohort	6	22	29.2 $\pm$ 6.2
	Clinical trial	5	12	28.8 $\pm$ 19.2

Abbreviations are Col, continuous infusion; InI, intermittent infusion; Pros, prospective; Ret, retrospective; RSD, relative standard deviation; SD, standard deviation.

Discussion and Conclusion

Differences in exposure variability between InI and Col treatment can largely be explained by the pharmacokinetic profile of vancomycin, which is best described by a two compartment model. After infusion the distribution phase lasts 30-60 minutes, after which equilibrium between the central and the peripheral compartment is reached (11, 42). The vancomycin peak concentration is determined after the distribution phase, in general one to two hours after infusion. The vast majority of vancomycin is cleared through renal excretion, more specifically through glomerular filtration. The degree of clearance thus depends on the glomerular filtration rate, which is estimated in clinical practice by the creatinine clearance (11). Under steady-state conditions with InI, $\Delta C_{peak-C_{trough}}$ is dependent of the dose and volume of distribution as described by equation 1

$C_{peak-C_{trough}} = D/V_d$ (equation 1)

where D and Vd represent the dose and volume of distribution, respectively. Consequently, a relatively high interpatient variability in volume of distribution will result in a high variation in trough levels between individual patients. Besides the volume of distribution, trough concentrations are affected by interpatient variability in clearance. For Col, clearance is the pharmacokinetic parameter that is correlated with C_{ss} as defined by equation 2.

$$C_{ss} = R_{inf}/Cl \text{ (equation 2)}$$

where C_{ss}, R_{inf} and Cl are the concentration in steady-state conditions, the rate of infusion and clearance, respectively. Furthermore, the timing of sampling is of great importance when obtaining trough serum concentrations (figure 3), especially since current IDSA guidelines indicate trough concentrations should be used to monitor efficacy of vancomycin treatment (4).

Vancomycin serum concentrations not drawn directly before the next administration require extrapolation to a true trough serum concentration, which introduces uncertainty and increases variability. In patients on Col, on the other hand, the timing of sampling is of less importance. Plateau concentrations depend on the infusion rate and clearance and therefore variability is caused by variation in clearance only (equation 2). Finally, the variation in duration of the distribution phase in patients on vancomycin InI will not affect trough concentrations, but can hinder accurate and precise estimation of AUC values derived from limited sampling strategies. In patients on vancomycin Col, on the other hand, this effect will be marginal or even absent under absolute steady state condition (43). This is eminent; if in the future vancomycin dosing guided by AUC targets would be preferred.

Independent of the mode of administration, the highest variability of exposure can be expected during the first 24 hours when steady-state conditions are not yet achieved as can be seen in figure 3. Importantly, it has been shown that the correlation between serum concentrations and estimated AUCs is greater in Col compared to InI, rendering plateau concentrations better predictors of vancomycin exposure in terms of AUC (E.M. van Maarseveen, S.G.H. Gipmans, E.C. Vasbinder, M. Petjak and A.R. van Zanten, submitted for publication).

Figure 3. Simulated pharmacokinetic curves of patients treated with intermittent (InI) or continuous infusion (Col) of vancomycin.

The interval between peak (C_{max}) and trough (C_{min}) serum concentration in patients treated with vancomycin InI (dashed line) is described by the dose (D) divided by the volume of distribution (V_d). C_{max} is determined after the distribution phase. The plateau serum concentration (C_{ss}) in patients treated with vancomycin Col (continuous line) is described by the infusion rate of vancomycin (R_{inf}) divided by the clearance (Cl). VAR 1 and VAR 2 illustrate the variability in serum concentration caused by a shift of sampling time.

Studies comparing clinical outcome measures of vancomycin InI and Col treated patients are often underpowered. In this review, an alternate strategy by investigating differences in the variability of exposure between Col and InI was performed. We found that the variability in vancomycin concentrations is larger for treatment with InI vancomycin compared to Col. A 50% greater RSD in serum concentration was detected for InI compared to Col. It is important to note that a 50% larger RSD results in a 2 times larger 95%-confidence interval of trough versus plateau concentrations. Furthermore, the studies that directly compared target attainment of InI and Col found more rapid and higher target attainment with Col (20, 25, 27, 38). Higher variability of exposure with InI compared to Col can be largely explained by the effect of variations in volume of distribution and sample timing. It remains to be elucidated whether the lower variability of exposure and better target attainment associated with Col can improve clinical outcomes.

To the best of our knowledge, this is the first systematic review to address exposure variability in InI and Col vancomycin treatment. Variability in serum concentration may be an important factor in achieving rapid target attainment, which is of critical importance in the treatment of critically ill patients. There are several limitations to this review. First of all, the majority of included RSDs were derived from retrospective cohort studies (32 out of 53 RSDs). Even though this may introduce more variability due to lower protocol adherence, it does represent data obtained in clinical practice. Secondly the heterogeneity of the patient population included could be considered a limitation, since targets and dosing protocols are known to differ between centres. However, in our analysis the relative and not the absolute serum concentration variability was examined, correcting for the different target concentrations in InI and Col treated populations. Finally, systematic reviews are at risk of selection bias. Nevertheless, to our best knowledge all studies reporting vancomycin serum concentration variability in adult patients not on renal replacement therapy were included in this review. Moreover, analysing the data stratified by patient population or methodological design showed that the variability in Col treated patients was lower in relation to InI treated patients in all reported comparisons.

Finally, the definition of target attainment varied largely between the included studies, which hampered the analysis of this outcome measure.

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Chapter 3.2

Exposure variability and target attainment of vancomycin intermittent and continuous infusion in critically ill patients –ready, aim, target locked?

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Abstract

Switching from intermittent to continuous infusion has been proposed to increase target attainment rates. Therefore, we investigated the exposure variability in critically ill patients on vancomycin infusion. A retrospective cohort study was conducted among critically ill patients admitted to a level 2 intensive care unit between April 2007 and November 2013. Vancomycin was administered through intravenous infusion and dosing was guided by therapeutic drug monitoring. In 2010 vancomycin administered was switched from intermittent to continuous infusion. Results showed the serum concentration variability was twice as high in intermittently dosed population compared to continuously infused patients. Importantly, concentration and area-under-the-curve correlations were stronger in patients on vancomycin continuous infusion. In conclusion, switching from intermittent to vancomycin continuous infusion can lower exposure variability and may thereby improve clinical outcomes in critically ill patients.

Introduction

The glycopeptide vancomycin has a time- or area-under-the-concentration-time-curve (AUC)-dependent bacterial killing profile. Based on in vitro and in vivo studies the area-under-the-concentration-time-curve over the minimal inhibitory concentration (AUC/MIC) is considered to be an important predictor of efficacy (1). In critically ill patients large variability in vancomycin distribution and clearance due to pathophysiological changes and concomitant therapies compromise the attainment of predefined exposure targets (2). For instance, fluid resuscitation and total parenteral nutrition expand the extracellular fluid content and dilute serum and extracellular vancomycin concentrations (3, 4). On the other hand, renal function is often impaired in the critically ill causing accumulation of the infused drug (2). To reduce the variability of vancomycin exposure and increase target attainment rates, switching the mode of administration from intermittent (InI) to continuous infusion (CoI) has been proposed (5). In this study we primarily investigated the within-population variability in vancomycin exposure of InI and CoI in critically ill patients. Secondly, vancomycin serum concentrations and AUC correlations and target attainment rates were compared between InI and CoI groups.

Methods

A retrospective cohort study was conducted among critically ill patients admitted to a level 2 intensive care unit between April 2007 and November 2013. Adult patients (18 years or older) treated with intravenous vancomycin for various indications including sepsis, pneumonia and endocarditis were eligible for inclusion. Patients were excluded if treated with any form of renal replacement therapy (RRT) or if vancomycin was prescribed in the three days prior to initiation of therapy. A waiver for consent was provided by the Medical Ethical Committee. Vancomycin was administered through intravenous infusion and dosing was guided by therapeutic drug monitoring (TDM). Before 2010, vancomycin was administered as an intermittent infusion. Vancomycin was started as a 15 mg/kg infusion every 12 hours. First trough (Conc1) and peak (two hours after the end of infusion) serum samples were obtained 24 hours after initiation of therapy and a second set of peak and trough (Conc2) samples were obtained at day 2 or 3 of therapy. The predefined target trough serum concentration for InI was 8-12 mg/L. In 2010, the mode of administration switched from InI to Col. A standard loading dose of 1000 mg infused over 2 hours directly followed by a glomerular filtration rate (GFR) based maintenance dose according to a prospectively validated nomogram (6)). First and second serum plateau samples were drawn approximately 20-36 (Conc1) and 36-72 (Conc2) hours after initiation of therapy, respectively. The target plateau concentration for Col was 15-20 mg/L. Vancomycin serum samples were assayed using a fluorescence polarization immunoassay (TDx, Abbott, Hoofddorp, the Netherlands). AUC_{0-24h} and AUC_{24-48h} were estimated in all individuals using a pharmacokinetic population model and pharmacokinetic software (Appendix 1). Variability in serum concentrations and AUC's was represented by the relative standard deviations (RSD's) as defined by the formula: $RSD = 100 * \text{mean}/SD$, in which SD represents the standard deviation. Subsequently, SD's were compared between InI and Col groups. Secondly, coefficients of determination (r^2) between serum concentrations and AUC's were obtained by univariate linear regression analyses. Finally, target attainment rates of InI and Col were determined for serum concentrations and AUC_{24h} values prior to (Conc1 and AUC_{0-24h}) and after (Conc 2 and AUC_{24-48h}) TDM-based dose adjustment. A 'therapeutic' AUC was defined as an AUC_{24h} ≥ 350 mg.h.L⁻¹ (7, 8). Clinical outcomes could not be

assessed due to the small number of patients.

Results

A total of 113 patients were selected. Five patients were excluded for RRT and 25 for recent prior treatment with vancomycin. Pharmacokinetic data were not available in five patients. Dosing protocol violations excluded seven patients. The remaining 71 patients were included. Baseline characteristics of included patients are shown in Table 1. No significant differences were seen between the two treatment groups. In all occasions the variability in serum concentrations and in AUC's was higher in patients treated with vancomycin InI compared to Col (Table 2). Linear regression analysis showed that coefficients of determination (r^2) were higher and 95%-confidence intervals were nearly twice as large in the InI versus the Col treated population on both sampling occasions (Figure 1). Target attainment rates were significantly higher in the Col versus the InI group for both AUC_{0-24h} (48.0 versus 18.5, $p < 0.001$) and Conc₁ (61.4 vs 14.8, $p < 0.001$).

Table 1 Patient characteristics

	InI (n=27)	Col (n=44)	p-value
Male gender, % (n)	66.7 (18)	65.9 (29)	0.948a
Age (y) (mean±SD)	67.8 ± 12.2	66.6 ± 12.5	0.702b
Weight (kg) (mean±SD)	79.8 ± 19.4	78.4 ± 18.5	0.765b
Height (cm) (mean±SD)	174 ± 9.0	173.9 ± 8.8	0.858b
SAPS II score (mean, range)	41 (12-79)	42 (13-91)	0.790c
Baseline creatinine clearance (ml/min) (mean±SD)	93 ± 52.91	78 ± 37.0	0.11b
Baseline serum creatinine (µmol/L) (mean±SD)	86 ± 38	99 ± 39	0.13b
Baseline serum urea (mmol/L) (mean±SD)	16.3 ± 6.3	14.4 ± 7.3	0.28b

P-values were determined by; a, Pearson Chi-Square test; b, Independent Samples Test; and c Mann-Whitney U Test. Abbreviations are Col, continuous infusion; InI, intermittent infusion; SAPS, simplified acute physiology score; and SD, standard deviation.

Table 2. Exposure variability and target attainment rates

	Intermittent infusion				Continuous infusion				p-value (target attainment)
	Mean	SD	RSD (%)	Target attainment (%)	Mean	SD	RSD (%)	Target attainment (%)	
Conc 1 (mg/L)	9.3	4.6	49.2	18.5	16.4	4.7	28.7	48.0	<0.001
Conc 2 (mg/L)	15.2	6.9	45.1	23.8	21.0	5.2	24.5	33.3	0.471
AUC _{0-24h} (mg.h.L ⁻¹)	289	88	30.4	14.8	365	91	24.9	61.4	<0.001
AUC _{24-48h} (mg.h.L ⁻¹)	475	161	33.3	76.2	476	108	22.7	88.9	0.242

Serum concentrations 1 and 2 were sampled shortly prior to the administration of the third and fifth to seventh administration in the intermittently infused patients. First and second serum samples in continuously infused patients were drawn approximately 20-36 (Conc1) and 36-72 (Conc2) hours after initiation of therapy, respectively. AUC's were estimated using a PK model and Bayesian based PK software. The target plateau concentration for continuous infusion was 15-20 mg/L. Predefined target trough serum concentration for intermittent infusion was 8-12 mg/L. Abbreviations are AUC, area under the concentration-time curve; Conc 1, serum concentration 1; Conc 2, serum concentration 2; Col, continuous infusion; InI, intermittent infusion; RSD, relative standard deviation; and SD, standard deviation.

Figure 1. Serum concentration and AUC Correlations

Serum concentration and AUC Correlations in intermittently infused patients are shown in figures 1A and 1C. Correlations in the continuously infused group are depicted in 1B and 1D. In intermittently dosed patients serum concentrations 1 and 2 were sampled shortly prior to the administration of the third and fifth to seventh administration, respectively. First and second serum samples in continuously infused patients were drawn approximately 20-36 and 36-72 hours after initiation of therapy, respectively. AUC's were estimated using a PK model and Bayesian based PK software. The outer lines represent 95%-confidence intervals. All correlations were significant at a p-level of <0.01.

Discussion

To the best of our knowledge, this is the first study to directly compare exposure variability of vancomycin InI and Col in critically ill patients. The results show lower variability and higher target attainment rates of serum concentrations and AUC's in patients treated with vancomycin Col compared to those treated with InI. Importantly, the correlation between serum concentrations and AUC's was better in the Col compared to the InI group. Consequently, a serum 'plateau' concentration in Col will be a better predictor of patients' exposure in terms of AUC compared to a trough concentration in InI. Since recent guidelines from the Infectious Disease Society of America (IDSA) recommend trough serum concentrations of 15-20 mg/L to attain a minimal AUC_{24h} of 400 mg.h.L⁻¹ (8, 10), we were also interested in the relation between concentrations and AUC's. Although it was not one of the primary aims of our study, we found that trough serum concentrations between 12-16 mg/L resulted in AUC_{24h} values over 400 mg.h.L⁻¹ in vancomycin InI (Figure 1). These findings are supported by recent data published by Neely and coworkers (11). Ultimately, dose adjustments in vancomycin InI based on current trough concentration recommendations render needlessly high exposure in critically ill patients infected by (suspected) pathogens with a MIC ≤ 1 mg/L with an increased toxicity risk as a resultant. The study is primarily limited by its open and retrospective design, although patient baseline characteristics were well balanced. Secondly, the poor performance of the InI regimen with regard to AUC target attainment may result from the fact that this regimen was not designed to target AUC's >350 mg.h.L⁻¹. Our findings can be explained by the fact that variability in 'plateau' concentrations during vancomycin Col is only dependent of variability in clearance, whilst trough concentration variability reflects the variation in clearance and volume of distribution. Moreover, 'trough sample timing' will significantly affect the variability in actually measured concentrations during InI. 'Plateau' concentrations in patients on vancomycin Col, on the other hand, are relatively independent of the sampling time under steady state conditions. (12). In conclusion, we have demonstrated that continuous infusion leads to more robust vancomycin exposure and can beneficially affect AUC target attainment in the critically ill. Whether this advantage leads to improved clinical outcomes warrants further investigations.

Funding

This work was not externally supported. All work was funded from the departments' research budgets.

Transparency declarations

None of the authors has conflicts of interest to declare.

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Appendix 1. Brief description of the design and validation of the pharmacokinetic model

Manuscript title: 'Exposure variability and target attainment of vancomycin intermittent and continuous infusion in critically ill patients – ready, aim, target locked?'

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Model design

Data were collected from patient clinical records and the electronic prescription database at the GHZ. A pharmacokinetic population model was designed using the KinPop module of MW\Pharm (version 3.80, MediWare, Zuidhorn, the Netherlands) for each of the groups using the trough and peak serum concentration obtained from patients that received vancomycin through intermittent infusion. MW\Pharm uses an iterative two-stage Bayesian approach to estimate the mean and coefficient of variation of relevant pharmacokinetic parameters. Population models were designed and validated according to previously published methodology (14). The following polynomial formula was used to estimate the SD (standard deviation) of the analytical assay error: $SD = 0.323 + 0.0265 * C + 0.0002 * C^2$. A two-compartment model was designed. The elimination rate constant was defined by the formulae: $Kel_{\text{total}} = K_{\text{elm}} + K_{\text{elr}}$ and $K_{\text{elr}} = K_{\text{slope}} * Cl_{\text{cr}}$, in which K_{elm} , K_{elr} and K_{slope} represent the metabolic elimination constant, renal elimination constant and slope constant, respectively. The free fraction of vancomycin was assumed to be constant at 0.5 ± 0.1 in the model.

The model was evaluated by bootstrap analysis (n=1000 replicates), an analysis of individual predicted versus observed concentrations, an analysis of the residual error versus the observed concentrations and model fit was checked visually for all individual patients. A pharmacokinetic model was established. Bootstrap analysis resulted similar parameters estimates (Table A1). Predicted versus observed serum concentrations plot showed good correlation (r^2 0.88, $p < 0.01$) and weighted residuals were randomly distributed over the concentration range (Figure A1). Visual check of all individual concentration-versus-time-curves showed good individual fits.

	PK model parameters		Bootstrap estimates	
	Estimate	CV (%)	Mean	RSE (%)
Kelm (h-1)	0.017	43	0.019	56
Kslope (h-1/(ml min-1))	0.0016	31.4	0.0015	17.1
V1 (L/kg)	0.47	18.4	0.51	14.6
K12 (h-1)	2.9	51.0	3.2	48.0
K21 (h-1)	1.1	106.5	0.9	40.4

Table A1. Pharmacokinetic population estimates and bootstrap analysis results of InI model in critically ill patients. Abbreviations: ICU, intensive care unit; InI, intermittent infusion; Kelm, metabolic elimination rate constant; Kslope, renal elimination rate constant per ml/min clearance; PK, pharmacokinetic; CV, coefficient of variation; RSE, relative standard error; V1, volume of distribution in the central compartment.

Figure A1. Plot A shows the individual predicted versus observed concentrations of the pharmacokinetic population model. In plot B the weighted residuals over the measured concentrations are depicted.

Chapter 3.3

Design and Prospective Validation of a Dosing Instrument for Continuous Infusion of Vancomycin: A Within-population Approach

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Eur J Clin Pharmacol. 2014 Aug 30. [Epub ahead of print]

Abstract

The clinical application of continuous infusion (Col) of vancomycin is rapidly gaining interest. Since no international guidelines on initial dosing of vancomycin Col exist, there is an urgent need for methods to facilitate the switch from intermittent to continuous vancomycin dosing algorithms in clinically infected populations. Therefore, the aim of this study was to describe the design and validation of an a priori dosing schedule for Col of vancomycin in clinical practice. A dosing table for Col of vancomycin based on estimated glomerular filtration rate (eGFR) was developed by simulation of continuous infusion of vancomycin using pharmacokinetic (PK) software and a PK population model designed from historical within-population data in intermittently dosed patients. The target range for steady state concentration (C_{ss}) at 24 hours was set at 15-20 mg/L corresponding with an area-under-the-curve (AUC) of at least 350 mg·kg·L⁻¹. The performance of the dosing schedule was primarily assessed by describing the percentages of patients attaining the predefined target. An eGFR-derived dosing schedule for Col of vancomycin was established and implemented in clinical practice. Prospective assessment in 35 general ward and 45 intensive care unit patients showed that AUC target was attained in 80% and 72% of patients and C_{ss} target was reached in 63% and 69%, respectively. An easy method to design and validate an eGFR-derived dosing algorithm for the continuous infusion of vancomycin to switch from intermittent to continuous dosing of vancomycin is made available.

Introduction

Vancomycin is a glycopeptide antibiotic used in (suspected or proven) bacteraemia, peritonitis or osteomyelitis with gram-positive micro-organisms. Antimicrobial killing of vancomycin is time and/or area-under-the-curve dependent and requires therapeutic concentrations of vancomycin at the site of infection (1, 2). Although resistance against vancomycin and other glycopeptides is emerging worldwide among methicillin-resistant *Staphylococcus aureus* and vancomycin-intermediate *S. aureus* (VISA) strains (3), vancomycin still is the antibiotic of choice for the therapy of methicillin-resistant *S. aureus* (MRSA) infections according to recent guidelines from the Infectious Diseases Soci-

ety of America (IDSA) (4). To preserve the current therapeutic armamentarium for Gram-positive infections, it is of paramount importance that guidelines for vancomycin dosing are based on patient outcomes and on in vitro and in vivo pharmacokinetic/ pharmacodynamic (PK/ PD) data to attain maximum efficacy with minimal toxicity and prevent the advent of resistance. Albeit there is in vitro evidence showing that the time above the minimal inhibitory concentration (MIC) and AUC/ MIC-values are the most important parameters associated with efficacy, most human studies failed to detect a clear PK/PD relationship (2). To date, only Moise-Broder et al. showed that for intermittent infusion of vancomycin an 'in vivo' AUC/MIC-ratio of <350 was associated with a higher cure rate ($P = 0.005$) in patients with MRSA pneumonia (1). Classically, vancomycin is administered as an intermittent infusion of 15-20 mg/kg every 12 hours. (5). Unfortunately, ascertaining AUC/ MIC values $>350 \text{ mg} \cdot \text{kg} \cdot \text{L}^{-1}$ and concentrations above MIC shortly after initiation of therapy can be difficult with conventional intermittent dosing in combination with trough level monitoring. To optimize the pharmacodynamics of vancomycin higher intermittent daily dosages have been suggested (6-8). Nevertheless, even with higher intermittent dosing attaining therapeutic exposure remains a challenge in patients with rapid clearance of vancomycin. Hence, it seems rational to explore alternative dosing strategies of vancomycin, e.g. continuous infusion (Col). Recent data suggest that administration of vancomycin for the treatment of Gram-positive infections by Col is associated with a significantly lower risk of nephrotoxicity compared to intermittent infusion, while efficacy parameters were comparable (9-11). In addition, continuous infusion of vancomycin is feasible due to demonstrated stability at temperatures up to 37°C for 72 hours (12) and might also be preferred due to a reduction in drug preparation time, staff workload, costs and hospital length of stay (13, 14).

Gradually, more data on the therapeutic range of Col are generated. Based on the pragmatic approach of achieving AUC's equal to intermittent dosing a minimum plateau level of 15 mg/L corresponding with an AUC_{24h} of $>350 \text{ mg} \cdot \text{kg} \cdot \text{L}^{-1}$ most studies used 15-25 mg/L as a target steady state concentration (C_{ss}) (15-19). These studies showed microbiological and clinical efficacy rates in the Col study arm that were equivalent or better than the study arm receiving vancomycin via intermittent infusion. Only scarce data on the correlation between plateau levels and rates of nephrotoxicity exist. Studies performed by Spapen et al. and Ingham et al. showed plateau levels exceeding 28-30 mg/L

were significantly associated with nephrotoxicity both in critically ill and outpatients populations (20, 21). For comparison, serum trough concentrations over 15-20 mg/L during intermittent dosing of vancomycin have been associated with an increased risk of nephrotoxicity (22, 23). As the clinical application of Col of vancomycin is rapidly gaining popularity and no international guidelines exist, there is an urgent need for methods to design dosing algorithms for continuous vancomycin dosing algorithms aiming for predefined AUC/MIC exposure targets in clinically infected populations. Therefore, the aim of this study was to describe the design and validation of a practical and easy to apply dosing schedule for vancomycin Col in clinical practice.

Methods

Study design

Prior to prospective validation a dosing table for Col of vancomycin was developed by simulation of continuous infusion of vancomycin using the PK software package MW\Pharm (version 3.60, MediWare, Zuidhorn, The Netherlands) (24) and a pharmacokinetic (PK) population model based on historical within-population data of patients on intermittently dosed vancomycin. The PK model was validated using a previously described methodology (25) (Appendix 1). The parameters of the PK model were: V_1 , 0.29 L/kg; k_{12} , 0.92 /h and k_{21} , 0.46 /h; K_{slope} , 0.00372 /h/(ml/min) and K_{elm} , 0.018 /h. The elimination rate constant was defined by the formulae: $K_{el} = K_{elm} + K_{elr}$ and $K_{elr} = K_{slope} * Cl_{cr}$, in which K_{elm} , K_{elr} and Cl_{cr} reflect the metabolic rate constant, renal elimination slope constant and creatinine clearance, respectively. The free fraction of vancomycin was assumed to be constant in the model. Vancomycin clearance is the key parameter for prediction of C_{ss} . The volume of distribution (V_d) is required to accurately determine the total serum AUC of the first 24 hours after start of infusion in individual patients. Patients with an estimated glomerular filtration rate (eGFR) based on the modification of diet in renal disease (MDRD) formula ranging from 10 to 150 mL/min/1.73 m² were simulated with intervals of 5 mL/min (26). Simulated patients' weights ranged from 50-100 kg. A target range for C_{ss} at 24 hours of 15-20 mg/L corresponding with an AUC/MIC ratio of >350, was set to determine a fixed loading dose and individualized maintenance dosages within a range of eGFR based on MDRD and simulated weights.

For the prospective validation adult patients admitted to a 400 bed teaching hospital starting with Col of vancomycin were eligible for inclusion. A waiver was provided by the medical ethical committee. Patients were excluded if they were on renal replacement therapy (RRT) or if the administered loading or maintenance dose deviated from the dosing table. Vancomycin clearances and AUC's of the first 24 hours after initiation of therapy were estimated in all patients with maximum a posteriori Bayesian estimation (MW\Pharm 3.60). A two compartment open PK model was used. Accuracy and precision of the dosing table were assessed by calculation of the percentages of patients reaching a C_{ss} of 15-20 mg/L or an AUC >350 mg·kg·L⁻¹. Finally, agreement between predicted and observed C_{ss} values was analyzed by testing correlation and by assessment of bias, means and 95% confidence intervals (95%CI) of observed versus predicted values.

Vancomycin and creatinine assays

Serum concentrations of vancomycin were drawn 18-28 hours after initiation of infusion, when steady state was assumed in the majority of patients. A second vancomycin serum concentration was drawn 72 hours after initiation of therapy. Samples were analyzed using a fluorescence polarization immunoassay (TDx, Abbott, Hoofddorp, The Netherlands). The lower limit of quantification and intraday and between-day coefficients of variation for vancomycin were 0.7 mg/L and 0.8 and 1.2 %, respectively. The linearity (r²) of the assay was 0.998. Baseline serum creatinine was analyzed using a colorimetric method (DxC, Beckman Coulter Inc., Brea, CA, USA). GFR was estimated using the modified diet in renal disease (MDRD) formula and serum creatinine values drawn within 24 or 48 hours before the initiation of vancomycin in the intensive care unit and general ward population, respectively.

Statistical analysis

As the PK parameters of the intensive care unit and general ward populations were expected to differ, patients were divided into 2 groups. Data were tested for normal distribution and variance homogeneity with Kolmogorov–Smirnov and Levene tests. Means and standard deviations of continuous variables are presented in normal distributions. Otherwise, medians and ranges were reported. A Student's t-test was used to determine statistical differences among normally distributed groups. If data were not normally distributed, a Mann-Whitney-U test was performed. A Chi-square test was used to compare dichotomous variables between groups. Predicted vs. observed C_{ss} levels at 18–28 hours after initiation of therapy were compared using Pearson correlation. Univariate and multivariate analyses of patient characteristics were performed to identify determinants of underexposure (AUC <350 mg·k·g⁻¹·L⁻¹ and C_{ss} level <15 mg/L) by binary logistic regression. A stepwise backward elimination procedure was carried out, retaining covariates in the model only when they were statistically significant. To account for multiple testing, only those covariates were tested that were shown to affect the risk of underexposure in univariate analyses and a stringent P value (<0.005) was selected. To evaluate the effect of renal function on exposure, patients were divided into 4 groups based on data distribution of MDRD-derived eGFR: MDRD <40 ml/min/1.73m² (n=17), 40 ≤ MDRD <60 ml/min/1.73m² (n=15), 60 ≤ MDRD <90 ml/min/1.73m² (n=19), and MDRD ≥90 ml/min/1.73m² (n=29). To test the effect of body weight on achieving therapeutic exposure were divided into quartiles based on data distribution. Statistical significance was defined by P-values <0.05. All tests were performed in SPSS Statistics version 20 (SPSS Inc., Chicago, IL, USA).

Results

A dosing table for Col was established (Table 1).

Table 1. Dosing table for vancomycin continuous infusion

eGFR ^a (ml/min/1.73m ²)	Dose ^{b,c} (mg/24 h)
<30	500
30-39	750
40-49	1000
50-59	1250
60-79	1500
80-89	1750
>90	2000

a glomerular filtration rate estimated with MDRD formula
b target C_{ss}: 15-20 mg/L
c preceded by a loading dose of 1000 mg administered over 2 hours

In total 103 patients were treated with Col of vancomycin between January 2009 and August 2012. Ten patients were excluded for dosing protocol violations and 13 patients were excluded of reasons of RRT. Patient characteristics are shown in Table 2.

Table 2. Baseline characteristics

	Population		p-value
	General ward	Intensive care unit	
No. patients	35	45	
Wards (%)			
<i>Surgical</i>	47	n.a.	
<i>Medical</i>	53		
SAPS-II score	n.a.	41 (12-79)	
age (y)	71 (12.4)	70 (12.3)	0.95 ^a
male gender (%)	54	58	0.36 ^b
weight (mg)	75.9 (14.3)	76.6 (15.8)	0.83 ^a
MDRD (mL/min/1.73m ²)	65 (13 - >90)	72 (24 - >90)	0.60 ^c

Baseline characteristics of included patients are shown. Differences in categorical variables were expressed as numbers and percentages, and continuous variables as means and standard deviations if normally distributed or as medians and range between parentheses if not normally distributed. Abbreviations are: eGFR, glomerular filtration rate estimated with MDRD formula; n.a., not applicable.

a Student's t-test,

b Chi-squared test,

c Mann-Whitney-U test

All eligible patients who received vancomycin demonstrated infections caused by documented or presumed vancomycin sensitive pathogens. Included vancomycin samples were drawn 18-28 hours after initiation of therapy. The dosing table produced good accuracy and precision of observed C_{ss} levels in intensive care unit and general ward patients (Figure 1 and Table 3).

No significant differences were found in mean C_{ss} levels at 24 hours, AUC's and clearance of vancomycin between the intensive care unit and general ward populations (Table 3).

Figure 1: Frequency distribution of vancomycin serum concentrations and total serum area-under-the-curves

Box-and-whisker plots of vancomycin total serum concentrations at 24 hours after initiation of therapy (1A) and total serum area-under-the-curves (AUC's) on day 1 of therapy (1B) are displayed for the general ward and intensive care unit populations. The line in the middle of each box represents the median; the box extends from the 25th to 75th percentile (interquartile range); and the lines emerging from the box extend to three quarters of the interquartile range, rolled back to where data are present. The shaded area in Figure 1A indicates the therapeutic target concentration range (15-20 mg/L). The interrupted line in Figure 1B indicates the minimal AUC target of 350 mg·h·L⁻¹.

Table 3 Dosing and pharmacokinetic parameters

	Population		p Value
	General ward	Intensive care unit	
Initial Dose (mg)	1500 (500-2000)	1500 (500-2000)	0.35c
Sampling time (h)	22.5 (4.2)	25.8 (3.1)	0.76a
C _{ss} (mg/mL)	17.4 (5.0)	17.4 (5.3)	0.94a
Cl _{vanco} (L/h)	3.6 (1.7)	3.6 (0.9)	0.61a
Total serum			
AUC _{0-24h} (mg*h/L)	427 (107)	415 (112)	0.63a
Dose adjustment (%)	33	39	0.81b

Dosing and pharmacokinetic parameters of included patient are presented. Differences in categorical variables were expressed as numbers and percentages, and continuous variables as means and standard deviations if normally distributed or as medians and range between parentheses if not normally distributed. Abbreviations are: C_{ss}, plateau serum concentration; Cl_{vanco}, vancomycin clearance; AUC_{0-24h}, area-under-the-concentration-time-curve of the first 24 hours.

a Student's t-test

b Chi-squared test

c Mann-Whitney-U test

Analysis of prospective data showed that rates of patients with a C_{ss} level >15.0 mg/L were 69 and 63 % and an AUC >350 mg·kg·L⁻¹ were 72 and 80 % and for the intensive care unit and general wards populations, respectively. A significant correlation ($r^2 = 0.35$, $p = 0.02$) between predicted and observed C_{ss} levels was found. Mean predicted (18.9 mg/L, 95%CI 18.6-19.3) and observed (17.4 mg/L, 95%CI 16.2-18.9) values were comparable. Mean bias was -1.2 mg/L (95% limits of agreement 0.50-1.88). In both groups one patient had a drug level over 28.0 mg/L. Finally, multivariate binary logistic regression analysis showed that an MDRD <40 (OR = 3.8 [1.2-11.8], $p < 0.001$) was the only parameter predictive of underexposure. After dosage correction based on linear pharmacokinetics 89 % of patients' C_{ss} levels at 72 hours were within the predefined target range.

Discussion

Ideally, a 'population-tailored' dosing algorithm for continuous vancomycin dosing is designed and prospectively validated based on historical within-population PK data. Therefore, the primary aim of our study was to describe an easy method to design and prospectively validate an algorithm for initial dosing of vancomycin CoI to facilitate the switch from intermittent to continuous dosing. Albeit the presented dosing table is in good agreement with the only other eGFR-derived dosing nomogram published by Pea et al. (17), we advice clinical centers to prospectively validate any dosing algorithm adopted from literature and recommend designing a 'population-tailored' algorithm if historical data and pharmacological knowledge are available. In addition to the data presented by Pea et al. we evaluated target attainment using the AUC as exposure parameter, since it is believed to be the major predictor with regard to efficacy. Furthermore, the nowadays more common MDRD formula was selected to estimate GFR instead of the Cockcroft&Gault formula. Remarkably, but also in accordance with results reported by Pea et al. the performance of the dosing nomogram with regard to reaching target serum concentrations was comparable between intensive care unit and general ward populations. Target AUC attainment rates were over 70% in both populations, therefore in the remaining 30% of patients exposure could be considered suboptimal. Yet, 100 and 90% of patients had an exposure above 200 and 300 mg*h/L respectively (Figure 1). Vancomycin will most likely be able to inhibit the growth of most pathogens at this exposure level during a short time period. Swift dose adjustments will optimize exposure and assure efficacy.

To rapidly achieve target trough concentrations and high tissue levels recent IDSA guidelines recommend a loading dose of 25-30 mg/kg in serious MRSA infections, including pneumonia and bacteremia (4). With respect to this recommendation our loading dose used may be too low and could be considered a limitation of the presented study next to scarce PK sampling, assumed steady state and stable vancomycin clearance, which are shortcomings faced by the majority of studies in this field and difficult to avoid due to the observational design. However, the loading dose recommendations of the IDSA guidelines are supported by limited safety data and further evaluation, primarily of the nephrotoxic effect is warranted (28).

The nephrotoxic potential of enhanced dosing of vancomycin is un-

known for patients with severe infections who are at risk of acute kidney failure. To our knowledge the only published PK data on these intensive dosing protocols of Col of vancomycin in a clinical study have been published by Roberts et al. (29). They found a 35 mg/kg loading dose followed by high maintenance dosages of 7-14 mg/kg/24h was necessary to rapidly achieve 20 mg/L vancomycin concentrations in intensive care unit patients. Nevertheless, the proposed dosing protocol was based on data generated by Monte Carlo simulations and has not been prospectively validated. Recently, Ampe et al. and Schmelzer et al. tested dosing nomograms for vancomycin continuous infusion aiming for plateau concentrations between 25-30 mg/L using higher loading doses in patients suffering from difficult-to-treat infections and 20-30 mg/L in critically ill patients respectively. They found that nephrotoxicity potentially related to vancomycin was observed in up to 25% of patients. Further research on the relation between vancomycin exposure and nephrotoxicity is urgently needed. For now the risk-benefit-ratio of 'enhanced' vancomycin dosing remains to be elucidated. When our study was designed, the IDSA guidelines had not been published yet, and a 1000 mg loading dose for a patient of 70 kg (15 mg/kg) was considered adequate at that moment (19, 30). Finally, our target C_{ss} levels were set at 15-20 mg/L, since in our center vancomycin is primarily used to target micro-organisms with MIC's in the lower range, which is not uncommon in Northwestern European countries. Globally, vancomycin is frequently used again MRSA with higher MIC's urging for higher C_{ss} levels. Using the here presented methodology this can readily be achieved by elevating the target levels during the design and validation of any dosing algorithm.

Previous studies have shown that the MDRD tends to be less accurate in the higher and lower GFR range (26), which may explain why patients with an MDRD <40 mL/min are at higher risk of underexposure. However, its accuracy and precision are comparable to eGFR values estimated with the Cockcroft & Gault formula (31). Levey and co-workers showed the MDRD formula can provide a good estimate of GFR values from 60 to 90 mL/min/1.73m² with just slightly higher bias (2.5 versus 5.5 mL/min per 1.73 m²) compared to the Chronic Kidney Disease Epidemiology Collaboration (CKD-EPI) equation, which is currently considered to provide the most accurate GFR estimate based on serum creatinine (32).

The MDRD formula was selected, since there is worldwide trend in clinical information systems automatically to report MDRD-derived eGFR

in patients' clinical records by clinical chemistry departments (33, 34). The major advantage of the MDRD over e.g. Cockcroft & Gault formula is that it does not require the input of data on the patient's weight and therefore can be automatically calculated from patient's age, gender, race and serum creatinine. Naturally, the MDRD formula also has its shortcomings. It tends to overestimate the actual creatinine clearance in patients that are bedridden and is based on a fixed body surface area of 1.73m². A secondary explanation for higher percentage of patients with MDRD values in the lower range not reaching the predefined exposure targets is provided by Rodvold et al. who showed that the apparent volume of distribution per kg body weight in patient with an impaired renal function was proximately 30% higher increasing the risk of underexposure (35). Still, as shown by De Waele et al. a weight-based loading dose followed by a standard maintenance dose results in reasonable target attainment rates in critically ill patients (36). In theory an a priori dosing strategy of a weight-based loading dose followed by an eGFR maintenance dose has the best chance of reducing interpatient exposure variability and warrant testing in future studies (37, 38). In the near future serum cystatin C concentration may serve as a better predictor of vancomycin clearance (39).

Apart from the ongoing debate on optimal vancomycin dosing and levels in clinically infected patients on Col, it is important to note that the IDSA expert panel advises against administration of vancomycin as continuous infusion given that a clear benefit in clinical outcomes over intermittent dosing has not been demonstrated (4). In contrast, based on PK/PD considerations Col may be the preferred mode of administration, whilst it is widely accepted that efficacy is best predicted by the AUC/MIC model. Considering vancomycin C_{ss} levels during Col can easily be converted to AUC values under steady state conditions and are associated with little AUC_{24h}-variability as opposed to trough level monitoring in intermittent dosing. This straightforward translation of C_{ss} to AUC's is, however, not applicable to the first 24 hours following initiation of Col, since then AUC values in individual patients are strongly affected by the loading dose and the volume of distribution. This is the main reason for using a PK model instead of an eGFR-derived linear algorithm to design the dosing table.

For current clinical practice, we propose 2 strategies depending on local availability of TDM services to obtain optimal exposure. If a TDM service is available, we advice to draw samples 2 and 8-14h after initiation of therapy. Using a PK model and Bayesian based PK soft-

were the plateau concentration at 24h as well as the AUC_{0-24h} can be estimated and the dose can swiftly be adjusted. If these PK 'tools' are not present, we advice to draw the first sample approximately 24 hours but not later than 36h after initiation of therapy balancing between rapid dose adjustment and reaching steady state conditions.

Conclusions

An easy method to design and validate an eGFR-derived dosing schedule for continuous infusion of vancomycin was presented. Prospective validation showed good precision and accuracy in clinical practice.

Funding

This work was not supported by outside funding. All work was funded by our department research budget.

Transparency declarations

None of the authors has conflicts of interest to declare.

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Appendix 1. Brief description of the design and validation of the pharmacokinetic model

Manuscript title: 'Exposure variability and target attainment of vancomycin intermittent and continuous infusion in critically ill patients – ready, aim, target locked?'

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Model design

Data were collected from patient clinical records, therapeutic drug monitoring (TDM) database and the electronic prescription database. A pharmacokinetic population model was designed using the KinPop module of MWPharm (version 3.60, MediWare, Zuidhorn, the Netherlands) and historical within-population data obtained from 18 surgical ward patients, 22 internal ward patients and 55 intensive care unit patients on vancomycin intermittent infusion. MWPharm uses an iterative two-stage Bayesian approach to estimate the mean and coefficient of variation of relevant pharmacokinetic parameters. Population models were designed and validated according to previously published methodology [1]. The following polynomial formula was used to estimate the SD (standard deviation) of the analytical assay error: $SD = 0.323 + 0.0265 * C + 0.0002 * C^2$. A two-compartment model was designed.

The elimination rate constant was defined by the formulae: $Kel = Kelm + Kelr$ and $Kelr = Kslope * Clcr$, in which Kelm and Kelr represent the metabolic and renal elimination constant, respectively. The glomerular filtration rate of vancomycin was assumed to be constant. The model was evaluated by bootstrap analysis (n=1000 replicates), an analysis of individual predicted versus observed concentrations, an analysis of the residual error versus the observed concentrations and model fit was checked visually for all individual patients. A pharmacokinetic model was established. Bootstrap analysis resulted similar parameters estimates (Table A1). Predicted versus observed serum concentrations plot showed good correlation ($r = 0.87, p < 0.01$) and weighted residuals were randomly distributed over the concentration range. Visual check of patients' concentration-versus-time-curves showed good individual fits.

PK model parameters			Bootstrap estimates	
	Estimate	CV (%)	Mean	RSE (%)
Kelm (h ⁻¹)	0.018	225	0.022	278
Kslope (h ⁻¹ /(ml min ⁻¹))	0.0037	31	0.0034	17
V1 (L/kg)	0.29	29	0.32	15
K12 (h ⁻¹)	0.92	47	0.61	15
K21 (h ⁻¹)	0.46	116	0.38	45

Table A1. Pharmacokinetic population estimates and bootstrap analysis results of InI model in critically ill patients. Abbreviations: ICU, intensive care unit; InI, intermittent infusion; Kelm, metabolic elimination rate constant; Kslope, renal elimination rate constant per ml/min clearance; PK, pharmacokinetic; CV, coefficient of variation; RSE, relative standard error; V1, volume of distribution in the central compartment

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Chapter 4

Summary, clinical implications and future perspectives

In this thesis we have investigated pharmacokinetically based strategies for optimization of aminoglycoside and vancomycin therapies. The current chapter will summarize and discuss the results of our studies investigating aminoglycoside and vancomycin pharmacokinetics in the studied patient populations with infections.

Once daily and extended interval dosing of aminoglycosides

As once daily dosing of aminoglycosides has gained popularity worldwide, more data are available on once daily dosing regimens in various clinically infected populations. In the study described in Chapter 2.1 we focused on adult cystic fibrosis (CF) patients. CF patients are frequently exposed to high doses of once daily dosed (ODD) tobramycin with relatively long treatment course duration in case of pulmonary exacerbations (1). The tobramycin dosages range from 10 to 15 mg/kg once daily in this population. Based on pharmacokinetic/pharmacodynamic (PK/PD) considerations a high dose per kg bodyweight is a consequence of the relatively large volume of distribution and high clearance in comparison to other clinically infected populations treated with aminoglycosides (2). Current once daily high tobramycin dosing protocols in pediatric and adult CF patient populations result in high peak and low trough concentrations ensuring efficacy and limiting the risk of toxicity, respectively (3). In Chapter 2.1 the influence of the time of administration on the PK of tobramycin in adult CF patients was investigated. This study was designed to test the assumption that dosing in the evening results in a lower clearance and an increased area under the concentration-time curve (AUC) of aminoglycosides due to the circadian rhythm of renal function. In close collaboration, the effect of the circadian rhythm on tobramycin PK was studied in pediatric CF patients by investigators at the University of Nottingham, UK. Results from the latter studies indicate no clinically relevant effect of the administration time on tobramycin PK in CF patients. In Chapter 2.2 we performed a retrospective cohort study that investigated the effect of the time of administration of aminoglycosides in general ward patients and in intensive care unit (ICU) patients. In these clinically infected populations also no effects of the time of administration could be detected on the PK of aminoglycosides. Furthermore, the incidence of nephrotoxicity was not affected by the time of administration in the general ward population. We suggest that the lack of an effect of the time of administration on

aminoglycoside PK in CF patients, general ward patients and ICU patients is caused by a distortion of the 'normal' circadian rhythm in these clinically infected patients. Previous reports have demonstrated that changes in immunological status affect the circadian rhythm, e.g. sepsis will abrogate the normal melatonin secretion over the day (4-6). The presence of a 'normal' circadian rhythm is influenced by distortion of immunological homeostasis. In fact, based on our observations we hypothesize that intravenous antibiotic agents will rarely show a diurnal pattern since they are prescribed to patients with serious suspected or proven infections.

In Chapter 2.3 we investigated the PK and nephrotoxicity of ODD gentamicin and ODD tobramycin in a large retrospective cohort of infected general ward patients. The most important finding of this study was a fifty percent lower incidence of nephrotoxicity in patients treated with tobramycin compared to those treated with gentamicin. To our knowledge no previous studies have compared the clinical toxicity of two or more ODD aminoglycosides. Importantly, this study offers a pharmacological explanation for the difference in nephrotoxicity between gentamicin and tobramycin, since the PK analysis showed that gentamicin clearance was lower than tobramycin clearance. As a consequence, gentamicin exposure in terms of daily AUC was higher compared to tobramycin exposure. Mechanistically, our findings can be explained by the higher renal accumulation of gentamicin versus tobramycin inducing renal injury (7). Finally, a multivariate analysis showed that the AUC of the first administration was a strong predictor of the incidence of nephrotoxicity indicating 'early therapeutic drug monitoring (TDM)' using limited AUC sampling can be of additive value identifying those patients at risk and therefore is of clinical relevance.

In Chapter 2.4 we prospectively evaluated a gentamicin dosing regimen which is used in newborns with suspected or proven neonatal sepsis. This study showed that a dosing regimen of 5 mg/kg every 36h resulted in therapeutic exposure of over 90% of included subjects. It is important to note that this study was not performed in a neonatal intensive care unit (NICU) but in a level 2 special care nursery population. In accordance with previous reports (8), we found a strong relationship between the volume of distribution (L/kg) of gentamicin with body-weight in our population, with a larger bodyweight-corrected volume of distribution in preterm infants due to differences in body composition. Gentamicin half-life decreased with lower gestational age, which has also been reported by others (9). A combination of low weight-corrected

volume of distributions with short half-lives will translate in concentration-vs-time-curves with high peak concentrations and steep slopes. Vice versa, in low-birth-weight premature newborns peak concentrations and half-lives were lower and longer, respectively. Interestingly, the AUC was not related to gestational age or body weight, meaning that using the proposed dosing regimen in low birth-weight more premature newborns on average had the same gentamicin exposure compared to older neonates with higher bodyweights.

Continuous infusion of vancomycin

In Chapter 3.1 we performed a review of the literature comparing the inter-patient exposure variability in clinically infected patients on continuous or intermittent infusion of vancomycin. This review showed that continuous infusion conferred a fifty percent lower inter-patient variability of vancomycin concentrations compared to intermittent infusion. This interesting finding based on the review of in total 34 published reports led to the design of a retrospective cohort study in an ICU population (Chapter 3.2). In this study we confirmed the hypothesis that the inter-patient variability in vancomycin exposure was significantly lower for continuously dosed versus the intermittently dosed critically ill patients. Furthermore, 'plateau' concentrations in continuous dosing showed a stronger correlation with estimated AUCs compared to trough concentrations during intermittent dosing. Two conveyable causes can largely explain the reported results in Chapter 3.1 and 3.2. First, since 'trough sample timing' can have a major influence on the measured vancomycin concentrations, an intermittent dosing regimen monitored by trough concentration is inherently prone to higher variability in comparison to continuous infusion and 'plateau' concentration monitoring (10). Second, when trough concentrations are monitored in case of intermittent dosing the variability of the AUC is not only affected by the vancomycin clearance, but also by the volume of distribution (Chapter 3.1 and 3.2). Patients with a relatively high volume of distribution will need higher intermittent dosages to maintain trough concentrations within the therapeutic range resulting in a large 'delta' between peak and trough concentrations and therefore in high AUCs, especially in once or twice daily dosing regimens. In Chapter 3.3 a simple method was presented to clinically implement a vancomycin continuous dosing nomogram based on historical within-population data obtained from

intermittently dosed patients. This chapter offers tools for clinicians considering a switch from intermittent to continuous dosing of vancomycin. Although not intended for use in other clinically infected populations the developed dosing nomogram was in accordance with the only other prospectively validated dosing algorithm (11).

Implications for clinical practice

In chapters 2.1 and 2.2 we showed that the time of administration did not affect the clearance or exposure in terms of AUC in CF patients, general ward patients and ICU patients. Hence, swift administration times to the morning period is not recommended and 24 hour intervals should be maintained unless therapeutic drug monitoring results indicate that the dosing interval needs to be extended (12). Furthermore, since early initiation of antibiotic treatment is paramount to improve outcomes, the first administration of an aminoglycoside should take place as soon as possible after the diagnosis (13).

Although our findings deserve to be confirmed, results from Chapter 2.3 show a higher nephrotoxic potential of ODD gentamicin over ODD tobramycin in clinically infected patients treated for at least 3 days which may lead to the revision of (inter)national and local treatment protocols. In patients who are at high risk of developing nephrotoxicity and/or are likely to be treated over a longer period, tobramycin should be preferred over gentamicin. In fact, European cystic fibrosis treatment guidelines already advocate to choose tobramycin over gentamicin for this reason (14, 15). The same concerns with regard to cumulative toxicity apply to other highly exposed populations, e.g. patients treated for endocarditis. In order to optimize treatment with aminoglycosides 'early' AUC monitoring in addition to 'trough levels' should be implemented in daily routine, since our data suggest the AUC of the first administration is a strong predictor of nephrotoxicity.

The added value to the present literature of our study results presented in Chapter 2.4 on gentamicin dosing in neonates lies in the specific neonatal population investigated. Current dosing regimens in neonates are largely based on data obtained from neonates admitted to a NICU (9, 16, 17), while the number of neonates exposed to gentamicin is likely to be higher in level 2 special care nurseries. Albeit our data deserve to be prospectively confirmed in a second independent cohort of neonates admitted to a level 2 special care nursery, our findings show that a more uniform dosing protocol of 5 mg/kg per 36 hours can be implemented in level 2 unit compared to more complex dosing protocols proposed in neonates admitted to a level 3 intensive care unit (16, 18, 19). As described in Chapter 3.1 vancomycin continuous dosing

protocols are rapidly gaining interest and are being implemented in clinical practice. Apart from some practical advantages over intermittent infusion, continuous infusion of vancomycin shows a more predictable exposure and thus the odds of an optimal target attainment as shown in Chapter 3.1 and 3.2 are better. In addition, continuous infusion of vancomycin might also be preferred since the drug preparation time, the staff workload and costs are reduced and it may also shorten hospital length of stay.(20, 21), (22) and has shown to be safe concerning stability (23). Of course, there are also potential disadvantages to continuous infusion. Sufficient numbers of volume controlled infusion pump systems have to be secured and compatibility with other simultaneously infused drugs can be an issue. Nevertheless, 'Y-site'-compatibility of vancomycin with many drugs has been demonstrated (24). Finally, based on our methodology described in Chapter 3.2 a 'tailored' dosing algorithm of vancomycin can be designed for clinically infected populations aiming to achieve a priori target exposure using historical within-population data, a PK model and PK software.

Suggestions for future studies

Toxicity studies in populations treated with 'traditional' dosing regimens of aminoglycosides or vancomycin have been performed in the past. However, limited data are available on the recommended dosing regimens from most recent guidelines with respect to clinical outcomes (25, 26). For example, the most recent treatment guidelines for the treatment of multi-resistant *S. aureus* (MRSA) of the Infectious Diseases Society of America (IDSA) promoting high exposure targets for vancomycin are sparsely supported by safety data (27). Moreover, most toxicity studies on aminoglycosides were performed in the multiple daily dosing era. Therefore, prospective studies investigating the toxicity of currently prescribed once daily or extended-interval dosed aminoglycosides in highly exposed populations e.g. patients suffering from endocarditis are warranted. Newly developed high dosing protocols of vancomycin for long-term MRSA treatment also raise questions about the risk/benefit ratio (28).

Furthermore, there is a need for pharmacokinetic studies investigating vancomycin exposure in patients on continuous infusion in detail during the first 24 hours of therapy (29-31). Therefore, we advocate future studies to address this issue first by incorporating intensive

PK monitoring during the first 24 hours to design proper PK models of continuous infusion of vancomycin and identify optimal sampling times for limited sampling strategies in clinically infected populations. Secondly, determinants for patients at risk of under- or overexposure may be identified. Next, we recommend the implementation of 'early TDM', which has recently also been suggested for intermittent dosing of vancomycin (32). The AUC_{24h} and steady state concentration can be estimated accurately shortly after initiation of vancomycin from two serum samples drawn within 12 hours after initiation of therapy using a validated PK model and PK software. This 'early TDM' approach offers the opportunity to swiftly identify patients that are or will be significantly under- or overexposed and may thereby enable clinicians to improve patient outcomes. Finally, the ongoing debate on the preferential mode of administration of vancomycin should be addressed by large multi-center investigations preferably of prospective randomized design.

Many experts believe that clinical usage of tobramycin and gentamicin will increase due to resistance to our current first line antibiotic armamentarium (33-36). This could result in selection pressure of first choice aminoglycosides on a global scale. Recently, the Swedish Reference Group for Antibiotics (SRGA) has carried out a risk-benefit analysis of aminoglycoside treatment based on clinical efficacy, antibacterial spectrum, and synergistic effect with beta-lactam antibiotics, endotoxin release, toxicity, and side effects. For instance, in cases of suspected infection caused by multidrug-resistant Enterobacteriaceae the SRGA nowadays promotes amikacin instead of gentamicin or tobramycin which is generally more active against extended-spectrum beta-lactamase (ESBL)-producing and quinolone-resistant *Escherichia coli* than other aminoglycosides (37).

Of interest, after the introduction of the SRGA guidelines the increase in Swedish orders of amikacin immediately lead to drug shortages in other EU countries. This is just one examples showing that antimicrobial agent shortages are of global concern and not limited to second or third world countries nowadays (38). It is a multifactorial problem and the current situation of antibiotic shortages is unfortunately more likely to aggravate than to disappear in the years to come.

With regard to the amikacin nephrotoxic and ototoxic potential, there is mechanistic and clinical evidence that the risk of developing nephrotoxicity is relatively low compared to the others within-class drugs (39, 40). Nevertheless, more studies on amikacin with current once daily

dosing regimens are needed. For similar reasons, netelmicin, a previously used aminoglycoside which is thought to have a low nephrotoxic potential (40, 41), will regain interest. It is also worthwhile to mention plazomicin, a promising aminoglycoside, that has been tested in phase 3 studies (42). Plazomicin is designed to overcome tobramycin and gentamicin resistance. Human studies to date have not yet reported nephrotoxicity or ototoxicity, but lack of plazomicin ototoxicity has been reported in the guinea pig model for plazomicin (42). Given the reported increase in bacterial resistance to current antimicrobial agents, plazomicin may be considered a welcome addition to the antibacterial armamentarium pending positive results from large-scale clinical trials and other required clinical studies. In conclusion PK/PD studies of 'new' dosing regimens of 'older' aminoglycoside antibiotics are urgently needed, anticipating broad clinical use of these antimicrobial agents in patient populations with serious infections and a high drug exposure.

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Nederlandse Samenvatting

In dit proefschrift worden strategieën ter optimalisatie van de behandeling met de antibiotica aminoglycosiden en vancomycine onderzocht. De blootstelling van de individuele patiënt aan deze antibiotica was hierbij het uitgangspunt. Door het meten van geneesmiddelconcentraties in het bloed van patiënten met infecties werd voor ieder individu naar de optimale dosering gezocht. Daarbij wordt gestreefd naar een zo gunstig mogelijke balans tussen effectiviteit en veiligheid van aminoglycosiden en vancomycine.

Het inleidende hoofdstuk 1 geeft een globaal overzicht van de werkzaamheid, farmacokinetiek, effectiviteit en veiligheid van aminoglycosiden en vancomycine. In hoofdstuk 1.2 is tevens een review naar de klinische uitkomsten van intermitterende versus continue infusie van vancomycine opgenomen. De belangrijkste conclusies van deze systematische review zijn dat continue infusie minder nefrotoxisch lijkt te zijn en dat er op basis van de beschikbare data geen verschil in effectiviteit tussen beide toedieningsvormen kan worden aangetoond.

In de hoofdstukken 2.1 en 2.2 is bestudeerd of het toedienen van de dagelijkse dosis van een aminoglycoside in de ochtend versus de avond invloed had op de farmacokinetiek van gentamicine en tobramycine. Van meerdere antibiotica is een effect van het circadiaans ritme, dat wil zeggen dag-nachtritme, op de farmacokinetiek van het geneesmiddel beschreven. Doordat meerdaagse regimes van aminoglycosiden voor vrijwel alle indicaties zijn vervangen door minder nefrotoxische eenmaal daagse doseringen, zou het tijdstip van toediening van een aminoglycoside een klinisch relevant effect op de blootstelling kunnen hebben. Wij hebben dit effect bestudeerd in een met tobramycine behandelde populatie van volwassen taaislijmziektepatiënten en bij met tobramycine en gentamicine behandelde patiënten die met een infectie opgenomen werden in een ziekenhuis. In deze onderzoeken werd geen effect gevonden van het tijdstip van intraveneuze toediening van deze aminoglycosiden op de blootstelling. Tevens vonden we geen aanwijzingen voor klinisch relevante verschillen in nefrotoxiciteit. De meest voor de hand liggende verklaring voor de gevonden resultaten is dat een patiënt met een ernstige infectie het 'normale' dag-nachtritme verliest, waardoor het tijdstip van de toediening geen effect op de blootstelling heeft. Bovenstaande studies zijn in nauwe samenwerking met onderzoekers van de Universiteit van Nottingham uitgevoerd.

In een vergelijkbaar onderzoek bij kinderen met taaislijmziekte kwamen zij tot dezelfde conclusies.

In Hoofdstuk 2.3 onderzochten we retrospectief de farmacokinetiek en nefrotoxiciteit van een eenmaal daagse dosering gentamicine en tobramycine in een groot cohort van in het ziekenhuis opgenomen patiënten met (een verdenking op) een infectie. De belangrijkste bevinding van deze studie was dat met tobramycine behandelde patiënten een 50% lagere kans hadden op het ontwikkelen van nefrotoxiciteit in vergelijking met patiënten die gentamicine toegediend hadden gekregen. Dit onderzoek is tot op heden de enige studie die de klinische nefrotoxiciteit van twee of meer eenmaal daags gedoseerde aminoglycosiden onderling heeft vergeleken. Verder toonden we aan dat het oppervlak onder de concentratie-vs-tijd curve (AUC) van de eerste gift de beste voorspeller was van het ontwikkelen van nefrotoxiciteit. Deze resultaten suggereren dat het bijstellen van de dosering en vooral het doseerinterval aminoglycosiden op basis van serum concentraties, afgenomen na de eerste gift, van belang zijn om het risico op nierschade bij individuele patiënten beter in te kunnen schatten.

In hoofdstuk 2.4 hebben we prospectief een nieuw gentamicine doseerregime voor pasgeborenen op een high care unit met (een verdenking op) neonatale sepsis geëvalueerd. Deze studie liet zien dat bij een dosering van 5 mg/kg elke 36 uur de vooraf vastgestelde 'therapeutische' streefwaarden werden gehaald in meer dan 95% van de geïncubeerde patiënten. Een belangrijke toegevoegde waarde van dit onderzoek is de selectie van een populatie pasgeborenen die opgenomen waren op een high care unit. Verreweg de meeste studies op dit terrein zijn uitgevoerd bij pasgeborenen die opgenomen waren op een neonatale intensive care unit (NICU). Zij kennen een veel grotere onderlinge variabiliteit in farmacokinetiek. Daardoor kennen doseerregimes voor een dergelijke populatie pasgeborenen vaak meerdere startdoseringen afhankelijk van het gewicht, de zwangerschapsduur en postnatale leeftijd van de pasgeborene. Wij concluderen dat het onderzochte doseerregime leidt tot een betrouwbare en gelijkmatige gentamicineblootstelling in de onderzochte populatie. Daarnaast suggereren de resultaten dat relatief complexe schema's niet noodzakelijk zijn voor het doelmatig doseren van deze antibiotica bij pasgeborenen die opgenomen zijn op een high care unit.

Tot slot liet deze studie zien dat de AUC van een gift gentamicine bij meer premature, lichtere pasgeborenen gelijk was aan de AUC bij a terme pasgeborenen met een hoger geboortegewicht.

De afwezigheid van een effect van geboortegewicht en zwangerschapsduur op de AUC kon verklaard worden doordat met een toename van gewicht en zwangerschapsduur de halfwaardetijd korter werd terwijl het verdelingsvolume (L/kg) afnam.

Hoofdstuk 3.1 is een systematische review van de bestaande literatuur waarbij de inter-patiënt variabiliteit van vancomycine serum concentraties tussen continue en intermitterende infusie onderling zijn vergeleken. Deze literatuurstudie liet zien dat in populaties die behandeld werden met continue infusie versus intermitterende infusie de inter-patiënt variabiliteit in serum concentratie gemiddeld vijftig procent lager was. Deze interessante bevinding is gebaseerd op 34 gepubliceerde artikelen en gaf aanleiding tot het opzetten van een direct vergelijkend onderzoek tussen intermitterende en continue infusie middels een retrospectieve cohort studie (hoofdstuk 3.2). Ook dit onderzoek liet zien dat de inter-patiënt variabiliteit van vancomycine serumconcentraties ongeveer tweemaal lager was bij de met continue infusie behandelde populatie in vergelijking met de groep patiënten die vancomycine via intermitterende infusie kreeg toegediend. Daarnaast toonde het laatstgenoemde onderzoek aan dat de correlatie tussen plateauconcentraties en de geschatte AUC bij continu doseren sterker was dan de correlatie tussen dalspiegels en geschatte AUC bij intermitterend gedoseerde patiënten. Aan deze resultaten liggen volgens ons twee oorzaken ten grondslag. Om te beginnen heeft de 'timing' van afname van het serummonster in geval van dalspiegels bij intermitterende infusie invloed op de gemeten concentratie. Een effect dat in geval van continu doseren en plateauspiegels vrijwel afwezig is na het bereiken van steady state condities. Verder wordt de inter-patiënt variatie in dalspiegels tijdens intermitterende infusie eveneens beïnvloed door verschillen in distributievolume in tegenstelling tot de variatie in plateauspiegel bij continu doseren. Patiënten met een relatief hoog distributievolume van vancomycine hebben hogere intermitterende doseringen nodig om therapeutische dalspiegels te bereiken, hetgeen zal resulteren in een grote 'delta' tussen piek- en dalspiegel en daarmee in een hoge AUC/dalspiegel ratio.

Wederom uitgaande van steady state condities zullen de plateauspiegels van continu gedoseerde patiënten niet beïnvloed worden door de inter-patiënt verschillen in distributievolume van vancomycine (hoofdstukken 3.1 en 3.2).

In hoofdstuk 3.3 wordt een methode gepresenteerd voor het ontwerpen van een vancomycine doseernomogram voor continue infusie waarbij gebruik gemaakt werd van historische data verkregen binnen de onderzochte populatie.

Vervolgens werd het ontworpen doseernomogram geïmplementeerd in de klinische praktijk en prospectief geëvalueerd. Dit hoofdstuk biedt handvatten ten behoeve van klinici die een switch van intermitterend naar continu doseren van vancomycine overwegen. Ondanks dat het niet onze intentie was om een doseernomogram voor de continue infusie van vancomycine te ontwerpen voor gebruik in andere klinisch geïnfecteerde ziekenhuispopulaties, komt het algoritme wel overeen met het enige andere prospectief gevalideerde doseernomogram opgesteld door Pea en collega's.

In hoofdstuk 4 worden allereerst de samengevatte resultaten van de hoofdstukken 2 en 3 beschreven. Daarna wordt dieper ingegaan op de klinische implicaties. Het hoofdstuk eindigt met enkele suggesties voor toekomstige studies. Met betrekking tot de behandeling met aminoglycosiden is ons belangrijkste advies tobramycine te verkiezen boven gentamicine bij patiënten voor wie een langere behandelduur geïndiceerd is. Volgens onze resultaten, welke gesteund worden door de resultaten van direct vergelijkend onderzoek uitgevoerd ten tijde van het tijdperk van meerdaags doseren, geeft dit een lagere kans op het ontwikkelen van nefrotoxiciteit. Er lijken nauwelijks argumenten te pleiten tegen dit therapieadvies. Er is bewijs dat tobramycine wereldwijd minder microbiële resistentie kent en tevens dieper penetreert in weefsels dan gentamicine. Daarnaast liggen de huidige inkooprijzen van intraveneuze tobramycine preparaten lager dan de prijs van de gentamicine equivalenten. Wij stellen dan ook voor om een prospectief gerandomiseerde studie te ontwerpen waarin klinische uitkomstmaten van eenmaal daags gedoseerd gentamicine en tobramycine worden vergeleken. Met betrekking tot de modus van toedienen van vancomycine blijkt uit ons literatuuronderzoek dat er weinig direct vergelijkend onderzoek is uitgevoerd naar de klinische uitkomstmaten van intermitterende en continue infusie.

Op grond van het momenteel beschikbare bewijs lijkt toediening per continu infuus bij de meeste indicaties niet inferieur aan intermitterende toediening en heeft het mogelijk voordelen bij de behandeling van osteomyelitis en pleuraempyeem. Daarnaast zijn er aanwijzingen dat continue infusie minder nefrotoxisch is dan intermitterende infusie. Tevens is continue infusie van vancomycine bewezen goedkoper en kent het enkele andere praktische voordelen, zoals eenvoudigere monitoring van de blootstelling en de mogelijkheid tot eenvoudiger doorbehandelen in de thuissituatie.

Tot slot zijn er geen grote bezwaren die pleiten tegen het toepassen van continue infusie van vancomycine. Gezien bovenstaande argumenten prefereren wij continue infusie van vancomycine boven intermitterende infusie. Echter, grote prospectieve multicenter gerandomiseerde klinische studies zijn nodig om de verschillen in behandeluitkomsten tussen continue en intermitterende infusie van vancomycine beter te karakteriseren.

Dankwoord

Dit promotieonderzoek is in 2009 begonnen als gevolg van een uit de hand gelopen onderzoeksstage te Den Haag gedurende mijn opleiding tot ziekenhuisapotheker in het Groene Hart Ziekenhuis. Tijdens deze stage ontstond mijn interesse voor klinisch onderzoek. Kort na aanvang van deze “buitenpromotie” aan de Universiteit Maastricht en mijn registratie tot ziekenhuisapotheker startte ik als ziekenhuisapotheker laboratorium te UMC Utrecht. Zo kon het gebeuren dat dit proefschrift gevuld met Goudse en Haagse data te Maastricht verdedigd zal worden door een Utrechtse ziekenhuisapotheker.

Bovenstaande constructie is allesbehalve standaard en voor het slagen ervan ben ik een groot aantal mensen dank verschuldigd. Graag wil ik een ieder bedanken die mij direct of indirect geholpen heeft bij mijn magnum opus. Zonder jullie werk en steun was dit proefschrift niet als dusdanig tot stand gekomen. Een aantal mensen wil ik hieronder in het bijzonder bedanken.

Allereerst mijn ‘promotieteam’. Promotoren Kees Neef en Daan Touw en co-promotor Arthur van Zanten. Daan, eigenlijk ben jij degene die mij motiveerde en inspireerde om mijn registratieonderzoek uit te bouwen naar een promotie. Je enthousiasme werkt aanstekelijk op jonge onderzoekers. Ook je toewijding voor klinisch (farmacologisch) onderzoek en voor je promovendi heb ik enorm gewaardeerd de afgelopen jaren. Daarnaast we zaten altijd snel op een lijn; van nulhypothese tot indiening van het manuscript. Heerlijk! Kees, samen met Daan vormde je mijn aanvankelijke ‘begeleidings- team’. Een promotor op bijna 200 km afstand is een uitdaging, maar gelukkig was je altijd goed benaderbaar en flexibel bij het inplannen van een volgend promotieoverleg. Afspreken in de centrale Domstad aansluitend aan de KKGt vergaderingen leek logistiek de beste oplossing. Het duurde overigens wel even voordat ik de juiste gangen in het doolhof richting NVKC gebouw wist te vinden. Bedankt voor je begeleiding! Arthur, je sloot je pas later aan bij het team. De timing was niettemin perfect. We hebben elkaar leren kennen op een studiereis naar een infusenfabriek in Baskenland. Het was me snel duidelijk dat je een intensivist was met buitengewone interesse in farmacologie van antibiotica en bevlogen innovator van de gezondheidszorg. Naast je onmisbare ‘klinische bril’ leerde ik onder andere van je hoe een artikel weg te zetten en hoe zo elegant mogelijk de discussie met referenten aan te gaan. Dank!

Beste (ziekenhuis)apothekers van het UMC Utrecht, mijn directe collega's, ook jullie steun en interesse heb ik als bijzonder ervaren. Onze open cultuur en bekwame vakgroep leent zich uitermate voor 'even sparren' of desgewenst raad inwinnen bij een ervaren onderzoeker. Met aan zekerheid grenzende waarschijnlijkheid zullen er dit jaar vier apothekers binnen onze groep promoveren. Een onwaarschijnlijke oogst, die niet alleen aan de inspanning van dat fortuinlijke viertal is toe te wijzen maar een verdienste van formaat van de hele vakgroep genoemd mag worden. Getuige het snel groeiende aantal onderzoekers en het gunstige research klimaat zullen er zeker nog vette jaren volgen. Het voert te ver om iedereen van jullie individueel te bedanken, maar tot een aantal van jullie wil graag persoonlijk het woord richten. Toine, werkelijk bijzonder dat je vanaf het moment van mijn indiensttreding mijn externe promotieonderzoek hebt gefaciliteerd. Naast de permissie voor de benodigde tijd stond je me altijd met raad en daad terzijde. Niet alleen op gebied van mijn promotieonderzoek maar eigenlijk ziekenhuisfarmaciebreed. Samen we hebben de afgelopen jaren veel bereikt en werken in jouw apotheek geeft mij nog dagelijks plezier. Ik hoop dit voort te kunnen zetten en daar waar mogelijk uit te bouwen. Karin, ons hoofd Triple O, je passie en je gedrevenheid voor onderzoek werken aanstekelijk. Age quod agis! Barbara, jou wil ik heel graag bedanken voor de fantastische tijd en gezelligheid in het UMCU. Ik heb aan jou veel steun gehad als medepromovendus, net als aan Esther trouwens - gedeelde smart is halve smart. Je bent een geweldige collega en je positieve energie zal gemist worden in 't Utrechtse met je vertrek naar 't schone Brabantse land. Bedankt en veel succes in Tilburg!

Marcel en Tessa, kamergenoten, wat een lief en leed hebben we met elkaar gedeeld de afgelopen jaren. Marcel, mijn partner in PK crime, iedere farmacologische discussie met jou is een genot - wie er uiteindelijk ook gelijk heeft of gelijk neemt! Je bent de collega die niet alleen in den lijve maar ook in farmaceutische gedachten het dichtst bij mij staat. Tessa, jou wil ik ten eerste heel graag bedanken voor mijn huidige prachtige baan. Bij mijn sollicitatiegesprek dacht ik werkelijk in het ootje genomen te worden, toen je zei dat je zelf de QC wilde gaan doen en tegelijk de TDM aan mij over te laten. Bedankt ook dat ik al jouw mooie ideeën voor onderzoeken uit heb mogen voeren. In een vergelijkbaar opgezet onderzoek bij kinderen met taaislijmziekte kwamen zij tot dezelfde conclusies.

– zeker de helft van mijn huidige wetenschappelijke output bevat een vleugje van jouw gedachtengoed. Met uitzondering van onze Kruyt periode delen we in onze kamer al 5 jaar onze ‘highs and lows’. Misschien kunnen we inmiddels een Brabants tegeltje overwegen: ‘Ge mot ‘t nie opkroppen, verrekte m.....’ of houd je het toch liever bij ‘GMP is our lifestyle’? Bedankt ook dat je me hebt geleerd ‘hoe de hazen lopen’ in een academische tent.

Graag wil ik ook alle analisten van het laboratorium van de UMCU apotheek danken. Vincent, Evelien, Sabine, Klaas, Annelies, Mohsin, Mandy, Kamiel, Yvette, Bea, Anneke en Janneke. Ik kan nog dagelijks genieten van het opzetten van innovatieve analyses en ben daarnaast trots op hoe jullie jezelf ontwikkeld hebben. Werken met jullie op ons lab blijft interessant. Bedankt voor jullie steun en interesse. Ook de analisten van de apotheeklaboratoria van het Groene Hart Ziekenhuis - Jan Leen, Ellen en Kees en alle analisten van de Apotheek Haagse Ziekenhuizen wil ik bedanken voor hun hulp bij het uitvoeren van de analyses en het verzamelen van de data gebruikt voor dit promotieonderzoek.

Verder wil ik mijn voormalige Goudse collega's bedanken. Aan mijn opleidingstijd in Gouda bewaar ik warme herinneringen en nog steeds goede contacten. Tot een aantal mensen wil ik een persoonlijk woord van dank richten. Allereerst Monique, ik ben je zeer erkentelijk voor de kans om mijn registratieonderzoek in Den Haag onder wetenschappelijke begeleiding van Daan uit te voeren. Natuurlijk hield je wel de planning strak in de gaten. Daarnaast heb ik enorm veel van je geleerd over het ziekenhuisapothekerschap. Deze laatste woorden gelden tevens voor Mignon, opleider te Gouda tijdens het staartje van mijn registratiefase. Sardha, Annemien, Charlotte, Hans, Katja, jullie hebben mij mede gevormd tot de ziekenhuisapotheker en onderzoeker die ik nu ben. Ik ben jullie dankbaar voor het overdragen van jullie ervaring en kennis en ook voor alle gezelligheid natuurlijk. Tot slot wil ik graag Franny en Anke bedanken, want jullie zijn allebei heel erg belangrijk voor mij geweest op zowel professionele als persoonlijke vlak.

Nog een woord van dank voor alle studentstagiairs die dit onderzoeksproject versleten heeft. Twee van jullie zijn voor mij van onschatbare waarde geweest. Walaa, wat heb jij hard gewerkt tijdens je stage. Alles begint met inzet en toewijding. Aan beide was bij jou geen gebrek. En

Suzan, als er zo iets als een ideale stagiair bestaat dan kom jij daar aardig bij in de buurt. Veel succes met afstuderen en in je aanstaande carrière. Bedankt ook alle verpleegkundigen en arts-assistenten - in het bijzonder Renske - op de afdeling longgeneeskunde van het Haga ziekenhuis voor de inclusie van de CF patiënten voor de CIRCA studie. Dank ook aan alle co-auteurs voor jullie hulp bij het opstellen van METC aanvragen, bij schrijven van de manuscripten en voor het delen van jullie netwerken.

Natuurlijk wil ik mijn familie graag bedanken - om te beginnen Els, mijn moeder en Gerard, mijn vader en John. Jullie hebben me altijd aangemoedigd - maar nooit gepusht - om mijn talenten te ontwikkelen. Dit boekje is ook jullie verdienste. Helaas zal mijn vader mijn 'hora est' niet mee kunnen maken, maar ik weet zeker dat hij trots op me zou zijn geweest. Ook een vermelding waardig zijn de grootste oppasactiviteiten van mijn moeder en John. Sam, Liv en Ties hebben bij jullie hun tweede thuis! Mijn bro's Jelle en Thomas, jullie zorgden vroeger voor de nodige 'serious competition' - ik herinner me onder andere rondvliegende tafeltennisbatjes. Juist voor die potjes tafeltennis wil ik jullie bedanken. De 'innerlijke drive', die nodig is om een proefschrift af te ronden, vindt vermoedelijk hierin zijn oorsprong. Ik ben gelukkig met twee fantastische broers en hoop dat we nog veel lol met elkaar kunnen hebben. Een bedankje aan mijn vrienden is op zijn plaats en in het bijzonder Bram en Erik, mijn vrienden voor het leven, mijn mede-brave huisvaders. Na ruim 16 jaar vriendschap kennen jullie mij van binnen - tot op orgaaniveau - en van buiten. Door promoties, werk en vooral het toegeven aan onze voortplantingsdrang - zonder oordeel een drang bij de een iets groter dan bij de ander - zien we elkaar wat minder in deze tropenjaren. Echter, onze kroost zal eens het nest verlaten en oude tijden zullen herleven! Dank voor jullie steun. Erik, bedankt ook dat je me bij zal staan als paranimf. Ferdi, mijn tweede paranimf, in Gouda waren we ooit 'concullega's', een gezonde competitie die louter winnaars en een vriendschap heeft opgeleverd. Ik vond het een eer dat ik tijdens jouw verdediging je paranimf mocht zijn en dat jij nu mijn paranimf bent.

Tot slot wil ik mijn Marieke bedanken en uiteraard niet alleen voor de opmaak van het binnenwerk en het ontwerp van de omslag. We zijn al tien jaar samen en hebben in die tijd al veel meegemaakt. Je bent mijn

grote liefde en mijn beste maatje. Dat dit boekje het levenslicht heeft mogen aanschouwen, is zonder enige twijfel ook de verdienste van mijn veelzijdige en multitaskende verloofde. De geboorte van dit boekje, mijn 'vierde kindje', is tot stand gekomen met geregeld hevig gepuf en gesteun van mijn kant. Gelukkig was jij er altijd om me te steunen, net als ik er omgekeerd voor jou was tijdens de geboortes van onze kinderen - althans in mijn herinneringen is dat zo. Lieve Marieke, we gaan samen verder met het verwezenlijken van onze dromen. Ik ben trots op je en hou van je.

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Valorization paragraph

As the current drug development pipeline wanes new classes of antibiotics, it is essential we learn to exploit our current antibiotic arsenal optimally by 'rediscovering' 'older' antimicrobial agents. Aminoglycosides, e.g. gentamicin and tobramycin and glycopeptides, e.g. vancomycin are considered antibiotics of older classes as they were marketed in the 1950s. Aminoglycosides and vancomycin are cornerstones of the current antibiotic armamentarium because of their activity against multidrug-resistant pathogens that can cause high rates of mortality and morbidity especially in hospitals. However, tobramycin and gentamicin have narrow therapeutic ranges and can induce both serious damage to the kidneys and ears limiting its clinical use. Clinicians prescribing vancomycin and other glycopeptides, on the other hand, are faced with emerging resistance among methicillin-resistant *Staphylococcus aureus* (MRSA) and vancomycin-intermediate *S. aureus* (VISA) strains worldwide. Although vancomycin still is the antibiotic of choice for these types of infections, the development of drug resistance is increasing. Furthermore, there is a risk of treatment failure if minimal drug exposure targets are not reached and maintained in individual patients. Vancomycin resistance is a serious threat that needs to be dealt with in the years to come if we want to preserve this vital antibiotic for future generations. Therefore, we explored pharmacologically-based strategies to optimize aminoglycoside and vancomycin therapies.

In this thesis data are presented on gentamicin, tobramycin and vancomycin pharmacokinetics and clinical outcomes in clinically infected populations. The patients populations investigated include cystic fibrosis patients, neonates, general ward and critically ill patients. Patient populations that show aberrant aminoglycoside and vancomycin pharmacokinetics and that are potentially susceptible to toxicity and treatment failure. Moreover, therapeutic drug monitoring (TDM) is recommended to optimize aminoglycoside and vancomycin therapy in individual patients.

In chapters 2.1 and 2.2 we showed that the time period of administration did not affect the clearance or exposure in terms of area-under-the-curve (AUC) in CF patients, general ward patients and ICU patients. Hence, in daily clinical practice 24-hour intervals should be maintained unless therapeutic drug monitoring results indicate that the dosing interval needs to be extended. Shifting administration times to the morning period - which is not uncommon in current practice - is not

likely to offer a benefit to these patients. It can, however cause side effects or treatment failure due to shortening or extending administration intervals, respectively. Although our findings deserve to be confirmed, results from Chapter 2.3 showed a higher nephrotoxic potential of once daily dosed (ODD) gentamicin over ODD tobramycin in clinically infected patients treated for at least 3 days which may lead to the revision of (inter)national and local treatment protocols. In patients who are at high risk of developing nephrotoxicity and/or are likely to be treated over a longer period, tobramycin should be preferred over gentamicin. In fact, European cystic fibrosis treatment guidelines already advocate choosing tobramycin over gentamicin for this reason. The same concerns with regard to cumulative toxicity may apply to other highly exposed populations, e.g. patients treated for endocarditis. In order to optimize treatment with aminoglycosides 'early' AUC monitoring in addition to 'trough levels' should be implemented in daily routine, since our data suggest the AUC of the first administration is a strong predictor of nephrotoxicity.

The added value to literature of our study results presented in Chapter 2.4 on gentamicin dosing in neonates lies in the specific neonatal population investigated. Albeit the results deserve to be prospectively confirmed in a second independent cohort of neonates admitted to a level II special care nursery, our findings show that a more uniform dosing protocol of 5 mg/kg per 36 hours can be implemented in level 2 unit compared to more complex dosing protocols proposed in neonates admitted to a level III intensive care unit. This offers the potential use of an easy one-dose-fits-all protocol, which is likely to reduce prescribing errors in neonates. The robustness of the investigated dosing regimen may also reduce the need for TDM and thereby the number of gentamicin blood samples. Reduction of blood sampling in neonates lowers the risk of infections and is one of focus points of the World Health Organization (WHO).

As described in Chapter 3.1 vancomycin continuous dosing protocols are rapidly gaining interest and are being implemented in clinical practice. Continuous infusion of vancomycin might also be preferred since the drug preparation time, the staff workload and costs are reduced and may also shorten hospital length of stay. Apart from the practical advantages over intermittent infusion, continuous infusion of vancomycin results a more predictable exposure and thus has better target attainment odds as shown in Chapter 3.1 and 3.2 are better.

Finally, based on our methodology described in chapter 3.2 a 'tailored' dosing algorithm of vancomycin can be designed for clinically infected populations aiming for the desired target exposure.

The results reported in this study are of considerable interest to clinical pharmacologists and clinicians especially infectious disease specialists, clinical microbiologists, pediatricians and intensive care physicians.

Furthermore, our data can contribute to future antibiotic (inter)national guidelines. Moreover, the cost savings achieved by higher cure rates, lower percentages of patients experiencing treatment related side effects and higher efficiency are of great interest to policymakers. The increase in drug resistance urges both the private and public sector to keep these older antibiotics available. In our opinion consortia should be formed to investigate potential applications and new indications of these antibiotics by designing innovative drug formulations and performing clinical studies. For instance, the European Union is currently running a program promoting private and public collaboration to test new formulations of inhaled antibiotics in patients with cystic fibrosis. Finally, the suggestions for future studies made in chapter 4 are of interest to other researches in the field of antibiotics. Studies call for more studies. To conclude, in an era of increasing antibiotic resistance against our current first line of defense, this thesis provides pharmacology-based strategies striving for a safer and more effective clinical usage of tobramycin, gentamicin and vancomycin.

Curriculum Vitae

Erik van Maarseveen was born on the 17th of May 1979 in Tiel.

He graduated at the Gymnasium of the Tiel College in 1998.

He studied Pharmaceutical Sciences from 1998 till 2004 at the University of Utrecht and received his Pharmacist Degree in 2005. That year he also completed all theoretical courses of the Master program “Business and Economics for Beta’s” at the University of Utrecht.

In 2005 he was employed as a pharmacist to work on a project promoting an early switch of intravenous to oral antibiotics at the Groene Hart Ziekenhuis Gouda. In 2006 he started his training as a hospital pharmacist at the Groene Hart Ziekenhuis. He has been working as a registered hospital pharmacist since 2010 at the University Medical Center Utrecht, where he is the lead pharmacist of the therapeutic drug monitoring and toxicology laboratory. In 2015 he will start his training as a clinical pharmacologist at the University Medical Center Utrecht.

In 2009 he started his PhD on the pharmacology of aminoglycosides and vancomycin at Maastricht University, which resulted in this thesis. Next to antibiotics his current research focusses on the pharmacology of immunosuppressive, antiretroviral and anti-cancer drugs.

Erik van Maarseveen is engaged to Marieke Velthuis. They are proud to be the parents of three beautiful children; one daughter Liv (2010) and two sons Sam (2008) and Ties (2012).