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A dual-respiration chamber system with automated calibration

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Schoffelen, Paul F. M., Klaas R. Westerterp, Wim H. M. Saris, and Foppe Ten Hoor. A dual-respiration chamber system with automated calibration. *J. Appl. Physiol.* 83(6): 2064–2072, 1997.—This study characterizes respiration chambers with fully automated calibration. The system consists of two 14-m³ pull-type chambers. Care was taken to provide a friendly environment for the subjects, with the possibility of social contact during the experiment. Gas analysis was automated to correct for analyzer drift and barometric pressure variations and to provide ease of use. Methods used for checking the system's performance are described. The gas-analysis repeatability was within 0.002%. Results of alcohol combustion (50–350 ml/min CO₂) show an accuracy of 0.5 ± 2.0 (SD) % for O₂ consumption and -0.3 ± 1.6 % for CO₂ production for 2- to 24-h experiments. It is concluded that response time is not the main factor with respect to the smallest practical measurement interval (duration); volume, mixing, gas-analysis accuracy, and levels of O₂ consumption and CO₂ production are at least equally important. The smallest practical interval was 15–25 min, as also found with most chamber systems described in the literature. We chose to standardize 0.5 h as the minimum measurement interval.

indirect calorimetry; energy expenditure; oxygen consumption and carbon dioxide production; measurement interval

ENERGY EXPENDITURE IN HUMANS can be determined by direct measurement of heat loss (direct calorimetry) or by calculation of heat production from O₂ consumption (V_{O₂}; ml/min), CO₂ production (V_{CO₂}; ml/min), and nitrogen loss in urine. V_{O₂} and V_{CO₂} may be determined with a variety of methods usually involving a mouthpiece, face mask, or ventilated hood (2, 9, 20), limiting the duration of the measurement to a few hours. For the determination of V_{O₂} and V_{CO₂} during a longer time interval (up to several days), a respiration chamber may be the method of choice (1, 3–6, 8, 11–19). During the measurement, the subject stays in an airtight room through which a stream of fresh air is directed. Composition and volume of the inlet and outlet airstream are measured. The respiration chambers described below feature a double set of gas analyzers with continuous automated calibration and automated data collection. This approach circumvents most problems due to ambient variations in gas composition and pressure and due to operator errors. Independent checks of the automated calibration procedure are performed regularly by using alcohol combustion or injection of gas with known composition. The chambers have been operational for over 10 years and provide an

easy-to-use and labor-saving service with minimum downtime.

METHODS

Subject Environment

Two equally sized (14 m³) and furnished chambers were placed next to each other (Fig. 1). They give the impression of a normal room, with windows positioned in the door for contact with researchers, in the wall (outside view), and between the chambers for visual contact between subjects. Curtains ensure privacy when needed. Each chamber has a full-sized foldaway bed, a bureau with built-in sink, a folding chair, a color television set, an audiocassette player, an alarm clock, a telephone, an automated intercom, and a computer-network connection. Confined spaces were avoided because of the adverse effect on the air-mixing process (APPENDIX A). Both chambers are occupied simultaneously, preventing subjects from feeling isolated and stimulating normal domestic behavior.

The chambers are equipped with a deep-freeze toilet (Special Product, Mulders) for collecting feces; urine is collected separately in bottles. Three air locks provide passage for the exchange of food, collection of feces and urine, and for sampling of blood. Safety precautions include a fire alarm and extinguisher, emergency lighting, and panic buttons. The door can be opened from both sides without hindrance. The chambers are checked once a year for electrical safety (S1 standard), and the climate is constantly regulated and monitored by an automated information system. Physical activity can be performed by using a cycle ergometer (Lode) or a treadmill (Quinton). The height of the chamber also allows the use of a stepping platform. Activity of the subject is measured by an analog ultrasound system (Advisor DU160).

Ventilation

For airtightness, each room was constructed of six prefabricated welded steel parts, bolted together with a sealing mass in between. Door and air locks have a flexible seal. The air in the surrounding laboratory is ventilated with fresh air at a rate of five times its volume per hour (1,200 m³/h). Fresh air from outside the building is drawn through the chambers by a ventilator with a capacity of 30–250 l/min at a negative pressure of 600–50 Pa, respectively (pull type). The flow is adjusted with a valve at the output. Negative pressure is adjusted to 250 ± 125 Pa with a valve at the input and is measured by using a U-type oil gauge against barometric pressure. Flow is measured with dry bellows meters (G4, Meterfabriek Schlumberger) calibrated by mass-flow (CO₂ weight) from a gas bottle to 0.2%. [This calibration method was periodically verified by sending a calibrated gas meter to the national standards laboratory (Nederlands Meet Instituut)] The flowmeters are equipped with a digital pulse output for continuous flow measurement. The air-conditioning has a capacity of 3,700 W, which is mainly needed for dehumidification of the air during exercise. The air in the chambers is mixed with a radial ventilator, forcing the air

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Fig. 1. Layout of dual respiration chamber. 1, Television set; 2, chair; 3, sink with hot and cold running water; 4, deep-freeze toilet; 5, air lock for blood samples; 6, cycle ergometer; 7, bed (folded down for sleeping or sitting); 8, body-weight balance; 9, air locks, one for food, one for feces; 10, folding chair, which may be stored under bed; 11, bed (folded up for more floor space).

through a draft-reducing perforated ceiling at a controllable rate of 3,300–10,000 l/min.

The fresh-air supply is routed directly into the air-conditioning for mixing and temperature control. The temperature variation is $\pm 0.1^\circ\text{C}$ during rest and $\pm 0.4^\circ\text{C}$ during exercise. The air leaves the chamber diagonally opposite the input at two levels. Volume and flow measurements are corrected to STPD by using data obtained from temperature (AD590, National Semiconductor), humidity (SA100c, Rotronic), and barometric pressure (4-801-1124, Bell & Howell) sensors that are calibrated on a yearly basis.

O₂- and CO₂-Measurement Systems

O₂ is measured by using paramagnetic 0–22% oxygen analyzers (Magnos 6G, Hartmann & Braun; OA184A, Servomex), and CO₂ is measured by using infrared 0–1% analyzers (Uras 3G, Hartmann & Braun). To improve the reliability of the measurement, each gas sample is analyzed in duplicate, reducing the risk of losing data because of hard-to-detect failures.

Samples from the input and the output of the chamber are drawn into a sample preparation unit by using membrane pumps (model 300, Wisa). When a sample is not selected, the sample line is still flushed to reduce dead time. Pressure and humidity variations are reduced through utilization of needle valves and oil-filled overflow bubblers (constant pressure to ambient) and by using membrane dryers (ME050-24-MFL, Perma Pure). The membrane dryers have an enhanced drying capacity obtained from using an outer hull with a counterflowing dry purge gas at 50 KPa negative pressure; this provides a steady drying capacity. The combination of fully flushed sample routes with identical delays and a membrane-drying tube resulted in a 90% response of the CO₂ analyzers of 5 s after the switch from N₂ to calibration gas. The system ensures that all samples are clean (1- μm filter) and are of equal pressure (± 10 Pa), temperature ($\pm 0.1^\circ\text{C}$), and humidity (-15°C dew point). The linearity error of each CO₂

analyzer was reduced by constructing a linearization curve for each apparatus. The range of the linearized curve is 0–0.8%; the CO₂ concentration inside the chambers normally never exceeds 0.8%.

The difference in gas composition (dg) between incoming and outgoing air and the ratio of dg to the difference in time (dt) in the chamber (derived from measurement of the outgoing air) have to be known for the calculation of $\dot{V}\text{O}_2$ and $\dot{V}\text{CO}_2$ (APPENDIX B). Air samples are measured in sequence (10) and alternated with samples of calibration and zero gases, thus eliminating errors because of differences in analyzers or sample preparation. The measurement of each sample requires 1 min. During each interval of 15 min, samples of fresh air and zero and calibration gas are measured in addition to the 12 chamber samples (Fig. 2). In this way, effects due to baseline drift, barometric pressure (3, 18), and temperature variation, factors that vary more slowly than in 15 min, are minimized. Because of the full automatization, no operator action is required, eliminating this source of error.

Fig. 2. Sample sequence. Inlet and outlet samples from chambers 1 and 2 are alternated with N₂ (zero control), fresh air, and calibration gas, thus obtaining zero control and calibration every 15 min.

Calibration and fresh-air measurements account for 12 min every hour. The remaining 48 min during the hour provide time for two concentration measurements for both chambers during each of twelve 5-min intervals. $\dot{V}O_2$ and $\dot{V}CO_2$ are calculated for each 5-min interval, and the 5-min results are integrated to 0.5-h values in the standard output file. Although it is possible to calculate 5-min values for $\dot{V}O_2$ and $\dot{V}CO_2$, the accuracy of these values will be low because the standard deviation (SD) in the measurement of the minute concentration changes is multiplied with the large volume of the chamber (13). The standard procedure is the calculation of $\dot{V}O_2$ and $\dot{V}CO_2$ over 0.5 h or longer time intervals.

The calibration gas contains 0.8% CO_2 -18% O_2 -remainder N_2 . The CO_2 concentration of this gas can be obtained with a certified accuracy of 0.008%. The O_2 concentration, however, has a certified accuracy of only 0.18%. For the O_2 analysis, we therefore rely on the accuracy of the overall O_2 concentration of the fresh air (4, 10, 14, 15) during a whole day, while using the 18% O_2 content of the calibration gas as a stable, but at first unknown, O_2 reference. The unknown O_2 concentration of the calibration gas is calculated on the basis of measurement of fresh air, N_2 , and calibration gas O_2 concentration by using mean values over the whole experiment. The calculated calibration gas O_2 value is then used to determine momentary O_2 concentrations during the experiment. Because the O_2 concentration of the calibration gas is calculated during each experiment, monitoring the obtained calibration gas values over a 3-mo interval (lifespan of a single calibration gas bottle) provides data on the accuracy of the O_2 measurement, including any drift in fresh-air O_2 concentration over the 3-mo interval.

A microcomputer (Macintosh, Apple) is used to monitor the parameters for determining $\dot{V}O_2$ and $\dot{V}CO_2$. Each analyzer and sensor has its own analog-to-digital converter (ADC; voltage-to-frequency, VFC62, Burr-Brown) and is optically isolated from the microcomputer, enabling optimal conversion of electrical signals by reducing electrical noise from long cabling and earth loops. Calibration of sensors is done in the software; the analog range of the converters was individually chosen to handle any long-term drift. Parameters used in the calculation of $\dot{V}O_2$ and $\dot{V}CO_2$ are temperature, humidity, flow, barometric pressure, and a digital reading of O_2 and CO_2 concentrations in sample and calibration gases.

On-line calculation enables continuous monitoring of the progress of the experiment. Final calculation is done after the experiment is completed, allowing the use of all data for calculation of calibration constants (12), specifically the O_2 concentration in the calibration gas bottle. The equations used in the calculation of gas exchange are based on the assumption of N_2 conservation ["haldane" correction (3, 5, 7, 10-12, 15-17)] with incorporation of differentiated changes in the chamber volume for the N_2 equation [dFN_2/dt ; determined at the outlet (3, 7, 12, 17)]. Water vapor is taken into account (3, 10) by first calculating all flow and volumes [including differentiated changes in the chamber volume (dH_2O/dt) to STPD. Energy expenditure is calculated from $\dot{V}O_2$ and $\dot{V}CO_2$ with the Weir formula (21).

To achieve flexibility, the software for the system is modular; data acquisition is based on a graphical engineering program (Labview, National Instruments), and calculation is performed with a spreadsheet macro program (Excel, Microsoft). Additional parameters can be incorporated by using the flexibility of the software and the network capability of the computer (network-connected ADCs and serial ports). The audio capability of the computer (speech) was used to synchronize the subject's behavior to a protocol by providing an audio signal to the subject when it was time for a certain activity.

Validation

Each month, an independent check of the whole system is obtained by combusting alcohol inside the chamber or, in some instances, by injecting gas with a known composition into the chamber. The alcohol (99.8% methanol pro analyse; Merck) is combusted by using a gas burner (Fig. 3A). The burner is placed on a calibrated balance connected to a computer to measure the rate of combustion during the experiment. When alcohol is combusted, O_2 is consumed and CO_2 is produced, mimicking normal measurement. With the use of gas injection with CO_2 , N_2 , or a combination of both (Fig. 3B; Refs. 4, 11-14, 17, 18), the accuracy of CO_2 and O_2 measurement can be checked.

Calculating produced CO_2 from weight should take into account that at barometric pressure the CO_2 compressibility (3a) accounts for a -0.6% deviation between molar and volume fraction, valid for both alcohol combustion and CO_2 injection. In this context, it should be pointed out that calibration gas certificates can therefore be obtained on the basis of molar or volume fraction. In our setting, all calculations were done by using volume (fractions) at STPD.

The duration of validation experiments is 24 or 2 h; both time intervals are relevant to actual experiments.

RESULTS

During operation of the system for 10 yr, only 2 of >2,000 subjects felt isolated and finished the experiment prematurely. No safety hazards have occurred.

Ventilation

The perforated ceiling reduced the noise at ear level to 45 dbA at the lowest recirculation flow of 3,300 l/min. At the highest recirculation of 10,000 l/min, the noise increased to 54 dbA. At the lowest recirculation flow of 3,300 l/min and a flow through the chamber of 50 l/min,

Fig. 3. Chambers are checked by combustion of alcohol (A) or infusion of CO_2 (B). Amount of combusted alcohol or injected gas is calculated from weight change of containers; connecting balance to a computer allows continuous measurement of rate of combustion or infusion. Computer can change this rate during experiment if computer-controlled valves (B) are used.

99% of a 5-min continuous injection was measured within 15 min, and 63% was measured in 5 min.

O₂- and CO₂-Measurement System

Key elements in the gas-analysis system are the sampling system and the reproducibility of the gas analyzers. The time needed to flush the sampling system after a change of sample was measured to be ≤ 5 s, leaving 55 s for stabilization [at least 10-fold; analyzer response time [reponse time to 90% (t_{90}) ≤ 3.5 s]] and measurement. The drying capacity of the sampling system, particularly important for measuring O₂, showed a steady sample dew point lower than -15°C . To check the reproducibility of the O₂ analyzers, the difference between two analyzers was measured over the 0–18% O₂ concentration range (Fig. 4) when identical gas samples were measured. The signal-to-noise levels had an SD $< 0.002\%$ ($n = 75$), illustrating the reproducibility of the O₂ gas analysis.

Validation

Alcohol-combustion experiments over the present year ($n = 44$) resulted in differences between “alcohol combustion” values and “chamber system” values of $-0.3 \pm 1.6\%$ for CO₂, $0.5 \pm 2.0\%$ for O₂ (Fig. 5) and a respiratory quotient of 0.663 ± 0.012 . No difference was found between 2- and 24-h tests. The results of calibration experiments using CO₂ injection over the past years were comparable with those found for alcohol combustion ($-0.6 \pm 2.3\%$, $n = 20$).

The system compares the stable O₂ concentration of the calibration gas with the mean 24-h fresh-air O₂ concentration; the variation in these 24-h O₂ concentration measurements can be determined. Maximum difference in 24-h measurements over the 3-mo lifespan of a calibration gas bottle was found to vary from 0.003 ± 0.002 to $0.006 \pm 0.004\%$ O₂. This variation can be attributed to both variation in fresh air %O₂ and variation in the measurement system; these factors

Fig. 4. Difference between 2 analyzer cells (y-axis, %) when identical samples are measured as a function of O₂ concentration in measured gas (x-axis, %).

Fig. 5. Results of alcohol-combustion experiments. Difference between amount calculated from alcohol combustion and amount measured by system (y-axis, %) as a function of CO₂ production by alcohol combustion (x-axis, ml/min) is shown.

cannot be separated. Because these variations are also both present in normal experiments, the maximum difference found is an indication of the performance and stability of the system with respect to calibration based on fresh air %O₂. If calculated O₂ concentration in the calibration gas was compared by using data of two O₂ analyzers, the maximum difference and SD increased by 50%, showing the advantage of measuring input and output concentrations sequentially (10, 14) with one analyzer compared with measurement of input and output concentrations with separate analyzers.

Registration of energy expenditure and physical activity of a subject is shown in Fig. 6. Energy expenditure data are given for 0.5-h intervals. Physical activity was synchronized with these 0.5-h intervals by using computer audio.

DISCUSSION

Subject Environment

After the creation of a friendly environment, only noise and draft due to air conditioning remain as the major factors compromising comfort. The rate of recirculation flow is therefore limited (12) and, because of its cooling effect, the room temperature is normally set a few degrees higher ($\leq 3^{\circ}\text{C}$) than at home.

Ventilation

The recirculation flow range of ventilating the 14-m³ volume at a rate of 15–42 times per hour calculates to a mixing time constant (12) of 1.4–4.2 min, which is comparable to those calculated from the literature (4–6, 12, 14–16), ranging from 0.4 to 4 min. The result

Fig. 6. Energy expenditure (EE) and physical activity of a subject (male, aged 32 yr, body weight 68 kg) as measured over 0.5-h intervals.

of observing 99% of a 5-min continuous injection within 15 min is slightly better than expected from the calculated 4.2-min time constant at the lowest recirculation rate. Extending the recirculation flow range will either compromise the mixing time constant (≥ 4 min) or the comfort of the subject. The negative pressure [pull type (5, 8, 13, 15)] ensures that airflow through leaks will only be from outside to inside the chamber. If the room around the chamber is well ventilated, this will have a negligible effect on the measurements. In some settings, factors like environment (4) or control of inlet air (11, 12) can necessitate the use of a positive pressure system. Such a system [push type (4, 6, 11, 12, 14, 16–18)] requires better sealing (12, 16) because it cannot be guaranteed that leaked air was already completely mixed and sampled.

Wherever possible, care was taken to avoid confined spaces, which would act as buffer volumes. A confined space behaves as a volume in which gas concentrations will slowly follow the concentration in the chamber. If a subject's expired air is directed to a confined space, the mixing time will increase. For this reason, no closed cabinets were provided inside the chambers and the cabinets around equipment like the deep-freeze toilets and television sets were perforated.

O₂- and CO₂-Measurement System

The automated system operates continuously, and thus the system is calibrated 96 times/24 h. The SD < 0.004% in the daily calculated O₂ concentration of the calibration gas over 3 mo shows the capability of the system for handling environmental variation and drift. As far as we know from the literature, this frequent automated calibration is a unique feature of the system, making it easy to use; to start an experiment, one has only to close the door of the chamber. Multiplexing samples in time on one analyzer (10, 14), rather than using multiple analyzers, combined with interleaved (frequent) calibration, eliminates the need for temperature and pressure correction (3, 18) when momentary concentration values are calculated, because the time of calibration is almost identical to the measurement time (≤ 15 min).

Validation

The results of the alcohol-combustion tests were $-0.3 \pm 1.6\%$ for CO₂. One factor determining the accuracy is the CO₂ concentration in the calibration gas, which in our case was determined to be $0.8 \pm 0.008\%$. Elimination of this possible source of deviation

would require a certificate with an accuracy of $0.8 \pm 0.0008\%$ for the CO_2 component and possibly further improved linearization of the analyzers. However, neither was available.

The $0.5 \pm 2.0\%$ result for O_2 is only achievable with analyzers that perform well within the factory specification and requires meticulous sample preparation and stable laboratory conditions. The reason for this is the $\sim 20\%$ O_2 background in all measured air because every type of analyzer has to deal with a 20% background in relation to a 1% measurement span, requiring a 21% physical measurement range for a 1% differential physiological range. This is easily understood for mass-spectrometer (17) and paramagnetic optical analyzers that measure one gas stream; they can only be differential in time (10, 14). However, it is also valid for dual-gas stream differential analyzers because these use dual compartments (magnetic wind principle) for comparison, and in each of these compartments the 20% background again largely determines the signal-to-noise ratio. Because the result of subtracting the background in differential analyzers is instantaneous, it is often erroneously assumed that the 20% background is eliminated from the physical measurement.

Alcohol combustion is normally used for checking experiments because it tests VO_2 and VCO_2 simultaneously and experiments are easy to perform. However, for troubleshooting, CO_2 and N_2 infusion is the method of choice because it is independent of a chemical reaction. The slightly larger error margin in the CO_2 -infusion experiments (over several years) compared with the regular alcohol-combustion experiments (over 1 yr) is attributable to the fact that gas infusion was mostly used for troubleshooting when a problem was detected.

Response Time and Measurement Interval

In the literature, two types of system responses are given. One is actually the time constant (therefore, not

referred to as response time in this study) of the mixing process (8, 12, 15), and the other is the (90–99%) response time of the complete system to a change in energy expenditure (4, 6, 11, 12, 14). The response time incorporates the mixing time constant (because all air should first be well mixed) and will therefore be larger than the mixing time constant. The response time is only important, in part, when measurement protocol is decided on, i.e., rate of change of energy expenditure to be measured. The volume of the chamber, the rate of gas flow through the chamber, and the accuracy of the gas measurement determine the interval (duration) needed to reach 95% of accuracy. Normally, this interval is at least 1 h, as can be seen from calibration experiments (Table 1, Refs. 4, 6, 8, 11, 12, 15–17, 19), even if the response time is much smaller (4, 6, 11, 12, 14, 15, 17). Furthermore, it is not proven that subjects will behave in the same way as the testing methods, specifically with respect to the mixing and leaking (push-type chamber) of the subject's expired air, because the subject can direct his breath and move in any direction inside the chamber in an unpredictable manner. This may be the reason that virtually all publications refer to a minimum measurement interval (duration) of 15–30 min, as illustrated in Table 1.

Today's line of research is often a combination of long-term observations with short-term changes in energy expenditure. If, for example, the O_2 concentration SD of 0.002%, which is the SD we found for our calibration gas O_2 concentration check over 3 mo, is applied to a subject with a VO_2 of 350 ml/min for a 0.5-h interval, the resulting SD in VO_2 will be $\sim 3\%$. In reality, the shorter intervals have a slightly lower SD because the short-term stability of the O_2 measurement is better than 0.002%. In addition, the change in concentration of the chamber volume is smaller, which decreases eventual errors due to nonlinearity of CO_2 analyzers.

Table 1. Overview of achieved accuracy of chambers in the literature in relation to test duration

ΔO_2 , %	ΔCO_2 , %	<i>n</i>	Tested Duration, h	Suggested Minimum Duration, min	Size, m^3	Reference No.	Checking Method
-1 ± 2.2 (CV)*	0 ± 2.1 (CV)*	5	3–24	60	27	4	Butane, N_2 + CO_2 inf
0.06 ± 1.21	-0.49 ± 1.12	5	4–6	20	30	6	Butane
$+0.44 \pm 0.34$	-0.36 ± 0.5	14	1	15–30	31	8	Butane
-0.22 ± 1.51	0.21 ± 0.68	14	24	30	34	12	N_2 , CO_2 inf
-0.53 ± 0.66	-0.05 ± 1.36	9	24		19	12	N_2 , CO_2 inf
2.3–5.8	1.9–4.9	2	0.5			12	N_2 , CO_2 inf
± 1.2	± 0.5				30	13	N_2 , CO_2 inf
$-2.8 \pm 2.8^*$	$-2.1 \pm 2.8^*$	18	3–6		19	15	Propane
1.5 ± 1.4 (CV)	1.5 ± 1.4 (CV)	9	18–22		20	16	Alcohol
0.3 ± 1.54	0.4 ± 1.34	30	>2–24	30	20	17	Alcohol
	0.4 ± 0.85	10	>2–24		20	17	CO_2 inf
± 0.43					15	18	N_2 inf
-0.5 ± 2.1	0.2 ± 2.3	3	24		16	19	Propane
0.5 ± 2.0	-0.3 ± 1.6	44	2–24	30	14	Present study	Alcohol
	-0.6 ± 2.3	20	2–24		14	Present study	CO_2 inf
Summary	$+0.13 \pm 1.26$						
	$+0.09 \pm 0.95$						

Summary mean $\pm$ SD is average deviation from 0. Δ , Change; *n*, No. of experiments; CV, coefficient of variation; inf, infusion. *Values calculated from literature.

Although the results of “ideal” injection experiments (good mixing, predictable injection flow) show that shorter interval measurements are feasible, we chose the standard of 0.5-h results as the smallest practical time interval with subjects. The smallest experiment duration allowed (limited in software) is 2 h, to provide at least eight automatic calibrations for the off-line calculation, although a minimum of 12 h is preferred (for instance, one night). The 2-h interval is also used in our standard checking experiments because it illustrates the accuracy for measuring sleeping metabolic rate, which is done over a 2- to 3-h interval (mostly over 3 h). Whenever possible, the experiment duration with subjects was chosen to be at least 24 h.

Conclusions

The automated system with its intermittent calibration showed stable performance and can effectively be used on a 24-h/day, 7-day/wk basis. The system has a low risk of operating errors. Variations in ambient temperature and pressure have little effect because of the intermittent-calibration method.

The accuracy of the respiration chambers is dependent on the measurement interval (duration) and the level of $\dot{V}O_2$ and $\dot{V}CO_2$ to be measured. In our setting, the accuracy, described as a mean error $\pm$ SD, is 2.1 ± 7.4 and 1.3 ± 3.7 ml/min for O_2 and CO_2 , respectively, for intervals ≤ 2 h. Calculated energy expenditure has an accuracy of $0.7 \pm 2.3\%$ for an adult consuming 300 ml/min O_2 .

On the basis of our experiments, the smallest time interval needed to measure a subject was 15–25 min. When measuring plateau values (constant metabolic rate), one should wait a few minutes (≥ 5 min) after changing the plateau to accommodate the mixing time constant. The smallest time constant possible was determined, for the most part, by the mixing properties of the chamber, but the smallest practical measurement interval (duration 0.5 h) was determined by volume and gas-analysis accuracy.

APPENDIX A

Glossary

A	Analyzer-output uncalibrated value
c	Chamber
cal	Calibration
F	Volumetric fraction of gas (STPD)
g	Any gas
i	Incoming
o	Outgoing
P	Pressure
r	Recirculation
Rh	Relative humidity
s	Time-derivative operator, d/dt
sat	Saturated with water vapor
T	Temperature
t	Time
t_{90}	Response time to 90%
τ	Time constant
τ_d	Delay time
V	Volume (STPD)
$\dot{V}$	Volumetric flow rate (STPD)
w	Water vapor

Mixing Process

Response time is understood, in general, to be the time needed for the outlet of a process to reach at least 90% of final value after a step change at the inlet. Percentages used to define the response time vary: 90% (t_{90}), 95% (t_{95}), and 99% (t_{99}) are often used. A response time may be the result of complex higher-order terms. In contrast, the time constant (τ) associated with a first-order process of type $Y(t) = X \cdot [1 - e^{-(t/\tau)}]$ or, better, in process notation $H = 1/(\tau s + 1)$, is well defined; e.g., it takes 3τ to reach 95% and 10τ to reach 99.995% (14.5-bit resolution) of final value.

The first-order system $1/(\tau_{c1}s + 1)$ normally associated with a respiration chamber is only valid for a completely mixed volume; in reality, there will always be a small initial mixing interval before a subject's air is well mixed. With the use of recent fast-response respiration chambers, the measurement interval approaches the mixing interval, raising the question

Fig. 7. Diagram of 1st-order system $1/(\tau_{c1}s + 1)$ normally associated with a respiration chamber augmented with higher-order terms (τ_{c2} and τ_{dc}) for mixing. Recirculation flow through chamber can be thought to be sum of several (n) partial pathways, each of which has its own delay time τ_{dn} and, where confined space is present, a time constant τ_n (τ_{dc} and τ_{c2} are a complex composite of values $\tau_{d1 \sim n}$ and $\tau_{1 \sim n}$). Subject's respiration and possibly leakage take place in some of pathways, depending on subject's position in chamber. For clarity, subject's respiration and confined volume influence have been drawn only in a single pathway. $\dot{V}O_2$ and $\dot{V}CO_2$, O_2 uptake and CO_2 production, respectively.

of how the mixing interval fits into the equation and which factors affect it.

The recirculation flow through the chamber (if evenly distributed) can be thought to be the sum of several (n) partial flows or pathways (Fig. 7), each behaving as a tube reactor. Because diffusion in a tube reactor (backward or axial mixing) is beneficial to mixing and we are interested only in the problematic dominant factors, we chose to consider each flow as an ideal tube reactor with a delay time ($e^{-\tau_{ds}}$). If a partial flow m passes a confined space, an additional time constant [$1/(\tau_{ms}+1)$] is added for that pathway. The subject's expired air enters one or more of the pathways. Leakage will also affect one or more of the pathways, possibly where the subject respires. In a negative-pressure chamber, the direction of the leakage flow prevents loss of subject air and has the same effect as fresh air normally entering the chamber.

Determination of each parameter involved (including position and level of energy expenditure) is difficult; however, most higher-order natural processes can be simplified to the form $Ke^{-\tau_{ds}}/[(\tau_1s+1) \cdot (\tau_2s+1)]$. This is usually also the maximum number of parameters that can be determined from measurement of standard input signals (pulse and step). When the simplified higher-order system $Ke^{-\tau_{ds}}/[(\tau_{c1}s+1) \cdot (\tau_{c2}s+1)]$ with a respiration chamber $\bar{K} = 1$ (response at $t = \infty$) is used, τ_{dc} and τ_{c2} are a complex composite of values τ_{d1-n} and τ_{1-n} from the n partial flows, and the dominant time constant τ_{c1} is determined by volume and flow (V_o/V_c). In the case of evenly distributed flow, τ_{dc} will be mostly determined by volume and recirculation flow ($V_c/\bar{V}_r$) and by the position of the subject in the chamber (i.e., at midpoint $0.5 \cdot V_c/\bar{V}_r$). τ_{c2} is the result of backward and axial mixing and exchange with confined spaces. If confined spaces are avoided, τ_{c2} will be very small. Determining chamber characteristics from standard input signals (pulse and step) should take into account possible variation of τ_{dc} and τ_{c2} .

APPENDIX B

Calculation of $\dot{V}O_2$ and $\dot{V}CO_2$

We define two moments in time: t_1 is the start time of a single measurement, and t_2 is the end time of the measurement, i.e., in our case, $t_2 - t_1 \geq 5$ min, the smallest usable interval with our sample sequence. A value at t_1 , t_2 , or t_{1+2} (average over interval t_1 to t_2) represents the best value calculated from multiple samples for that point in time. Algorithms used ensure that summing values calculated over small intervals are mathematically identical to calculation over one long interval.

The frequent calibration technique allows pressure- and temperature-independent calculations of momentary gas concentration

$$F_g(t) = F_{cal} \cdot [A_g(t) - A_0(t)] / [A_{cal}(t) - A_0(t)]$$

For calculation of $\dot{V}O_2$ and $\dot{V}CO_2$, the following parameters can be derived directly

$$F_{iN_2}(t) = 100 - F_{iO_2}(t) - F_{iCO_2}(t)$$

$$F_{oN_2}(t) = 100 - F_{oO_2}(t) - F_{oCO_2}(t)$$

$P_{wsat}(t)$ is derived from $T_c(t)$ by means of a lookup table.

$$P_w(t) = Rh_c(t) \cdot P_{wsat}(t) / 100$$

$$\dot{V}_o(t) = \dot{V}_{ATP}(t) \cdot [P_o(t) - P_w(t)] / [1013.25 \cdot (1 + 0.00367 \cdot T_o(t))]$$

$$V_c(t) = V_{cATP} \cdot [P_c(t) - P_w(t)] / [1013.25 \cdot (1 + 0.00367 \cdot T_c(t))]$$

$$\dot{V}_{oO_2}(t) = \dot{V}_o(t) \cdot F_{oO_2}(t) / 100$$

$$\dot{V}_{oCO_2}(t) = \dot{V}_o(t) \cdot F_{oCO_2}(t) / 100$$

$$\dot{V}_{iN_2}(t) = \dot{V}_o(t) \cdot F_{iN_2}(t) / 100 = \dot{V}_o(t) - \dot{V}_{oO_2}(t) - \dot{V}_{oCO_2}(t)$$

Next, parameters have to be derived that involve $\dot{V}_i(t)$, the flow of the input in STPD. $\dot{V}_i(t)$ may be calculated by using the "haldane" correction. The formula used must incorporate changing N_2 fractions and STPD correction of the chamber volume as a function of time

$$\dot{V}_i(t_{1+2}) = [\dot{V}_o(t_{1+2}) \cdot F_{oN_2}(t_{1+2}) + V_c(t_2) \cdot F_{oN_2}(t_2) - V_c(t_1) \cdot F_{oN_2}(t_1)] / F_{iN_2}(t_{1+2})$$

Now that $\dot{V}_i(t)$ is known, the following parameters are derived

$$\dot{V}_{iO_2}(t_{1+2}) = \dot{V}_i(t_{1+2}) \cdot F_{iO_2}(t_{1+2}) / 100$$

$$\dot{V}_{iCO_2}(t_{1+2}) = \dot{V}_i(t_{1+2}) \cdot F_{iCO_2}(t_{1+2}) / 100$$

$$\dot{V}_{cO_2}(t_{1+2}) = [V_c(t_2) \cdot F_{cO_2}(t_2) - V_c(t_1) \cdot F_{cO_2}(t_1)] / [100 \cdot (t_2 - t_1)]$$

$$\dot{V}_{cCO_2}(t_{1+2}) = [V_c(t_2) \cdot F_{cCO_2}(t_2) - V_c(t_1) \cdot F_{cCO_2}(t_1)] / [100 \cdot (t_2 - t_1)]$$

This leaves only $\dot{V}O_2$ and $\dot{V}CO_2$ to be calculated

$$\dot{V}O_2(t_{1+2}) = \dot{V}_{iO_2}(t_{1+2}) - \dot{V}_{cO_2}(t_{1+2}) - \dot{V}_{oO_2}(t_{1+2})$$

$$\dot{V}CO_2(t_{1+2}) = \dot{V}_{oCO_2}(t_{1+2}) + \dot{V}_{cCO_2}(t_{1+2}) - \dot{V}_{iCO_2}(t_{1+2})$$

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REFERENCES

1. **Atwater, W. O., and F. G. Benedict.** *Experiments on the Metabolism of Matter and Energy in the Human Body*. Washington, DC: US Government Printing Office, 1903. (USDA Office of Experiment Stations Bull. No. 136)
2. **Benedict, F. G.** A helmet for use in clinical studies of gaseous metabolism. *N. Engl. J. Med.* 203: 150–158, 1930.
3. **Brown, D., T. J. Cole, M. J. Dauncey, R. W. Marrs, and P. R. Murgatroyd.** Analysis of gaseous exchange in open-circuit indirect calorimetry. *Med. Biol. Eng. Comput.* 22: 333–338, 1984.
- 3a. **Centre de Recherche Claud-Delorme de L'Air Liquide.** *Encyclopedie des Gaz*. Amsterdam: Elsevier, 1976.
4. **Charbonnier, A., C. D. R. Jones, Y. Schutz, P. R. Murgatroyd, R. G. Whitehead, E. Jéquier, and G. Spinnler.** A whole body transportable indirect calorimeter for human use in the tropics. *Eur. J. Clin. Nutr.* 44: 725–731, 1990.
5. **Dauncey, M. J., P. R. Murgatroyd, and T. J. Cole.** A human calorimeter for the direct and indirect measurement of 24 h energy expenditure. *Br. J. Nutr.* 39: 557–566, 1978.
6. **Henning, B., R. Löfgren, and L. Sjöström.** Chamber for indirect calorimetry with improved transient response. *Med. Biol. Eng. Comput.* 34: 207–212, 1996.
7. **Heysfield, S. B., D. B. Allison, F. X. Pi-Sunyer, and Y. Sun.** Columbia respiratory-chamber indirect calorimeter: a new approach to air-flow modelling. *Med. Biol. Eng. Comput.* 32: 406–410, 1994.
8. **Jéquier, E., and Y. Schutz.** Long-term measurements of energy expenditure in humans using a respiration chamber. *Am. J. Clin. Nutr.* 38: 989–998, 1983.

9. **Kinney, J. M., A. P. Morgan, F. J. Domingues, and K. J. Gildner.** A method for the continuous measurement of gas exchange and expired radioactivity in acutely ill patients. *Metabolism* 13: 205–211, 1964.
10. **McLean, J. A., and P. R. Watts.** Analytical refinements in animal calorimetry. *J. Appl. Physiol.* 40: 827–831, 1976.
11. **Moon, J. K., C. L. Jensen, and N. F. Butte.** Fast-response whole body indirect calorimeter for infants. *J. Appl. Physiol.* 74: 476–484, 1993.
12. **Moon, J. K., F. A. Vohra, O. S. Valerio Jimenez, M. R. Puyau, and N. F. Butte.** Closed-loop control of carbon dioxide concentration and pressure improves response of room respiration calorimeters. *J. Appl. Physiol.* 125: 220–228, 1995.
13. **Murgatroyd, P. R., H. L. Davies, and A. M. Prentice.** Intra-individual variability and measurement noise in estimates of energy expenditure by whole body indirect calorimetry. *Br. J. Nutr.* 58: 347–356, 1987.
14. **Prentice, A. M., G. R. Goldberg, H. L. Davies, P. R. Murgatroyd, and W. Scott.** Energy-sparing adaptations in human pregnancy assessed by whole-body calorimetry. *Br. J. Nutr.* 62: 5–22, 1989.
15. **Ravussin, E., S. Lillioja, T. Anderson, L. Christin, and C. Bogardus.** Determinants of 24-hour energy expenditure in man: methods and results using a respiratory chamber. *J. Clin. Invest.* 78: 1568–1578, 1986.
16. **Rumpler, W. V., J. L. Seale, J. M. Conway, and P. W. Moe.** Repeatability of 24-h energy expenditure measurements in humans by indirect calorimetry. *Am. J. Clin. Nutr.* 51: 147–152, 1990.
17. **Seale, J. L., W. V. Rumpler, and P. W. Moe.** Description of a direct-indirect room-sized calorimeter. *Am. J. Physiol.* 260 (*Endocrinol. Metab.* 23): E306–E320, 1991.
18. **Shetty, P. S., M. L. Sheela, P. R. Murgatroyd, and A. V. Kurpad.** An open-circuit indirect whole body calorimeter for the continuous measurement of energy expenditure of man in the tropics. *Indian J. Med. Res.* 85: 453–460, 1987.
19. **Treuth, S. M., G. R. Hunter, R. L. Weinsier, and S. H. Kell.** Energy expenditure and substrate utilization in older women after strength training: 24-h calorimeter results. *J. Appl. Physiol.* 78: 2140–2146, 1995.
20. **Webb, P., and S. J. Troutman, Jr.** An instrument for continuous measurement of oxygen consumption. *J. Appl. Physiol.* 28: 867–871, 1970.
21. **Weir, J. B.** New methods for calculating metabolic rate with special reference to predicting protein metabolism. *J. Physiol. (Lond.)* 109: 1–9, 1949.