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Citation for published version (APA):

Hemker, H. C., Willems, G. M., & Beguin, S. (1986). A computer assisted method to obtain the prothrombin activation velocity in whole plasma independent of Thrombin decay processes. *Thrombosis and Haemostasis*, 56(1), 9-17. <https://doi.org/10.1055/s-0038-1661594>

Document status and date:

Published: 01/01/1986

DOI:

[10.1055/s-0038-1661594](https://doi.org/10.1055/s-0038-1661594)

Document Version:

Publisher's PDF, also known as Version of record

Please check the document version of this publication:

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A Computer Assisted Method to Obtain the Prothrombin Activation Velocity in Whole Plasma Independent of Thrombin Decay Processes

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Key words

Prothrombin activation velocity – Thrombin generation velocity – Antithrombin activities – α_2 -Macroglobulin

Summary

A method is described that, on the basis of the time course of amidolytic activity after the triggering of thrombin generation in normal plasma, allows the calculation of the velocity of prothrombin conversion independent of thrombin inactivating processes.

It is shown how the reaction constants for the α_2 M-dependent and the α_2 M-independent thrombin inactivation processes can be obtained in a sample of whole plasma.

The method is verified by demonstrating that the experimentally observed time courses of residual prothrombin and of α_2 M-thrombin complex coincide with those calculated from the time course of amidolytic activity, and by showing that the course of prothrombin conversion in plasma without α_2 -macroglobulin or AT III is adequately described if the α_2 M or AT III-dependent breakdown constants are taken zero in the calculations.

It appears that the inactivation of thrombin, endogenously generated in whole plasma, is about half as fast as that of exogenous thrombin added to the plasma.

A computer program is presented that carries out the relevant calculations.

Introduction

The thrombin generation curve is one of the oldest tools of the coagulation trade. It is the basis of the so-called two-stage prothrombin estimation (1, 2, 3) but its roots go as far back as the very discovery of prothrombin (4). The rise and fall of thrombin concentration after triggering of coagulation in plasma results from the combined activity of the prothrombin activating enzyme complex (prothrombinase) and the thrombin inactivating processes, i. e. the binding of thrombin by antithrombins, such as antithrombin III and α_2 -macroglobulin (5, 6).

The general shape of the curve shows a short lag time and a fairly quick rise. After the maximum has been reached a gradual decline is observed that, if the clotting of fibrinogen is used to estimate thrombin, will eventually approach a zero level. Because the complex of a protease with α_2 -macroglobulin has a persistent activity towards small molecular weight substrates (6–10) the final level of activity will not be zero if the thrombin concentration is determined with the aid of chromogenic substrates.

The classical thrombin generation test suffers from an important drawback: the samples from the mixture in which thrombin is generated have to be tested immediately on a fibrinogen containing reagent. Estimation of low concentrations of thrombin tends to be very imprecise and time-consuming when clotting methods are used. It therefore is virtually impossible to achieve in this way a sampling pattern with enough measuring points and of sufficient accuracy over the whole time range. A detailed analysis of the thrombin generation curve obtained by this method is therefore impossible.

This article describes a method to obtain thrombin generation curves with sampling intervals that can be as small as 5 sec. The thrombin is measured by its action on a chromogenic substrate. It is shown how these curves can be used to compute the time course of the process that generates thrombin (prothrombinase) independently of the processes that inactivate thrombin (antithrombins). These calculations can be carried out with the aid of a personal computer.

The following abbreviations will be used: AT III = Antithrombin III; α_2 M = α_2 -macroglobulin; Tris = tris (hydroxymethyl)aminomethane; EDTA = Ethylenediamino tetracetate; SBTI = Soybean Trypsin Inhibitor.

Materials and Methods

Plasmas

Blood from healthy donors was collected on 0.13 M trisodium citrate; nine parts of blood to one part of citrate solution.

A first and a second centrifugation were performed at 3,000 g, at 15° C for 15 min. A third centrifugation was done at 4° C, for one hour at 23,000 g. The platelet free plasma thus obtained was stored at –80° C. It was checked that the clotting factors and the antiproteases were in the normal range. The clotting factors were determined according to refs. 11 and 12. Radial immuno diffusion was used to estimate α_2 -macroglobulin and α_1 -antitrypsin (M. Partigen, Behring Werke, Germany). Antithrombin III was determined according to ref. 13.

Defibrinated plasma was obtained by mixing an aliquot of plasma with 1:500 volume of a reptilase solution, letting a clot form for 10 min at 37° C and keeping the clotted plasma at 0° C for 10 min. The fibrin formed was discarded by centrifugation (10 min, 5000 g, 0° C) or by winding on a small plastic spatula. The concentrations of factors II, V, VII, VIII, IX, X, XI, and XII did not significantly change by the reptilase treatment. For the factors V and VIII series of dilutions (1–10%) of the plasma before and after the treatment were compared. They showed clotting times that were identical within the limits of experimental error thus excluding the possibility that a combined effect of activation and inactivation of the factors accidentally mimicked as identical clotting factors.

The euglobulin fraction from defibrinated plasma was obtained by acid precipitation at low ionic strength as described in ref. 11. The precipitate was dissolved in Michaelis buffer pH 7.35 containing 0.02 M trisodium citrate, so as to obtain a concentration of the clotting factor II, V, VII and X that was not lower than 50%. No antithrombin activity could be demonstrated in this mixture.

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Thrombin was formed in the euglobulin fraction by the addition of human thromboplastin (1:240 final dilution) and CaCl_2 (16 mM final conc.). The development of thrombin activity was measured as described later in this section.

Human brain thromboplastin was prepared as described in ref. 14. It was subsequently centrifuged at 2000 g for 15 min and stored in 0.1 ml aliquots at -20°C . It was thawed and incubated at 37°C for one hour, then diluted as indicated with buffer A containing 100 mM of CaCl_2 and kept at room temperature. In this way it remained stable for at least 4 h.

Reptilase was obtained from Laboratories Stago (Asnières, France), and prepared according to the indications of the manufacturer.

Soybean trypsin inhibitor (batch no 43 F-800) was obtained from Sigma (St. Louis, USA). A 10 mg/ml solution in buffer A was used.

Staphylocoagulase was prepared as described in Hendrix et al. (15). Chromogenic substrate for thrombin was H.D.-phe-pip-arg-pNA. 2 HCL (S2238, from Kabi, Sweden).

Other chemicals were the highest grade commercially available.

Buffers

A: 0.05 M Tris-HCl, 0.1 M NaCl pH 7.5, with 0.5% egg albumin (Sigma)

B: Buffer A with 20 mM EDTA.

Inhibitor-Deficient Plasmas

Plasma without $\alpha_2\text{M}$ was obtained by addition of anti- $\alpha_2\text{M}$ globulin to normal plasma and elimination of the immunoprecipitate. The γ -globulins were isolated according to Steinbuch et al. (16) from the serum of rabbits immunized against purified human $\alpha_2\text{M}$ (gift from J. Tapon-Bretonnière). No $\alpha_2\text{M}$ could be demonstrated by an immunoelectrophoretic method that would detect $\alpha_2\text{M}$ at concentrations $>2\%$ of the normal plasma concentration. Plasma without AT III was prepared by gently stirring 2 ml of normal human plasma with 0.8 ml of a slurry of sepharose to which anti AT III was bound (gift from Dr. Tran Huu Tri, Basel) for 18 h at 4°C . The antithrombin activity in the plasma after adsorption was not influenced by the addition of 1 U/ml of heparin.

Determination of the Time Course of Amidolytic Activity

An aliquot of plasma is defibrinated as indicated above. To 200 μl of defibrinated plasma is added 50 μl of buffer A, that may contain the substances of which the influence on thrombin generation has to be investigated (e. g. heparins).

At zero time, thrombin generation is triggered by the addition of 50 μl buffer A containing 100 mM of CaCl_2 and a suitable trigger of coagulation. In the experiments described here human brain thromboplastin, diluted as indicated, was used as a trigger. If necessary the volumes indicated may be proportionally altered. At fixed intervals (10, 15, 20, 30 or 60 sec) after zero time, a 10 μl aliquot of the mixture is added to a disposable plastic microcuvette containing 465 μl of buffer B and 25 μl of a 4 mM solution of S 2238 at 37°C .

The aliquots are handled with a Microman pipette (Gilson, France). It is essential to keep the plunger of the pipette down after subsampling and to wipe the pipette tip carefully in order to avoid contamination of the incubation mixtures with EDTA from the buffer in the cuvette. Plastic tubes can be used instead of cuvettes but glass tubes adsorb thrombin and give unpredictable results.

A 15 sec rhythm is easily managed, 10 sec intervals require some practice. If shorter intervals are required one can make use of pipettes equipped with a pushbutton connected to an input gate of a personal computer (Apple IIe) programmed to record the moment of sampling. After sampling the cuvette is quickly and rigorously mixed on a Vortex mixer. After 120 sec the reaction in the cuvette is stopped by adding 0.3 ml of concentrated (98%) acetic acid, the cuvette is again thoroughly mixed and can be kept at room temperature until reading. If a pushbutton pipette and automatic time recording is used for sampling, then stopping the reaction is also carried out with a time recording pipette.

The cuvettes are read at 405 nm in a suitable spectrophotometer (LKB-Ultrospec). The generation of p-Nitroaniline was linear in time up to an O.D. of 0.900. If necessary, incubation times longer or shorter than two minutes can be used so as to obtain a suitable sensitivity and not to exceed 0.9 O.D.

The thrombin concentration data are processed by a computer program to obtain the time course of prothrombin conversion velocity. A full description of this procedure is to be found in the annexe.

Determination of the Thrombin Breakdown Constant

The same mixture as for thrombin generation curves is used, but with a volume of 44 μl of buffer A instead of 50 μl . After 2 to 8 min, at a moment that sufficient thrombin is formed, 6 μl of SBTI solution (10 mg/ml) is added to the main incubation mixture, so as to stop further prothrombinase activity. SBTI at this concentration could be shown to stop completely the activity of prothrombinase and not to influence the amidolytic activity of thrombin (17).

At the shortest possible intervals after addition of the SBTI solution, 10 μl aliquots of the incubation mixture were added to cuvettes by the use of a time recording pipette. The cuvettes were further treated as described for the thrombin generation curves.

Activation by Staphylocoagulase

A solution of staphylocoagulase of 50 nM is used. Of this solution 100 μl was added to 100 μl of a suitably diluted plasma sample. Normal plasma can be diluted 1:50. The incubation mixture remaining after a thrombin generation experiment is best diluted 1:5 or 1:10. The thrombin activity resulting from the formation of coagulase-thrombin is measured by adding 10 μl of the mixture to a cuvette containing buffer B and chromogenic substrate as used for the thrombin generation curves. The staphylocoagulase-prothrombin complex has the same molar activity towards small molecular weight substrates as thrombin (15, 18). Commercially available staphylocoagulase preparations can be used. In that case the own amidolytic activity of the preparation should be determined and its activating capacity has to be checked on normal pooled plasma.

Results

Experimental

The method that we propose here has the purpose to determine the velocity of prothrombin conversion in whole plasma. To this end we determine the amidolytic activity that generates and disappears in plasma after suitable triggering of the coagulation process, using a substrate specific for thrombin. Then we analyse the time-activity curve by a mathematical procedure that separates the process of thrombin formation (prothrombinase activity) and thrombin inactivation (antithrombin activity). In order to carry out the calculations it is necessary to know the

Table 1 Activities and concentrations of $\alpha_2\text{M}$ -thrombin and prothrombin in serum

Thromboplastin dilution	n	Activities (nM thrombin eq)		Concentrations (nM)	
		$\alpha_2\text{M-T}$ (1)	$\alpha_2\text{M-T} + \text{SC}$	$\alpha_2\text{M-T}$ (2)	F II
1:40	10	117 (± 2)	265 (± 5)	65 (± 6)	148 (± 5)
1:100	4	123	280	69	157
1:200	4	109	273	61	162
1:400	10	111 (± 5)	266 (± 8)	62 (± 7)	155 (± 9)

The activities were determined in the reaction mixtures of the experiments shown in Fig. 1, between $t = 10$ and $t = 20$. They are expressed as the equivalent amount of thrombin that would give the same rate of conversion of chromogenic substrate. $\alpha_2\text{M-T}$ (1) is the spontaneous activity, $\alpha_2\text{M-T} + \text{SC}$ is the total activity that could be elicited by addition of an excess of staphylocoagulase. $\alpha_2\text{M-T}$ (2) is the concentration of the $\alpha_2\text{M}$ -thrombin complex obtained by multiplying the $\alpha_2\text{M-T}$ activity with the relevant constant (f, see text). FII is the prothrombin concentration obtained by subtraction of the first two columns. SEM are indicated when $n = 10$

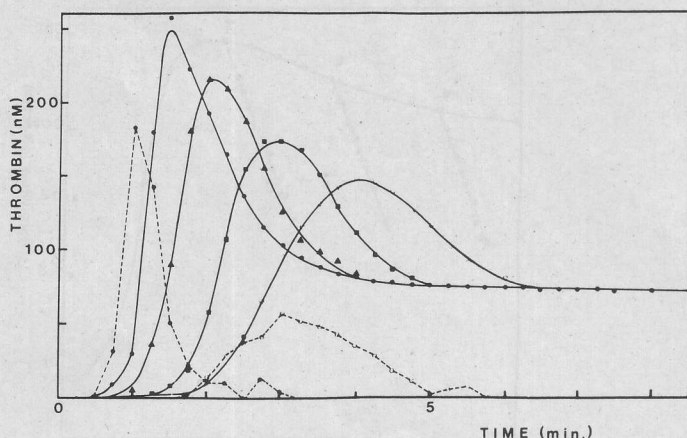


Fig. 1 Thrombin generation and the rate of thrombin formation in normal plasma triggered with different dilutions of thromboplastin. The reaction mixture consisted of 240 μ l defibrinated plasma 60 μ l buffer and 60 μ l thromboplastin diluted as follows 1/40 (\bullet); 1/100 (\blacktriangle); 1/200 (\blacksquare); 1/400 (\circ). The dotted lines represent the thrombin generation velocities of the 1/40 and 1/400 curves resp. The scale for these curves is the thrombin generation velocity in $\text{nM} \cdot \text{min}^{-1}$

reaction constants of the breakdown processes. In the first part of this section we will discuss how these data are obtained. In the second part we verify the mathematical model.

I. The Thrombin Generation Curve and the Reaction Constants

Fig. 1 shows the thrombin activities generated in normal plasma by different dilutions of tissue thromboplastin. As a rule the standard derivation of each point is below 4% except for the points located in the steep rising parts of the generation curve. With the aid of pipettes provided with a signalling pushbutton connected to an input port of an Apple IIe computer we could estimate the experimental error in the sampling time to be between 0.4 and 1 sec. In the horizontal parts of the curve the coefficient of variation of the thrombin determination is 2.75%.

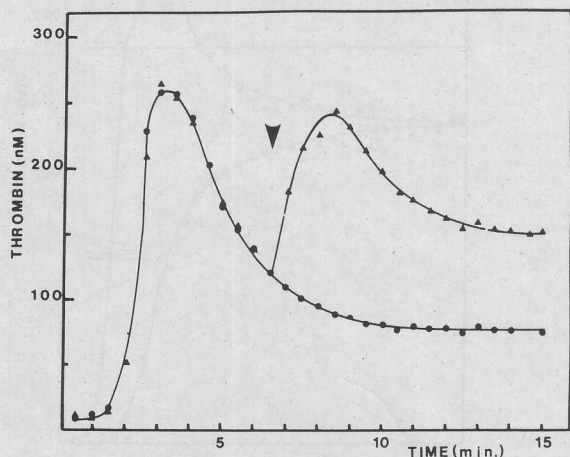


Fig. 2 Effect of addition of prothrombin during thrombin generation. To a thrombin generation experiment as in Fig. 1 (thromboplastin dilution 1/40) at 7 min (arrow) 1500 nM of purified human prothrombin was added

When the tangent of the curve is steep, the time error adds significantly to the overall error so as to make it 15–18% at tangents exceeding $1000 \text{ O.D.U.} \cdot \text{min}^{-1}$. With the aid of the time recording pipettes it is possible to eliminate the time error and to obtain almost perfectly overlapping curves in repeated experiments.

The amounts of residual prothrombin left in the mixtures after the amidolytic activity has reached a steady end-level, were determined with staphylocoagulase (Table 1). They are apparently not dependent upon the variations in velocity of thrombin generation induced by dilution of thromboplastin.

From Fig. 2 it appears that thrombin generation can be resumed in the course of the experiment if purified human prothrombin is again added. The method thus determines thrombin generation velocities that are dependent upon both prothrombin concentration and prothrombinase activity.

Fig. 3 shows experiments in which the breakdown constant of thrombin in plasma was determined. The decay curve of thrombin generated endogenously in plasma was obtained by stopping the factor X_3 activity in thrombin generating plasma by addition of SBTI and measuring the thrombin disappearance (see methods section). Alternatively 200 nM of thrombin that had been formed in a euglobulin mixture was added at zero time to defibrinated plasma diluted in the same way as the reaction mixture, SBTI included. In the latter case the decay was two fold quicker. The slower decay of endogenously formed thrombin cannot be attributed to ongoing thrombin formation because the amount of prothrombin that can be demonstrated with staphylocoagulase in the plasma does not change after the moment of stopping the reaction with SBTI (results not shown). During the course of thrombin generation the decay constant of thrombin does not change (Fig. 4), showing that in the course of the experiment there is not a time-dependent change in the properties of thrombin that could explain the difference in half-life time. For the moment we have no explanation for this phenomenon (see discussion).

The form of the decay curve is adequately described by a logarithmic approach to a steady end level (Figs. 3 and 4). From experiments like these overall decay constants can be obtained. This overall constant (k_{exp}) is equal to the sum of the $\alpha_2\text{M}$ -

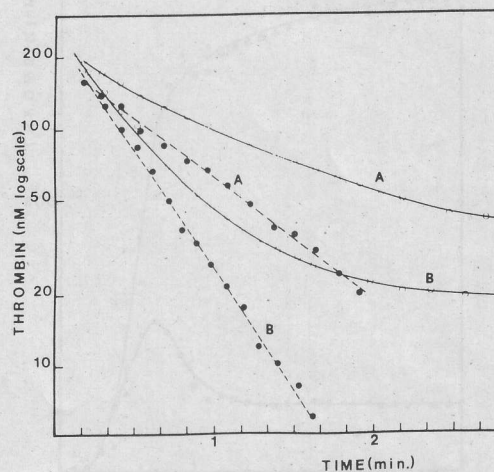


Fig. 3 Thrombin inactivation in plasma. A. (\circ - \circ) Decay of endogenous amidolytic activity after addition of SBTI in a standard thrombin generation experiment, 3 min after triggering the reaction with thromboplastin 1/40. (\bullet - \bullet) The same data after subtraction of the steady end-level of activity. B. (\circ - \circ) Decay of amidolytic activity of 200 nM exogenous thrombin. (\bullet - \bullet) The same data after subtraction of the steady end-level of activity

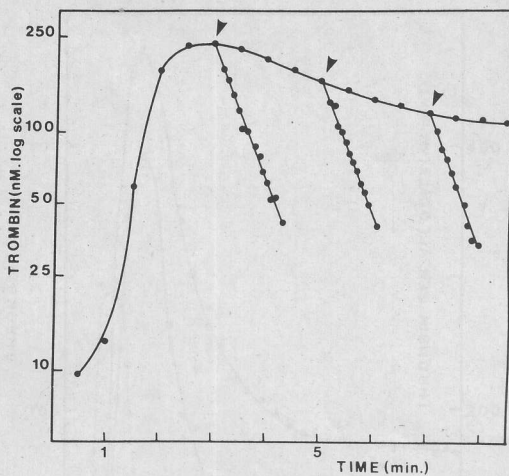


Fig. 4 Decay of endogenous thrombin at different intervals after the start of thrombin generation. In thrombin generation experiments triggered with thromboplastin 1/40, SBTI was added at $t = 3, 5$ and 7 min. The decay constants observed were 1.24, 1.17 and 1.23 respectively. The data are plotted on a log scale after subtraction of the steady end-levels

dependent decay constant (k_1) and the non- α_2 M-dependent constant (k_2), mainly due to antithrombin III. Table 2 lists these constants for various kinds of normal and deficient plasma. Also, in this table the ratio of the amount of prothrombin consumed during the experiment (i. e. the total amount of thrombin formed) and the amount eventually taken up by α_2 M is given, as well as the values of k_1 and k_2 that can be calculated from these data. For this calculation it is necessary to know the value of f ; i. e. the ratio of the amidolytic activity of one mole of the α_2 M-thrombin complex to one mole of free thrombin. This value was obtained by preparing thrombin in a euglobulin precipitate of defibrinated plasma and estimating its activity. Then an excess of an α_2 macro-

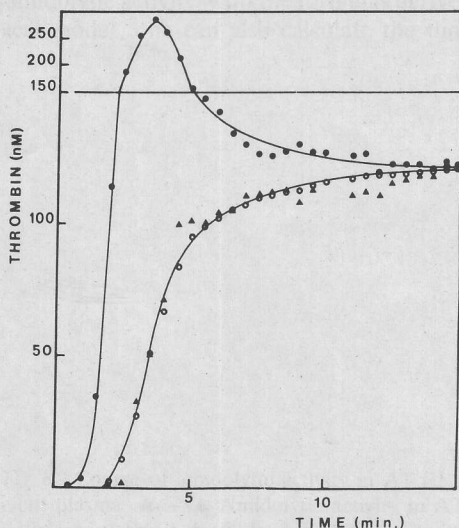


Fig. 5 Comparison of observed and computed levels of the α_2 M-thrombin-complex. A standard thrombin generation experiment was carried out. Thromboplastin diluted 1:40 was used as a trigger. From the amidolytic activity (●) the concentration of α_2 M-T complex was computed (○). The levels of α_2 M-T complex found experimentally are represented by ▲

Table 2 Reaction constants of inactivation in protease-inhibitor deficient plasmas

Plasma	Dil.	k_{exp} (min^{-1})	R_{exp}	k_1 (min^{-1})	k_2 (min^{-1})
Control	1.00	1.260	0.271	0.342	0.920
α_2 M def.	0.78	0.733	—	—	0.733
Control		0.856	0.236	0.200	0.756
AT III def.	0.45	0.249	—	0.167 ¹⁾	0.082 ²⁾
Control		0.630	0.270	0.170	0.460
Double def.	0.32	0.058	—	—	0.058
Control		0.462	0.240	0.111	0.352

Dil.: Dilution due to immunoadsorption, the control plasmas were diluted to the same extent

k_{exp} : Overall decay constant observed (min^{-1})

R_{exp} : Ratio of α_2 M-T to At III-T in the control plasma

k_1 and k_2 : Pseudo first order reaction constants of the complexation of thrombin with α_2 M and non- α_2 M respectively (min^{-1})

¹⁾ Obtained from the double deficient plasma (accounting for the difference in dilution)

²⁾ Obtained by subtracting k_1 from k_{exp}

globulin preparation devoid of AT III activity was added and the residual amidolytic activity was determined, f was estimated to be 0.558 ± 0.043 ($n = 10$; S.E.M.) (cf. Table 1).

II. Verification of the Model

The adequacy of our model was tested in two fundamentally different ways a) by comparing the time course of reaction products as predicted by the model with those actually found, b) by studying the effect of selective elimination of AT III and/or α_2 M.

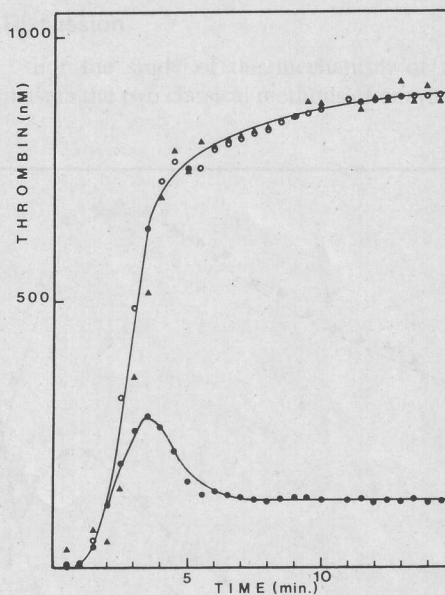


Fig. 6 Comparison of observed and computed levels of consumed prothrombin. In a standard thrombin generation experiment (trigger: thromboplastin 1:40) the amount of prothrombin converted (○) was computed from the amidolytic activities (●) and compared to the difference between the original amount of prothrombin and amount remaining during the time course of activation (▲)

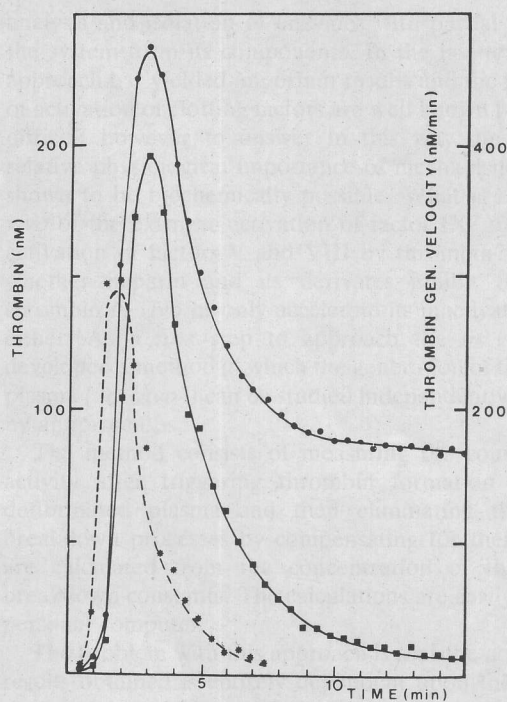


Fig. 7 Course of amidolytic activity in α_2 -macroglobulin deficient plasma. \blacksquare — \blacksquare Amidolytic activity in α_2 M deficient plasma. \bullet — \bullet Amidolytic activity in control plasma. Dotted line: Thrombin generation velocity calculated from the amidolytic activity curves. \circ : points obtained from the α_2 M deficient plasma. \star : points obtained from the control plasma. The constants used for calculation are given in Table 1

In Fig. 5 two sets of experimental data are plotted. The overall amidolytic activity as determined by our standard procedure (\bullet) and the level of α_2 M-thrombin (\blacktriangle). The latter was determined by subsampling from the reaction mixture into a solution containing an excess (1 U/ml) of heparin and 20 mM EDTA. In this mixture the generation of thrombin stops immediately and, because the half-life of thrombin is <10 sec, the residual activity measured after 10–20 min is due to the amount of α_2 M-bound thrombin that was present at the moment of subsampling. From the data on amidolytic activity, with the formulas derived from the mathematical model, one can also calculate the time course of the α_2 M-

thrombin concentration (\circ). From Fig. 5 it is seen that the calculated values and the values actually determined coincide within the limits of experimental error. Fig. 6 also shows the equivalence of a set of experimental and calculated concentrations, viz. those of the total amounts of prothrombin converted into thrombin. Experimentally these concentrations were determined by subsampling into a mixture containing 20 mM EDTA, 1 U/ml of heparin and $2\mu\text{M}$ of staphylocoagulase. In this mixture prothrombin conversion into thrombin stops, the thrombin present is inactivated by AT III and the prothrombin present converts quickly (second order rate constant $3.3 \times 10^6 \text{M}^{-1}\text{s}^{-1}$; [12]) into the amidolytically active prothrombin-staphylocoagulase complex that is not inactivated by AT III. Thus one obtains the amount of residual prothrombin (after subtraction of the activity due to α_2 M-thrombin determined independently) while the amount of prothrombin converted into thrombin is found by subtraction from the starting level. The equivalence between the concentrations found experimentally and those derived from the amidolytic activity via the kinetic model is again satisfactory.

A different approach to test the validity of the model is shown in Figs. 7 and 8. Here the inactivation system of the plasma was modified by selective immunoadsorption of α_2 -macroglobulin and AT III respectively. Control plasmas were obtained by treating normal plasma in a comparable way with a gammaglobulin fraction from serum from non-immunised rabbits and adjustment of the dilution if necessary.

In Table 2 it is shown that the decay constants calculated for the non-deficient plasma are identical to those found experimentally in the deficient plasmas, if one allows for a small amount of inhibition that is due to neither α_2 M or AT III.

In Fig. 7 the thrombin activity curves obtained with the α_2 M deficient plasma and its control are shown as well as the thrombin generation velocity curves obtained from them, using the relevant constants from Table 2. In Fig. 8 the same is done for the AT III deficient plasma. In both Fig. 7 and Fig. 8 the thrombin generation velocity curves obtained with the deficient and the control plasmas superimpose within the limits of experimental error, as would be expected from the model.

Discussion

For the study of the mechanism of thrombin formation in plasma the two classical methods of enzymology are open, kinetic

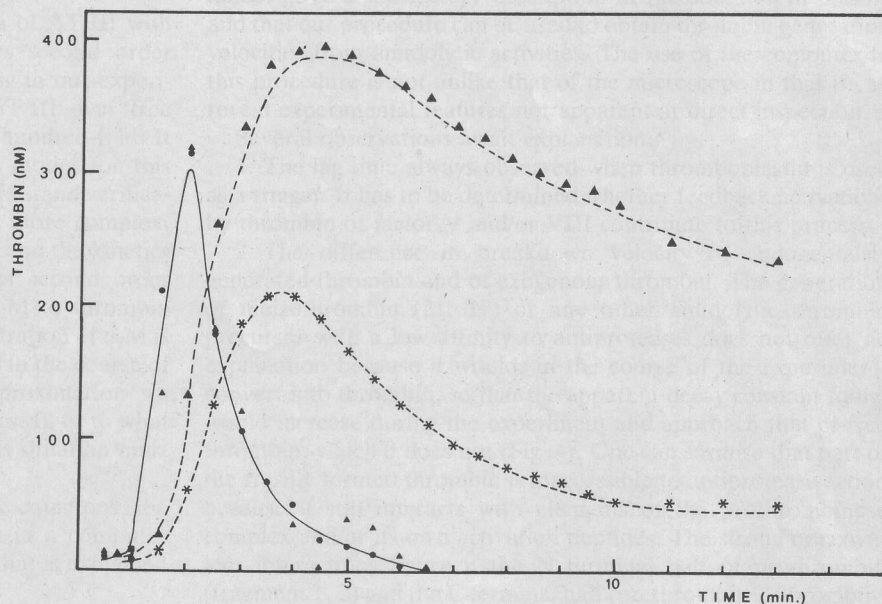


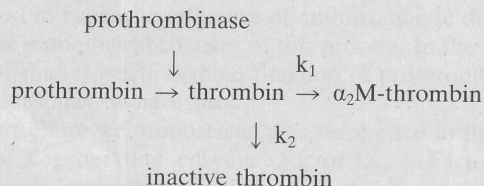
Fig. 8 Course of amidolytic activity in AT III deficient plasma. \blacktriangle — \blacktriangle Amidolytic activity in AT III deficient plasma. \star — \star Amidolytic activity in control plasma. Drawn line: Thrombin generation velocity calculated from the amidolytic activity curves. \blacktriangle : points obtained from the AT III deficient plasma. \bullet : points obtained from the control plasma. The constants used for calculation are given in Table 2

analysis and isolation of enzymes with partial reconstruction of the system from its components. In the last ten years the latter approach has yielded important results and the main mechanisms of activation of clotting factors are well known by now. It remains difficult however to answer in this way questions about the relative physiological importance of mechanisms that have been shown to be biochemically possible. What is the importance *in vivo* of the extrinsic activation of factor IX? or of the feedback activation of factors V and VIII by thrombin? The question of whether heparin and its derivatives inhibit the formation of thrombin *in vivo* or only accelerate its inactivation is not settled either. As a first step to approach the *in vivo* situation we developed a method in which the generation of thrombin in whole plasma ("*ex vivo*") can be studied independently of its breakdown by antiproteases.

The method consists of measuring the course of amidolytic activity after triggering thrombin formation in a sample of defibrinated plasma and then eliminating the effect of the breakdown processes by compensating for their velocities, that are calculated from the concentration of thrombin and the breakdown constants. The calculations are easily carried out on a personal computer.

The problem with this approach is that the acceptability of the results obtained is entirely dependent upon the acceptability of the model and the mathematical procedure. Therefore it is important to test extensively the validity of the method.

We assume that the situation in the plasma when thrombin generates can be depicted by the following diagram:



The assumptions at the basis of the kinetic model are:

1. Thrombin breakdown occurs via two processes, one (caused by AT III and others antiproteases) leading to inactive products, the other ($\alpha_2\text{M}$ -dependent) leading to a product with a residual amidolytic activity.
2. Both reactions are of pseudo first order with regard to thrombin.

It has been demonstrated that the interaction of AT III with thrombin is a bimolecular reaction and shows second order kinetics (19, 20). Under the conditions prevailing in our experiments there is at least a ten-fold excess of AT III over free thrombin and the excess of α_1 -antithrombin is a hundred-fold. It therefore seemed reasonable to try whether a model for this reaction based on pseudo first order kinetics would stand verification. The case of $\alpha_2\text{M}$ -dependent inactivation is more complex. Here the stoichiometry of the reaction is variable and the kinetics of the reaction are complicated (20, 6-10), yet second order reaction kinetics describe the situation if the $\alpha_2\text{M}$ to thrombin ratio is higher than 2 to 1 (20). The initial concentration of $\alpha_2\text{M}$ is 2 μM . No more than 300 nM of this is complexed in the course of the experiment (Figs. 1 and 5). As a first approximation we therefore again assumed pseudo first order kinetics. If, or to what extent it is indeed an adequate description of this situation must follow from the experiments.

The translation of the reaction model into rate equations, the solution of these equations and their utilization in a computer program is a technical problem in mathematics, that is explained in the annexe and will not be discussed here.

The following qualitative arguments support the correctness of the model:

a. The partition between the AT III- II_a complex and the $\alpha_2\text{M}$ - II_a complex of the thrombin generated in an experiment is independent of the course of thrombin generation. This can be seen in Fig. 1 and Table 1, where the total amount of prothrombin as well as the levels of $\alpha_2\text{M}$ - II_a eventually obtained are independent of the dilution of thromboplastin used. Our model predicts such behaviour (formula 8).

b. The decay of thrombin activity is characterized by a logarithmic approach to a steady end-level (Fig. 3). This proves the pseudo first order character of the breakdown constants (formula 4c).

c. The breakdown constant of thrombin does not change over the experiment (Fig. 4).

d. The level of activity remaining in the plasma after activation is explained by the level of $\alpha_2\text{M}$ - II_a present (Fig. 5), in the absence of $\alpha_2\text{M}$, the activity approaches the zero level (Fig. 7).

In the absence of both $\alpha_2\text{M}$ and AT III there is a residual antithrombin activity of about 18% of the total (Table 2). This activity is not heparin dependent and consumes thrombin in a pseudo first order process (results not shown). In our formulas this decay is included in the AT III dependent decay (k_2). This inhibition activity is presumably due to α_1 antitrypsin and other antiproteases.

From the observations (a to d) it can be concluded that the qualitative features of our model are supported by the experiments. The validity of our model is further corroborated by the similarity of the concentration courses calculated and those actually observed. In principle the time courses of all the reactants can be computed from the time course of the amidolytic activity. In practise the amounts of remaining prothrombin and of the $\alpha_2\text{M}$ -thrombin complex can be determined independent of amidolytic activity, thus offering the opportunity to compare the results of the calculation with the time-course of concentrations actually found. Figs. 5 and 6 shows that the observed and the computed points seem to follow the same curve.

Further qualitative arguments for the correctness of our procedure can be derived from Figs. 7 and 8. Here it is seen that the velocities of thrombin generation computed from plasma specifically deficient in either $\alpha_2\text{M}$ or in AT III coincide with those of the control plasma if the respective breakdown constants are eliminated from the computation.

We conclude that, within the limits of experimental error our model gives a satisfactory description of the situation in plasma and that our procedure can be used to obtain thrombin generation velocities from amidolytic activities. The use of the computer in this procedure is not unlike that of the microscope in that it can reveal experimental features not apparent at direct inspection.

Several observations await explanation:

1. The lag time always observed when thromboplastin is used as a trigger. It has to be determined whether feedback activations by thrombin of factor V and/or VIII contribute to this process.
2. The difference in breakdown velocity of endogenously generated thrombin and of exogenous thrombin. The generation of meizothrombin (21, 17) or any other amidolytic thrombin precursor with a low affinity to antiproteases does not offer an explanation because it would, in the course of the experiment, convert into thrombin, so that the apparent decay constant found would increase during the experiment and approach that of free thrombin, which it does not (Fig. 4). One can surmise that part of the freshly formed thrombin is inaccessible to antiprotease action because it still interacts with elements of the prothrombin complex and/or its own activation peptides. The strong noncovalent interactions between the N terminal half of prothrombin (fragment 1, 2) and the C terminal half (prethrombin 2, thrombin)

(22) make such interactions not *a priori* unlikely. This problem is at present under investigation. The same phenomenon probably explains why the pseudo order rate constants that we find for endogenous thrombin breakdown in whole plasma are below the lower limits of ranges found in the literature with free thrombin and in (partly) purified systems. For α_2M we found 0.342 min^{-1} (Table 1) compared to $0.4\text{--}0.8 \text{ min}^{-1}$ in the literature. For AT III the value was 0.920 min^{-1} compared to $1.4\text{--}3.6 \text{ min}^{-1}$ (6–10, 23). The value of 0.09 for the decay constant of thrombin in the presence of the normal plasma concentration of α_2M that can be calculated from ref. 20, has been supposed by others to be based on an underestimation of the second order rate constant of this inhibition (10).

3. The model behaves as if the inhibitory capacity of α_2M is immediately available, although it is known that this molecule has to be activated by a proteolytic enzyme, e. g. by the proteolytic enzyme that it will complex with (5, 6 and references therein).

4. Even though prothrombinase is not exhausted (Fig. 2), the reaction comes to a stop when 5–10% of the original prothrombin is left. This suggests the possibility of product inhibition of prothrombin activation in whole plasma. Our results indicate that in 2:3 diluted normal platelet free plasma (our standard system) the pseudo first order rate constant of α_2M , AT III and non- α_2M -non-ATIII antiproteases contribute 23%, 64% and 13% to the overall disappearance rate of endogenous thrombin. This is in accordance with the 20–30% of plasma antithrombin activity assigned to α_2M by several authors (10, 19, 20).

We think that our method may be a valuable means to investigate the physiology of thrombin generation in whole plasma and to study the influence of antithrombotic drugs on, as well as the pathological changes of this process. In the latter case additional studies, such as the estimation of prothrombin activation peptides may be necessary.

The procedure we propose can also be applied to the study of the factor X generating enzyme (Factor IX_a – Factor VIII_a – Phospholipid) in plasma on basis of the factor X_a levels observed.

Acknowledgements

This work has partly been financed by grants from the “Nederlandse Thrombosesichting” and the “Fondation de France”. Partly it was carried out during a sabbatical leave of one of the authors (H.C.H.) at the Dept. of Hematology CHU Necker-Enfants Malades (Paris). We thank Mme. J. Tapon-Bretonnière for a gift of purified α_2 -Macroglobulin and Dr. Tran Huu Tri for the sepharose-bound anti AT-III.

References

- 1 Wagner R H, Graham J B, Penick G D, Brinkhous K M. Estimation of prothrombin by the two stage method. In: Blood coagulation, Hemorrhage and Thrombosis. Tocantins M L, Kazal L A (eds) Academic Press, New York – London 1965.
- 2 Biggs R, Macfarlane R G. Human blood coagulation and its disorders. Blackwell Scientific Publications, Oxford 1967.
- 3 Hemker H C, Hemker P W, v d Torren K, Devilee P P, Hermens W Th, Loeliger E A. The evaluation of the two-stage prothrombin assay. Thromb Diathes Haemorrh 1971; 25: 545–54.
- 4 Pekelharing C A. Over de betrekking van het fibrineferment van het bloedsersum tot de nucleoproteide van het bloedplasma. Versl Kon Acad Wetensch Amsterdam 1895; 3: 272–97.
- 5 Travis J, Salvesen G S. Human plasma proteinase inhibitors. Ann Rev Biochem 1983; 52: 655–709.
- 6 Rosenberg R D, Harpel P C. α_2 -Macroglobulin and antithrombin-heparin cofactor: modulators of hemostatic and inflammatory reactions. Prog Hemost Thromb 1976; 3: 145–98.
- 7 Barret A J, Starkey P M. The interaction of α_2 macroglobulin with proteinase. Biochem J 1973; 133: 709–15.

- 8 Harpel P C. Studies on human plasma α_2 -macroglobulin enzyme interactions. Evidence for proteolytic modification of the subunit chain structure. J Exp Med 1973; 138: 508–21.
- 9 Rinderknecht H, Feling R M, Geokas M C. Effect of α_2 -macroglobulin in some kinetic parameters of trypsin. Biochim Biophys Acta 1975; 377: 150–65.
- 10 Fischer A M, Tapon-Bretonnière J, Bros A, Josso F. Respective roles of antithrombin III and α_2 -macroglobulin in thrombin inactivation. Thromb Haemostas 1981; 45: 51–4.
- 11 Josso F, Prou-Wartelle O. Exploration de l'hémostase. In: Techniques en hématologie. Alagille D et al. (eds). pp. 101–208. Flammarion, Paris 1972.
- 12 van Dam-Mieras M C E, Muller A D, van Diejen G, Hemker H C. Methods of enzymatic analysis. Enzymes 3: Peptidases, proteinase and their inhibitors. Verlag Chemie, Weinheim 1984; 5: 352–94.
- 13 Ødegard O R, Lie M. On the use of chromogenic substrates for the study of coagulation inhibitors. Haemostasis 1978; 7: 121–6.
- 14 Owren P A, Aas K. The control of dicumarol therapy and the quantitative determination of prothrombin and proconvertin. Scand J Clin Lab Invest 1951; 3: 201–18.
- 15 Hendrix H, Lindhout T, Mertens K, Engels W, Hemker H C. Activation of human prothrombin by stoichiometric levels of staphylocoagulase. J Biol Chem 1983; 258: 3637–44.
- 16 Steinbuch M, Audran R. Isolement de l'immunoglobuline IgG du plasma humain à l'aide de l'acide caprylique. Rev Franç Etud clin biol 1969; 14: 1054–8.
- 17 Lindhout T, Baruch D, Schoen P, Franssen J, Hemker H C. Thrombin generation and inactivation of antithrombin III and heparin. (Submitted for publication).
- 18 Kawabata S, Morita T, Iwanaga S, Igarashi H. Staphylocoagulase-binding region in human prothrombin. J Biochem 1985; 97: 329–31.
- 19 Rosenberg R D, Damus P S. The purification and mechanism action of human antithrombin-heparin cofactor. J Biol Chem 1973; 248: 6490–505.
- 20 Downing M R, Bloom J W B, Mann K G. Comparison of the inhibition of thrombin by three plasma protease inhibitors. Biochemistry 1978; 17: 2649–53.
- 21 Rosing J, Zwaal R F A, Tans G. Formation of meizothrombin as an intermediate in Factor X_a-catalyzed prothrombin activation. J Biol Chem 1986; (In press).
- 22 Esmon C T, Jackson C M. The conversion of prothrombin to thrombin. IV. The function of the fragment 2 region during activation in the presence of factor V. J Biol Chem 1974; 249: 7791–7.
- 23 Jesty J. The kinetics of inhibition of thrombin by antithrombin in the presence of components of the hemostatic systems. Blood 1985; 66: 1189–95.
- 24 Hemker H C, Willems G, Béguin S. Thrombinoscope, a personal computer program for the analysis of thrombin generation curves. Computers in Biol and Med. (Submitted).

Received February 24, 1986 Resubmitted May 1, 1986

Accepted May 2, 1986

Annexe

Mathematical Analysis of the Thrombin Generation Curve

(G. M. Willems, H. C. Hemker).

The amidolytic activity that we observe in the experiment is the sum of the activities of free thrombin and α_2M -bound thrombin. The experimental procedure gives this activity at a series of points in time. It is our purpose to obtain the velocity of thrombin generation alone as a function of time $[g(t)]$, independent of the breakdown processes.

We can make no assumptions as to the form of $g(t)$ because the enzyme that causes prothrombin activation (prothrombinase) itself increases and decreases as an unknown function of time. We describe the disappearance of thrombin as the sum of two pseudo first order reactions, one giving the inactive product of the

interaction of antithrombin III and thrombin (AT III-T) and minor amounts of other inactive forms of thrombin and the other the amidolytically active complex of α_2 -macroglobulin and thrombin (α_2 M-T). (see discussion for experimental justification).

The chemical model can be represented by a set of differential equations that describes the relations between the reaction velocities and the concentrations of the reactants in the system. We will use the following symbols:

- A(t) = amidolytic activity found at time t
P(t) = prothrombin concentration at time t
T(t) = thrombin concentration at time t
M(t) = α_2 M-thrombin concentration at time t
g(t) = thrombin generation from prothrombin as a function of time
 k_1 = pseudo first order rate constant for the binding of thrombin to α_2 M
 k_2 = pseudo first order rate constant for the binding of thrombin to AT III
f = the ratio of the enzymatic activity of one mole of α_2 M-T over that of one mole of thrombin

The subscripts *thr* and *mcr* will indicate the activities due to thrombin and to α_2 M-thrombin respectively, the total activity is unsubscripted.

The amidolytic activity will be expressed in thrombin equivalent-amounts, so that $A = T$ in a solution containing only thrombin and $A = f.M$ in the presence of α_2 M-thrombin only.

At any moment the net velocity of thrombin formation will be equal to the sum of the velocity of thrombin generation from prothrombin and the velocity of thrombin breakdown:

$$\frac{d}{dt} T(t) = g(t) - k_1 T(t) - k_2 T(t) \quad (T[0] = 0) \quad (1a)$$

The velocity of the formation of α_2 M-thrombin is proportional to the concentration of free thrombin,

$$\frac{d}{dt} M(t) = k_1 T(t) \quad (M[0] = 0) \quad (1b)$$

and the activity measured is the sum of the activities of thrombin and α_2 M-thrombin:

$$A(t) = T(t) + fM(t) \quad (1c)$$

These equations have the solution

$$g(t) = \frac{d}{dt} \left\{ A(t) + [(1-f)k_1 + k_2] \int_0^t e^{-fk_1(t-\tau)} A(\tau) d\tau \right\} \quad (2)$$

In order to use this formula we need estimates of k_1 , k_2 and f separately. The value f can be obtained by comparing the amidolytic activity of thrombin with that of α_2 M-thrombin (see experimental section). If, at any moment s during thrombin generation we stop the conversion of prothrombin (e. g. by the addition of SBTI) then the course of the remaining activity is given by:

$$\frac{d}{dt} T(t) = -(k_1 + k_2)T(t); T(s) = T_s \quad (3a)$$

$$\frac{d}{dt} M(t) = k_1 T(t); M(s) = M_s \quad (3b)$$

$$A(t) = T(t) + fM(t) \quad (3c)$$

where t is counted from moment s on and M_s is the amount of α_2 M-thrombin present at moment s .

From the formulas 3 it can be derived that:

$$A(t) = T_s e^{-(k_1 + k_2)(t-s)} + f \left\{ \frac{k_1}{k_1 + k_2} T(s) \left[1 - e^{-(k_1 + k_2)(t-s)} \right] + M(s) \right\} \quad (4a)$$

for $t \rightarrow \infty$ this gives

$$A(\infty) = f \left\{ \frac{k_1}{k_1 + k_2} T(s) + M(s) \right\} \quad (4b)$$

so that

$$\log[A(t) - A(\infty)] = \log \left\{ \frac{(1-f)k_1 + k_2}{k_1 + k_2} \right\} T_s - (k_1 + k_2)(t-s) \quad (4c)$$

from which it follows that $k_1 + k_2$ can be obtained from the slope of the plot of $\log [A(t) - A(\infty)]$ against time. The activity found at some late point(s) in the time curve are taken as $A(\infty)$. In order to obtain k_1 and k_2 separately we integrate (2):

$$P(0) - P(\infty) = \int_0^\infty g(\tau) d(\tau) d\tau = A(\infty) + \frac{(1-f)k_1 + k_2}{f k_1} A(\infty) \quad (5)$$

from which it follows that:

$$A(\infty) = \frac{f k_1}{k_1 + k_2} \int_0^\infty g(\tau) d\tau = \frac{f k_1}{k_1 + k_2} [P(0) - P(\infty)] \quad (6)$$

where $P(\infty)$ is the amount of prothrombin present after the activation is over, so that $P(0) - P(\infty)$ is the total amount of thrombin that has been formed during the experiment from formula (6), and equal to the amount of prothrombin converted during the experiment. We calculate k_1 as

$$k_1 = \frac{(k_1 + k_2)A(\infty)}{f [P(0) - P(\infty)]} \quad (7)$$

and k_2 from:

$$k_2 = (k_1 + k_2) - k_1 \quad (8)$$

Once the constants f , k_1 and k_2 are known we can use the following rearrangement of formula 2 to calculate $g(t)$:

$$\int_0^t g(\tau) d\tau = A(t) + \frac{(k_1 + k_2) - f \cdot k_1}{k_1 + k_2} e^{-fk_1 t} \int_0^t A(\tau) e^{+fk_1 \tau} d\tau \quad (9)$$

In practice $A(n)$ is obtained at a series of times, $t(n)$; n being the running number of the observation. The integral in the right hand term of formula (9) $[I(n)]$ can be obtained as the area under the curve generated by plotting

$$S(n) = A(n) e^{fk_1 t(n)} \quad (10)$$

against $t(n)$, so that

$$I(n) = \frac{1}{2} \sum_1^n \left[(t(n) - t(n-1)) \cdot (s(n) + s(n-1)) \right] \quad (11)$$

The amount of thrombin that would have been formed at the moment of observation if no inactivation had taken place then is obtained as:

$$T(n) = A(n) + [(k_1 + k_2 - fk_1)/(k_1 + k_2)] \cdot e^{-fk_1 t(n)} \cdot I(n) \quad (12)$$

The following series of Basic commands, operating on the arrays $A(n)$ and $t(n)$ carries out these calculations.

100 CE = F * K1 : CT = (K1 + K2 - CE)/(K1 + K2)

110 A(0) = 0 : T(0) = 0 : IN = 0 : SN = 0

120 FOR N = 1 TO E : REM E = number of data points

130 S = A(N) * EXP [CE * T (N)]

```

140 I = IN + [T(N) - T(N-1)] * [(S + SN)]/2
150 TT(N) = A(N) + CT * I * EXP [- CE * T(N)]
160 V (N-1) = [TT(N) - TT(N-1)]/[T(N) - T(N-1)]
170 SN = S : IN = I
180 NEXT N

```

It will yield the array TT(N) and V(N) that, when plotted against T(N) (sampling time) give the total amount of prothrombin activated and the velocity of prothrombin conversion as a function of time.

This routine is the core of the program "Thrombinoscope" (24) that a) stores the experimental data on disk b) calculates prothrombinase activity c) stores the results of the calculation on disk and d) displays the data and results in graphic and/or alphanumeric format on the monitor or the printer. The program was developed in Applesoft Basic on an Apple IIe personal computer with 128 k RAM and Applesoft in ROM. Both data obtained at regular and at irregular intervals can be accommodated. It also contains the subprogram "Stopwatch" that is used to record sampling and stopping times from pipettes provided with a pushbutton. The program can be obtained from the authors.

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Adrenergic Receptors: Molecular Properties and Therapeutic Implications

Symposium St.-Paul-de-Vence, France, October 21st-24th, 1984

Symposia Medica Hoechst 19

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ISBN 3-7945-1055-0

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