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Quantification of 3D microstructural parameters of trabecular bone is affected by the analysis software

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\textbf{ARTICLE INFO}

Keywords: Quantification, Microstructural parameters, Analysis software

\textbf{ABSTRACT}

Over the last decades, the use of high-resolution imaging systems to assess bone microstructural parameters has grown immensely. Yet, no standard defining the quantification of these parameters exists. It has been reported that different voxel size and/or segmentation techniques lead to different results. However, the effect of the evaluation software has not been investigated so far. Therefore, the aim of this study was to compare the bone microstructural parameters obtained with two commonly used commercial software packages, namely IPL (Scanco, Switzerland) and CTan (Bruker, Belgium). We hypothesized that even when starting from the same segmented scans, different software packages will report different results.

Nineteen trapezia and nineteen distal radii were scanned at two resolutions (20 μm voxel size with microCT and HR-pQCT 60 μm). The scans were segmented using the scanners' default protocol. The segmented images were analyzed twice, once with IPL and once with CTan, to quantify bone volume fraction (BV/TV), trabecular thickness (Tb.Th), trabecular separation (Tb.Sp), trabecular number (Tb.N) and specific bone surface (BS/BV).

Only small differences between IPL and CTan were found for BV/TV. For Tb.Th, Tb.Sp and BS/BV high correlations ($R^2 \geq 0.99$) were observed between the two software packages, but important relative offsets were observed. For microCT scans, the offsets were relative constant, e.g., around 15% for Tb.Th. However, for the HR-pQCT scans the mean relative offsets ranged over the different bone samples (e.g., for Tb.Th from 14.5% to 19.8%). For Tb.N, poor correlations (0.43 $\leq R^2 \leq 0.81$) for all tested cases were observed.

We conclude that trabecular bone microstructural parameters obtained with IPL and CTan cannot be directly compared except for BV/TV. For Tb.Th, Tb.Sp and BS/BV, correction factors can be determined, but these depend on both the image voxel size and specific anatomic location. The two software packages did not produce consistent data on Tb.N. The development of a universal standard seems desirable.

1. Introduction

The use of high-resolution imaging systems to assess bone microstructural parameters in animals and humans has grown immensely during the last decades [1,2]. Several commercial scanners are on the market, each one with its own analysis software. Yet, no standard exists either for quantification of the morphological measures, or for calibration of the analysis software. It has been reported that different voxel size and/or segmentation techniques lead to different results and hence that it is not possible to compare the absolute values of bone parameters obtained from different voxel size scans and/or segmentation techniques [3–6].

In commercially available scanners, two voxel size ranges are commonly used for imaging and quantification of bone microstructural parameters for human beings. For \textit{ex vivo} analyses, a voxel size of 10 − 20 μm is commonly accepted [7] as achieved by micro-computed tomography (microCT). State-of-the-art human \textit{in vivo} scanners can reach a somewhat lower resolution. For \textit{in vivo} investigations, the high resolution peripheral computed tomography (HR-pQCT) technology offers the highest resolution, with the XtremeCT-II scanner (Scanco...
Medical AG, Switzerland) providing a reconstructed isotropic voxel size of 60.7 μm. Next to HR-pQCT, cone-beam computed tomography (CBCT) can be used for high-resolution \textit{in vivo} imaging, but at a slightly lower resolution (75 μm voxel size for the NewTom 5G scanner (Cefla, Italy)) [8].

Several software packages exist for a quantitative evaluation of the images acquired by the scanners. Thus far, the results delivered by those software packages have not been subjected to direct comparison. Nevertheless, this comparison would be important to correctly interpret and compare the results of various research studies. Hence, the aim of this study was to compare the bone microstructural parameters calculated with different software packages. We opted to compare the two most widely used commercial software packages, more specifically, the Image Processing Language (IPL) software implemented in Scanco systems [9] and the CTan software delivered with Bruker scanners [10,11].

The second aim was to evaluate the effect that different voxel size and anatomical locations could have on the comparison. We hypothesized that, even when starting from the same segmented images, the results of the two analysis software packages would show differences in the calculated bone microstructural parameters due to different implementations and quantification methods.

2. Materials & methods

2.1. Sample collection

The software comparison was performed on two different bones of the wrist, namely the trapezium, being one of the carpal bones, and the distal radius that is the most commonly investigated peripheral site in osteoporosis research. Two sets of human cadaveric bone samples had been collected in previous studies [9,10]. In short, nineteen trapezia (18 female, 1 male; 7 left, 12 right) of severe osteoarthritic patients were harvested via trapeziectomy by a hand surgeon (FS) in the hospital AZ Groeninge (Kortrijk, Belgium). The age of the donors ranged from 53 to 76 years (mean ± standard deviation (SD): 63.4 ± 7.0 years). All donors consented to use their data and their extracted specimen for anonymous research use. No extra medical interventions were done for this study. In addition, nineteen cadaveric radii (14 females, 5 males; 8 left, 11 right) were obtained from Science Care (USA). Donor age ranged from 25 to 93 years (mean ± SD: 67.9 ± 16.2 year). All samples were conserved fresh-frozen at ~20° Celsius.

2.2. Image acquisition and processing

Each specimen was scanned twice: at a high resolution close to 20 μm voxel size and at a moderate resolution close to 60 μm voxel size (Fig. 1). Specifically, the trapezia were scanned at a voxel size of 19.84 μm using an \textit{ex vivo} microCT-scanner (Skyscan 1172, Bruker, Belgium), and at a voxel size of 60.7 μm using a HR-pQCT device (XtremeCT-II, Scanco Medical AG, Switzerland). The radii were scanned at voxel sizes of 19 μm and 60.7 μm using a small animal microCT scanner (VivaCT40, Scanco Medical AG, Switzerland) and the XtremeCT-II, respectively. All images were reconstructed and segmented using the standard software and the default settings of the respective scanner [8–10].

For the trapezia, the whole trabecular bone compartment was used for the analyses. For the radii, two 9 mm sections were defined as volumes of interest (VOI) (Fig. 1). Specifically, the trapezia were scanned at a voxel size of 19.84 μm using an \textit{ex vivo} microCT-scanner (Skyscan 1172, Bruker, Belgium), and at a voxel size of 60.7 μm using a HR-pQCT device (XtremeCT-II, Scanco Medical AG, Switzerland). The radii were scanned at voxel sizes of 19 μm and 60.7 μm using a small animal microCT scanner (VivaCT40, Scanco Medical AG, Switzerland) and the XtremeCT-II, respectively. All images were reconstructed and segmented using the standard software and the default settings of the respective scanner [8–10].

For the trapezia, the whole trabecular bone compartment was used for the analyses. For the radii, two 9 mm sections were defined as volumes of interest. The first section was selected strictly adjacent to the most proximal point of the subchondral endplate and perpendicular to the long bone axis and will be addressed as ‘subchondral section’ in this paper. The second section was selected directly distal to the first section and corresponded closely to the measurement area recommended for clinical scanning, termed ‘standard section’ throughout this work. The trabecular volumes of interest (VOI) were selected automatically on the microCT or vivaCT images using an in-house C++ script which was based on a method developed by Buie et al. [9,11]. The corresponding VOIs of the HR-pQCT images were identified by image registration and transferring the microCT masks to the HR-pQCT scans. In more detail, the HR-pQCT images were spatially registered to the microCT images by a rigid voxel-based gray value registration in AMIRA software package (v6.2, Thermo Fisher Scientific, USA). The obtained transformation matrix was used to transform the microCT mask to the HR-pQCT.
2.3. Calculation of trabecular bone microstructural parameters

Starting from the segmented images, bone microstructural parameters were calculated with the IPL software (Scanco Medical AG) as well as with the software CTan (Bruker). It is important to note that, for a given specimen and scan resolution/voxel size, both software packages were provided with the same segmented images. For both software packages, the standard 3D direct analysis method was used.

### Table 1

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Function</th>
<th>Technique</th>
<th>Manual</th>
<th>Explanation of calculation</th>
</tr>
</thead>
<tbody>
<tr>
<td>BV/TV</td>
<td>vox_scanco_param and tri_da_metric for MicroCT or vox_scanco_param for XtremeCT-II</td>
<td>Distance transform [15]</td>
<td>No explanation</td>
<td>Total volume of each binarized 3D object within the VOI divided by the total volume of the VOI. The 3D volume measurement is based on the marching cube volume model.</td>
</tr>
</tbody>
</table>
| Th.Th              | dt_object_param for MicroCT or dt_thickness for XtremeCT-II | Distance transform [15]       | No explanation       | Local thickness definition is based on the definition of Hildebrand and Ruegsegger [15]. They defined local thickness for a point in solid as the diameter of the largest sphere which fulfills two conditions:  
- The sphere encloses the point (but the point is not necessarily the center of the sphere)  
- The sphere is entirely bounded within the solid surfaces. The base of the implementation is the distance transform method described by Remy and Thiel [20]. |
| BS/BV              | tri_da_metric_db                               | Marching cube [14]            | No explanation       | The 3D surface measurement is based on triangulation (marching cubes model) of the segmented object. |

### Table 2

<table>
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<tr>
<th>Parameters</th>
<th>vox_scanco_param</th>
<th>dt_object_param</th>
<th>dt_thickness</th>
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</tbody>
</table>

Used for

| MicroCT | x | x | x | x | x | x | x |
| XtremeCT-II | x | x | x | x | x | x | x |

<table>
<thead>
<tr>
<th>Parameter</th>
<th>BV/TV</th>
<th>Th.Th</th>
<th>Th.Th</th>
<th>Th.Sp</th>
<th>Th.Sp</th>
<th>Th.N</th>
<th>Th.N</th>
<th>BS/BV</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>x</td>
<td>x</td>
<td>x</td>
<td>x</td>
<td>x</td>
<td>x</td>
<td>x</td>
<td>BV/TV</td>
</tr>
</tbody>
</table>

images in Matlab R2016a (The Mathworks, United States).

2.3. Calculation of trabecular bone microstructural parameters

Starting from the segmented images, bone microstructural parameters were calculated with the IPL software (Scanco Medical AG) as well as with the software CTan (Bruker). It is important to note that, for a given specimen and scan resolution/voxel size, both software packages were provided with the same segmented images. For both software packages, the standard 3D direct analysis method was used.
The evaluated trabecular bone parameters included bone volume fraction (BV/TV), trabecular thickness (Tb.Th), trabecular separation (Tb.Sp), trabecular number (Tb.N - named Tb.N° in Scanco’s user manual) and specific bone surface (BS/BV) [12]. The calculation techniques following the software’s manual are summarized in Table 1. For the IPL algorithms, the standard settings as recommended by Scanco were used (Table 2). Remark that Scanco is using different settings for microCT as for XtremeCT-II. Hence, different results will be obtained when evaluating the same data with the microCT or with the XtremeCT-II script. For CTan, there were no adjustable parameters of the evaluation procedure.

### Table 3

<table>
<thead>
<tr>
<th></th>
<th>Mean IPL</th>
<th>$R^2$</th>
<th>Relative offset</th>
<th>Mean IPL</th>
<th>$R^2$</th>
<th>Relative offset</th>
<th>Mean IPL</th>
<th>$R^2$</th>
<th>Relative offset</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>MicroCT scans</strong></td>
<td></td>
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<td></td>
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<td></td>
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</tr>
<tr>
<td>BV/TV (%)</td>
<td>23.41(2.75)</td>
<td>1.00</td>
<td>−0.18%</td>
<td>16.03(3.36)</td>
<td>1.00</td>
<td>−0.28%</td>
<td>12.45(3.89)</td>
<td>1.00</td>
<td>−0.37%</td>
</tr>
<tr>
<td>BV/TV* (%)</td>
<td>23.75(2.84)</td>
<td>1.00</td>
<td>−1.61%</td>
<td>16.23(3.44)</td>
<td>1.00</td>
<td>−1.51%</td>
<td>12.58(3.97)</td>
<td>1.00</td>
<td>−1.37%</td>
</tr>
<tr>
<td>Tb.Th (mm)</td>
<td>0.16(0.02)</td>
<td>1.00</td>
<td>14.76%</td>
<td>0.15(0.01)</td>
<td>1.00</td>
<td>15.04%</td>
<td>0.15(0.01)</td>
<td>1.00</td>
<td>14.89%</td>
</tr>
<tr>
<td>Tb.Sp (mm)</td>
<td>0.58(0.07)</td>
<td>0.99</td>
<td>4.99%</td>
<td>0.70(0.11)</td>
<td>1.00</td>
<td>6.30%</td>
<td>0.82(0.18)</td>
<td>1.00</td>
<td>4.64%</td>
</tr>
<tr>
<td>Tb.N (1/mm)</td>
<td>1.78(0.18)</td>
<td>0.66</td>
<td>−26.70%</td>
<td>1.45(0.22)</td>
<td>0.79</td>
<td>−35.00%</td>
<td>1.26(0.26)</td>
<td>0.81</td>
<td>−42.06%</td>
</tr>
<tr>
<td>BS/BV (1/mm)</td>
<td>16.67(1.97)</td>
<td>0.99</td>
<td>24.30%</td>
<td>17.76(1.71)</td>
<td>0.99</td>
<td>18.49%</td>
<td>18.29(1.85)</td>
<td>1.00</td>
<td>19.08%</td>
</tr>
<tr>
<td><strong>XtremeCT scans</strong></td>
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</tr>
<tr>
<td>BV/TV (%)</td>
<td>35.67(4.67)</td>
<td>1.00</td>
<td>−0.75%</td>
<td>17.53(4.99)</td>
<td>1.00</td>
<td>−2.00%</td>
<td>13.72(0.05)</td>
<td>1.00</td>
<td>−2.46%</td>
</tr>
<tr>
<td>Tb.Th (mm)</td>
<td>0.28(0.03)</td>
<td>1.00</td>
<td>19.79%</td>
<td>0.21(0.01)</td>
<td>1.00</td>
<td>15.07%</td>
<td>0.21(0.02)</td>
<td>1.00</td>
<td>14.53%</td>
</tr>
<tr>
<td>Tb.Sp (mm)</td>
<td>0.60(0.07)</td>
<td>0.98</td>
<td>13.18%</td>
<td>0.81(0.15)</td>
<td>1.00</td>
<td>11.79%</td>
<td>0.98(0.30)</td>
<td>1.00</td>
<td>8.80%</td>
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<tr>
<td>Tb.N (1/mm)</td>
<td>1.60(0.15)</td>
<td>0.43</td>
<td>−32.46%</td>
<td>1.24(0.20)</td>
<td>0.80</td>
<td>−44.99%</td>
<td>1.07(0.25)</td>
<td>0.81</td>
<td>−48.19%</td>
</tr>
</tbody>
</table>

* IPL-based parameter and CTan-based parameter are significant different (5% significance level) following the Kolmogorov-Smirnov test.

The aim of this study was to compare bone microstructural parameters obtained with both software packages, linear regression analysis was performed. The coefficient of determination ($R^2 = (Pearson\ correlation\ coefficient)^2$) which describes the goodness of the fit on a line (linear regression) and relative offset ($mean(\frac{CTan - IPL}{IPL})$) which describes the distance from the ideal line through 0 and with a slope of 1, were evaluated.

Furthermore, trendlines and Bland–Altman plots were generated to assess how well the software packages correspond as well as to assess the influence of the anatomical regions on this correspondence. The two-sample Kolmogorov-Smirnov test was used to evaluate for each bone parameter and for each anatomical region whether the values of IPL and CTan were significantly different at 5% significance level [13].

### 4. Discussion

The aim of this study was to compare bone microstructural parameters as calculated by the software packages IPL and CTan, and to assess their agreement at ex vivo microCT resolution (~20 μm voxel size) as well as at in vivo HR-pQCT resolution (~60 μm voxel size). A strength of our approach was that identical segmented images were used when comparing IPL to CTan. This prevented that differences in image segmentation could have affected the outcome of our study as a confound factor; hence, the identified differences in the quantified bone parameters were due only to differences in the software implementations for calculation of those parameters.

The results obtained for BV/TV were highly similar for both software packages ($R^2 = 1$ and relative offsets $\leq 2.46\%$), despite the differences in the software implementations. The implementation used in IPL is mainly based on voxel counting, whereas the implementation in CTan is based on the surface mesh model triangulated with the marching cubes algorithm [14]. For microCT, Scanco also implemented a marching cube based algorithm to calculate BV/TV (indicated as BV/TV* in this manuscript), however, this did not improve the relative offset between the software packages. High correlations ($R^2 \geq 0.98$) were found for Tb.Th, Tb.Sp and BS/BV, but the obtained absolute values were higher when CTan was used and these differences reached statistical significance for Tb.Th and BS/BV. As both software packages are based on similar theoretical backgrounds (distance transformation for Tb.Th and Tb.Sp, marching cube for BS/BV) for those parameters, we hypothesize that the respective implementations are slightly different. For Tb.N, only poor correlations were obtained; this can be explained by the different methodologies used to calculate this parameter. The IPL implementation utilizes the distance transform [15], whereas CTan’s calculation uses the parallel plate model [16,17]. It has
been demonstrated that the parallel plate model introduces a volume-dependent offset for Tb.Th and Tb.Sp [18]. We suggest to follow the reporting recommendations made by Bouxsein et al. in 2010 (Guide lines for assessment of bone microstructure in rodents using micro-computed tomography) [12] which are being adopted by many in the research community. They proposed to calculate Tb.N with the distance transform method as the inverse of the mean distance between the mid-axis of the structure. This corresponds to the IPL implementation.

For the microCT-based analyses, the relative offsets for Tb.Th and Tb.Sp were fairly constant, ranging from 14.8% to 15.0%, and from 4.6% to 6.3%, respectively. This suggests that correction factors could be used to compensate for the different implementations of the analyzing software for those parameters. Remark that those correction factors only compensate for the difference in the analyzing software and do not include other aspects which can cause discrepancies in the results, such as segmentation and voxel size which are explained later in this manuscript. We suggest to use a correction factor of 1.15 (1 + relative offset) for Tb.Th and 1.05 for Tb.Sp to convert data obtained with IPL to CTan. For the HR-pQCT-based analyses, relative offsets were higher (ranging from 14.5% to 19.8% for Tb.Th and from 8.8% to 13.2% for Tb.Sp) and calculated correction factors differed per anatomical location; hence, these need to be determined for each specific anatomical site. These findings for HR-pQCT can be explained by the lower resolution of the scanner, where even small differences in both the implementation procedure and the imaged bone microstructure will have a larger effect on the quantified microstructural parameters.

In this study, we demonstrated that even when starting from the same segmented images, different software packages using default settings calculate significantly different results for bone microstructural parameters. This problem could be solved by standardizing the analysis. The lack of standardization of the analysis software is not the only reason limiting the direct comparison between absolute values of bone microstructural parameters.

In Fig. 2, we show regression plots and Bland Altman plots for the microCT scans analyzed once with IPL (Scanco Medical AG, Switzerland) and CTan (Bruker, Belgium) for bone volume fraction (BV/TV), trabecular thickness (Tb.Th), trabecular separation (Tb.Sp), trabecular number (Tb.N) and specific bone surface (BS/BV). The ‘subchondral section’ is the section strictly adjacent to the most proximal point of the subchondral endplate. The ‘standard section’ is the section proximal to the ‘subchondral section’ and corresponds to the standard patient measurement area. The solid line on the regression plot indicates the line y = x.
scanner as well as the scanning protocol will have an important influence. It has e.g. already been thoroughly demonstrated that the image voxel size has a significant influence on the calculated absolute values of the bone microstructural parameters [3,5,6]. Standardization of the scanner as well as the scanning protocol should not be aimed for, because these are application specific; e.g., in vivo scanning will typically require short scanning times and low radiation dose, whereas achieving a high level of detail may be critical in an ex vivo scan.

Another factor influencing the absolute values of the bone microstructural parameters is the segmentation procedure. Also here, standardization of the segmentation process is not desired as specific filtering operations and adaptive segmentation techniques may be required to reduce the noise and to improve the contrast.

Also the well-accepted gold standard to calculate bone microstructural parameters, namely microCT [7], is not standardized (e.g., voxel size; segmentation protocol) This scanner is used by most research groups to validate other scanners and segmentation techniques. No standard protocol exists for either the scanner device, the used resolution/voxel size, or the segmentation procedure.

The authors believe that there is an important need for the bone research community to define a ‘gold standard’ to which scanners and their respective analysis software can be calibrated. This can be achieved by developing bone-like phantoms of trabecular and cortical bone and this of human bone samples as well as of animals (rodents, big animal) with well specified properties to be used for calibration. Like this, researchers and clinicians can use the scanner, segmentation and analysis method of their choice, which allow optimal usage and further innovation of all components, but at the same time parameters are quite well standardized. On short term, the authors advise to use, if possible, the standard methods of the scanners, to calibrate all scans to Hounsfield units and to report in detail scanning, segmentation, analysis methods as well as correction factors (if used). This will not make everything standardized, but this makes it at least reproducible. Specifically, if compared to a gold standard microCT-image, we advise to do this against a 20μm microCT scan which is segmented with a global threshold. The correction values calculated in this paper could be used as correction values to convert from Bruker to Scanco and vice versa. We advise to determine own correction values when using other analysis software.

A limitation of the current study is that our findings may not be representative for other sites and image resolutions/voxel sizes. Moreover, only two commercial software packages were compared in this study. However, these two software packages already show large differences, which demonstrates the need for standardization. A third limitation is the limited sample size subjected in our study. The used specimens may not represent a larger population and thus the results cannot be generalized. Fourth, the current investigation was performed in an ex vivo environment and this may have influenced the results. The image quality is expected to be superior compared to what is achievable under in vivo conditions. Another limitation is that only a restricted set of morphological parameters was analyzed. Other parameters such as connectivity, skeletal muscle index (SMI), etc. were not investigated. This study could still be extended by overcoming the above-mentioned limitations; however, this extension would not alter the core message. With this investigation we intend to highlight the need for standardization to enable comparison of bone microstructural parameters between different scanners and studies in the future.

5. Conclusion

With this study we demonstrated that trabecular bone microstructural parameters obtained with IPL and CTan cannot be directly compared except for BV/TV. Good correlations were obtained between the two software packages for Tb.Th, Tb.Sp and BS/BV; however,
correction factors need to be applied to match their absolute values. General correction factors could be determined with regard to Tb.Th and Tb.Sp for the microCT scans, but not for the HR-pQCT scans. Hence, for in vivo HR-pQCT scanning, we advise to calculate different correction factors for different anatomical locations. The outcomes for Tb.N did not correlate well between IPL and CTan for our tested cases. The development of a universal standard for calculation of bone microstructural parameters seems desirable, not only for calibration of different software packages to each other, but also for compensation of other aspects, such as resolution/voxel size, scanner type and segmentation technique.

CRediT authorship contribution statement

Karen Mys: Conceptualization, Methodology, Software, Validation, Formal analysis, Investigation, Data curation, Writing - original draft, Writing - review & editing, Visualization, Project administration, Funding acquisition. Peter Varga: Conceptualization, Investigation, Resources, Data curation, Supervision. Filip Stockmans: Conceptualization, Resources, Supervision. Boyko Gueorguiev: Conceptualization, Resources, Supervision. Caroline E. Wyers: Investigation, Resources. Joop P.W. van den Bergh: Investigation, Resources. G. Harry van Lenthe: Conceptualization, Data curation, Supervision, Project administration, Funding acquisition.

Declaration of competing interest

Karen Mys, Peter Varga, Filip Stockmans, Boyko Gueorguiev, Caroline E. Wyers, Joop P.W. van den Bergh and G. Harry van Lenthe declare that they have no conflict of interest.

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References


