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Food and fluid related aspects in highly trained athletes
Food and fluid related aspects in highly trained athletes

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To my parents
In memory of Lars Hermansen
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Parts of the present work have already been published.
ABBREVIATIONS

Aceto A  aceto acetate.
ACTH  adrenocorticotropic hormone
AM  morning
AMP  adenosine monophosphate
ATP  adenosine triphosphate
BHBA  beta hydroxybutyric acid
BW  body weight
CCK  cholecystokinin
CHO  carbohydrate
Δ E  difference between energy intake and energy expenditure
EB  energy balance
EE  energy expenditure
EI  energy intake
en%  % of total energy
FFA  free fatty acids
FM  normal diet composed of carbohydrate rich foodstuffs and supplemented with a beverage containing 50% of fructose and 50% of maltodextrin
g  gram
mg  milligram (10⁻³ gram)
ng  nanogram (10⁻⁹ gram)
pg  picogram (10⁻¹² gram)
μg  microgram (10⁻⁶ gram) (mcg)
GE  gastric emptying
GI  gastro intestinal
GIP  gastro intestinal peptide
³H₂O  water labeled with Tritium
HPLC  high pressure liquid chromatography
kJ⁻¹  kilocalorie.
kJ  kilojoule
l  liter
l⁻¹  per liter
Max  maximal
Mf  normal diet composed of carbohydrate rich foodstuffs and supplemented with a beverage containing a high fraction of maltodextrin and a low fraction of fructose
min  minute
MJ  megajoule
ml  milliliter (10⁻³ liter)
mmol  millimole (10⁻³ mol)
nmol  nanomole (10⁻⁹ mol)
µmol  micromole (10⁻⁶ mol)
mosm  milliosmoles
N  normal diet composed of carbohydrate rich foodstuffs
n  number
NADH  reduced nicotine adenine di-nucleotide
NH₃  ammonia
Nu  urinary nitrogen
PM  afternoon
R  respiratory quotient
RDA  recommended daily allowance
RIA  radio immuno assay
SEM  standard error of the mean
t ½  half time
TG  Triacylglycerol
IU  international units
mU  milli units
uv  ultra violet
VCO₂  carbon dioxide production (volume/time)
VIP  vaso intestinal peptide
VO₂  oxygen uptake (volume/time)
vol  volume
W  Watt
Wmax  maximal achieved performance capacity expressed in Watt
INTRODUCTION

Nutrition as one of the factors influencing performance has received attention from athletes and their coaches since the classical athletic competitions in Olympia. However, it is only since the last 50 years that scientific studies have been focussed more specifically on the role of nutrients in human performance and physical fitness.

In 1939 Christensen and Hansen observed that a change in the proportions of carbohydrate (CHO) and fat in the daily diet influenced the respiratory quotient. They concluded that food intake was one of the determinants of substrate selection for oxidation. It was also observed that physical activity raised the value of the respiratory quotient (R) and the conclusion was that CHO metabolism is enhanced as a result of physical activity.

More recently it has been suggested that fat is the dominant substrate for energy exchange in resting conditions and that CHO plays a role as immediate energy source, whenever energy exchange from fat is too slow to meet the requirements (Newsholme, 1976). Based on this suggestion it can be explained that CHO degradation, which leads to a larger energy flow per second than fat degradation (McGilvery, 1973), is enhanced at the onset of exercise when aerobic metabolism still has to be increased. It may also explain that increased fatty acid availability, in the course of exercise caused by neural and hormonal adaptations, reduces CHO oxidation so that endogenous CHO stores can be spared (Newsholme, 1976, Jansson, 1984).

Since the studies of Bergström and Hultman (1967, 1967a) it is known that local CHO stores play a limiting role for the ability to perform exercise of a high intensity. This may explain the adaptation of the body to increase fat metabolism and to spare CHO for emergency actions. The same authors also observed that increasing the amount of CHO in the daily diet results in increased glycogen stores and that this increase in glycogen availability is related to longer exercise times before exhaustion is reached. The general advice thereafter was that athletes should consume more CHO.

With respect to the observations of Christensen and Hansen (1939a, 1939b) this may sound conflicting. Increasing the amount of CHO in the diet stimulates CHO oxidation and inhibits fat metabolism, whereas the natural adaptations of the body lead to the opposite.

Meanwhile a large number of studies with respect to nutritional factors and exercise have been performed and extensively reviewed (Christensen, 1939b; Åstrand, 1977; Bergström. 1967, 1972; Blom, 1984; Costill, 1978, 1980, 1981; Hermansen, 1967, 1979; Lamb, 1986; Saltin, 1967; Sherman, 1981; Brotherhood, 1984). However, only very few studies have been done with highly trained athletes whereas practically no information is available on the effect of highly intensive long lasting exercise performed on sequential
days. Particularly this type of exercise may have its impact on nutritional parameters and related energy exchange as well as the occurrence of gastrointestinal problems often observed in endurance events. Based on the available data advice is given to athletes and in general this advice is the same for the moderately active jogger as for the elite athlete. In a large number of situations however, theory and praxis fall wide apart. On the one hand studies in which moderately trained athletes are tested may lead to false conclusions with respect to the counseling of highly trained athletes and vice versa. On the other hand nutritionists often claim that a normal composed diet will always be adequate in any situation. These statements are often made without substantial analysis of what is happening in praxis. Therefore it was decided to analyze nutritional problems in elite athletes in the field. More specifically, the question was raised whether or not as a result of daily exhausting exercise athletes differ from a normally active population with respect to eating patterns, food selection and nutritional requirements (Chapter I).

In a second step it was decided to perform a controlled experiment in which highly trained cyclists performed until exhaustion on sequential days and in which the effect of a conventional CHO rich diet on nutritional and metabolic parameters was compared to the effect of the same diet supplemented with a CHO beverage. The idea behind this experiment was to examine the efficacy of diet intervention with respect to meeting the nutritional needs during days of exhausting longlasting physical exercise. Secondly, to study the rational for nutritional supplementation under these circumstances. Such a rational could be obtained from the analysis of the changes in nutritional and biochemical parameters as well as performance capacity (Chapter II, III, IV).

A third step was to study the effect of CHO intake during warming-up on the regulation of the blood glucose concentration during exercise. In contrast to the available studies, which have been performed after an overnight fast and in which CHO intake took place in resting conditions, this study was performed under praxis like circumstances, i.e. after a standardized breakfast and a warming-up procedure (Chapter V).

Finally, in order to give input to a field which until now has been largely ignored by exercise physiologists (Fogoros, 1980; Sullivan, 1981, 1984), it was decided to analyze the available literature in the field of gastro-intestinal physiology and regulation at rest and during exercise. The latter should serve as a reference for future research (Chapter VI).
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CHAPTER I  DIETARY PROBLEMS IN THE CASE OF STRENUOUS EXERTION

F.J.P.H. Brouns, W.H.M. Saris and F. ten Hoor

Part I: A literature review.

Physical exertion and energy requirement.
It is generally known that any form of activity is based on the usage of energy. The more intensive the activity and the longer it lasts, the larger the total energy expenditure. This applies particularly to physical exertion. It is generally assumed that the human body has limited stores of energy. The most important of these are carbohydrates (in the form of glucose and glycogen) and fat.
Since increasing intensity is accompanied by increasing combustion of carbohydrates and decreasing combustion of fat, the quantity of glycogen stored in the body is an important performance-limiting factor (Hermansen, 1967). Without adequate substrate availability the body is unable to adapt to the required rate of ATP synthesis so that the muscle contractions are hampered. Because the store of glycogen is limited, the athlete who has to perform intensively over a longish period will fall off in performance as soon as very low glycogen levels are reached. 'Hitting the wall' in this manner is a well-known phenomenon in sport. Meanwhile, a number of investigations have shown that this occurrence coincides with the low level of muscle glycogen content at that moment (For review see Blom, 1984). Since muscle glycogen is formed principally from the glucose carried by the blood and obtained from the food that the athlete has ingested, it is obvious that proper attention must be paid to food-intake in order to maintain normal muscle glycogen stores. Because a number of enzymes play a key role in substrate combustion and storage, it is reasonable to suggest that the availability of coenzymes such as vitamins, is also of importance. It can be supposed that small deficiencies can lower performance. Although a number of other factors affect the final result, these aspects should not be overlooked now that the difference between winning and losing often can be measured in tenths or even hundredths of second. Because in a number of sports the performance level has become so high that the differences between individuals are only small, endeavors to make further progress have pushed training effort to the limits of mechanical load-ability and energetic power. The latter aspect in particular has led to extremely high energy expenditures in some sports. Whereas for the ordinary active person an energy expenditure of approximately 2.000-2.500 kcal/day (8.3-10.4 MJ) is considered as normal,
endurance athletes under intensive training may have much higher expenditures. Strauzenberg (1978) reports energy intakes ranging from 5.200 to 6.800 kcal (21.7-28.4 MJ) in various kinds of sport. For long-distance runners he reports an average caloric intake of 4.887 kcal/day (20.4 MJ), measured over ten consecutive days, the intake figure for the least intensive day being 3.620 kcal (15.1 MJ) and for the most intensive day 6.000 kcal (25.1 MJ). These values agree with those from other publications (e.g. Mellerowicz and Mellor, 1975; Donath and Strauzenberg, 1975; Haralambie, 1987; Kirsch and von Ameln, 1981; Weicker, 1976). Sometimes for extreme conditions even higher energy consumption figures are recorded. For instance, Biener as cited from Strauss (1979) reports for alpine sports such as climbing an expenditure of 7.000-8.000 kcal (29.3-33.4 MJ) per day. To complete the Wasalöp, a long distance ski-race over 85 km it has been calculated that an energy expenditure of 9.000 kcal (37.6 MJ) is required. Very recently Saris et al. (1986) calculated a mean expenditure of 6.500 kcal per day in professional cyclists in the Tour the France. The expenditure on the most intensive day was approximately 9.000 kcal (37.6 MJ). In the case of top athletes training intensively high energy expenditures happen not occasionally but one day after another.

Therefore in summary as it can be said that, depending on type of sport and duration of event or training, athletes may have an energy requirement two to five times greater than the normal daily consumption of someone who does not go in for sport.

Therefore it is important that the body's stores of glycogen be replenished in time so that optimal initial glycogen levels will be assured.

Physical effort and appetite.

It is generally known that light to moderate physical exercise has a stimulating effect on appetite. Everyone knows the effect of a good walk in the country. However for an athlete involved in strenuous physical exercise the reverse is often the case.

For example it is reported by Karvonen et al. (1978) that with intensive physical activity, carried out repeatedly every hour, the appetite of the person tested decreased sharply so that the next meal was postponed until late evening. Only a few athletes tolerated solid food. Though normal at first, the desire to drink decreased or was suppressed as exhaustion set in. These findings can be related to the negative effect of raised body core temperature and catecholamine levels on appetite (changes that take place during intensive physical exercise). Voelkers (1977) reports from practical observation that after an exhausting day's training American football players sometimes take liquid food because the are just too tired to think of eating anything solid. Donath and Strauzenberg (1975) suggest that an inverse relation between energy consumption and food intake may exist. On days of intensive physical
activity most athletes appeared to have insufficient appetite to compensate for the high energy expenditure. However on days of reduced intensity this energy was again made up, the intake being then greater than the requirement at that moment. Also Åstrand and Rodahl (1977) write: It is a common experience that athletes in active training often are unable to maintain regular meal-schedules and may incur detrimental effects.

The effect of mental stress.
Besides physical stress it appears that psychological stress can also adversely affect appetite and hence influence the intake of food. According to Canham and Consolazio (1965) athletes about to take part in an important competitive event often react in the same way as soldiers about to go into battle, namely with anorexia, nausea and sometimes vomiting. Rose and Fuening (1960), state that there is objective evidence that functional changes (e.g. in motility) take place in the gastrointestinal system immediately before and during an important sport event. Upjohn (1953) reports that it takes about 3-4 hours for the stomach to empty after a normal meal but that this time increases to about 6 hours due to the psychological stress that precedes a competitive event. If this situation occurs frequently it may have harmful consequences for the athlete if the right measures are not taken.

In summary it can thus be said that physical and psychological stress have an adverse influence on appetite. This loss of appetite occurs chiefly before, during and after the competitive event. As a result of this, when there is strenuous physical effort on successive days or at short intervals, inadequate carbohydrate intakes may occur.

Sport and frequency of meals.
As the energy expenditure of athletes increases, it appears that the normal feeding pattern suffers more and more. This is in fact a logical result of the fact that an increased energy expenditure is the direct corollary of a lengthened physical working time. Thus, if training is done for a great many hours per day, this will mean that the time for preparing and digesting well composed and varied meals is limited (Wilmot and Freund, 1964; Brouns, 1984). We know from practical experience that many people find it does not suit them to indulge in sport on a full stomach. This naturally has its repercussions on the feeding pattern of an athlete under intensive training who necessarily worries about it. In some sports a full stomach may even be a 'contraindication', e.g. boxing, karate and jumping, owing to the risk of possible stomach injuries. But here too there is a real problem in that the athlete must try to cater for his high energy requirement. Therefore a direct relation between daily physical activity and food intake can be assumed. In a normal situation one can say that the
problem can usually be solved by eating more. In practice however this is
often difficult for an athlete. This is also recognized by Mathews, and Fox
(1979) who write that it is perfectly possible to divide a 5,000 kcal diet over 5
meals, but that this sort of eating creates a problem for the athlete faced with
long periods of consecutive physical exertion. E.g. in the case of the
decathlon or tennis championships when singles and doubles have to played
on the same day, and also in tournaments. In such a situation there may be a
time limitation combined with a decreased appetite and a bad tolerance for
sporting on a full stomach.
Haralambie (1978) states that a consumption of 5,000-6,000 kcal (20.9-25 MJ
) will probably force an athlete to resort to high energy food. From cycle
racing, Tour de France etc., we know that cyclists have been taking liquid
baby food on competition days for the past decade, as this is fairly easily
digested. This is not generally appreciated by the athletes, however, the
alternative may be not eating at all.
In view of the advice often given by dieticians to the effect that a normal
composed menu always suffices and that the athlete needs no adaption of his
dietary regimen at all must be considered as illogical. There is a nutritional
advice, generally advanced, that no food should be eaten within 3-5 hours
before a strenuous physical exercise (competitive event) or at any case only a
very small quantity of easily digested food may be taken (Mathews and Fox,
1979; Smith, 1979; Lamb, 1980; American Dietetic Association, 1980, a.o).
Top athletes say that the training effort is probably on a par with or even
greater that the required for competition. On that basis this advice would thus
hold good for any training session. This would leave a top athlete training
twice a day with about 7 hours in which to consume the greatly increased
volume of food spread over several meals. But practical experience shows
this to be impossible. In consequence one falls into a continuous eating
pattern, consuming snack food to a large extent and food with a high energy
content. At the same time frequent use is made of sugar and sugar solutions.
The continuous eating pattern (lots of snacks spread over the day) is
supported by a study by Kirsch and von Amelia (1981). The authors studied a
group of long-distance runners with a moderately elevated energy
expenditure averaging 3,316 kcal (13.8 MJ) per day and a group of racing
cyclists with a drastically raised energy expenditure averaging about 6,280
kcal per day (26.2 MJ). They found that the food intake frequency was
governed by the total energy expenditure. The runners took 4-7 meals a day
but this fragmented pattern was far more pronounced in the case of the
cyclists with 6-8 meals a day. They concluded that the cyclists were more or
less continuously eating and drinking; in 57% of cases they ate and drank at
intervals of less than 1,5 hours. In 90% of cases the interval was less than 3
hours. In these circumstances athletes must have access to food and drink
throughout the whole day since they tend to eat anything that is easy to
prepare or ready for use, tasty and with a high energy content (preference for sweet foods). This often results in frequent consumption of cream cakes, chocolate and sweets. In addition a lot of sweetened soft drinks are drunk and, in some sports also beer (Saris, 1980).

As regards the situation in Holland, this has been confirmed in a study of De Wijn and Van Erp-Baart (1980), who found that in the case of competition rowers some 35% or more of the total daily energy input is obtained from 'bites of food in between', such as snacks, sandwiches, biscuits, etc. As these foodstuffs mostly have a high energy content but little protein and vitamins. Such a nutritional pattern leads to a lowering of the nutrient density. (The presence of micro nutrients related to macro nutrients).

In summary it can be stated that:
1. Athletes with a high energy consumption often get into a continuous eating pattern consisting of many small 'meals' at short intervals.
2. Increased training time is attended by increased energy expenditure so that a greater volume of food must be assimilated but with less and less time for eating.
3. Intensive sports activity is inadvisable on a full stomach.
4. To cover the energy (carbohydrate) requirement on days of very strenuous training or competition, it is recommended to take liquid food with a high carbohydrate content in addition to the 'normal meals'.
5. The advice that athletes can at all times manage with a normal composed (solid) diet is in fact erroneous and based on insufficient know how of sports practice.

Liquid food concentrates.
From the foregoing it is clear that an adequate energy supply for athletes is actually at its most important in periods when the feeding pattern is most exposed to disturbing factors. Should this be inadequate, then premature depletion of glycogen stores may have a very decisive effect on the end result. Athletes with a high glycogen content perform longer (for review see Blom 1984) and when a low value is reached (about 3 g/kg muscle weight) the athlete is forced to discontinue his activity (Karlsson and Saltin, 1971).

For the same reason the supply of carbohydrates shortly before, during and directly after regular prolonged intensive physical effort is an important measure if one wants to stay in the race (Sherman and Costill, 1984). In this case only liquid food may be tolerated optimally.

From studies in which solid food was replaced by liquid food before a competition it transpires that nausea, vomiting and stomach cramps were eliminated and dryness of the mouth did not occur so much (Rose et al., 1961;
Cooper et al., 1962). At the same time it is possible by this means to better maintain the blood glucose level, whereby a glycogen sparing effect can be achieved (Pirmay et al., 1977), and exhaustion of energy averted. Just how important it is for an intensively training athlete frequently exposed to strenuous physical exertion - as for instance a racing cyclist - to secure a large carbohydrate intake, is apparent from recent studies. It is known that the resynthesis of muscle and liver glycogen after prolonged exhausting effort can take 2-3 days with a normal diet (solid food with a composition of about 50 en% carbohydrates 35 en% fat and about 15 en% protein) (Costill, 1979). (energy percent is the proportion in the energy intake, e.g. intake 2000 kcal, carbohydrate 100 gram = 100 x 4 kcal = 20 en%).

This means that if exercise is repeated within this period, the athlete will cope with a decreased endurance capacity due to reduced muscle glycogen stores. By ensuring an elevated carbohydrate content in the food and a supply of food starting immediately after the sports effort, the time needed for glycogen resynthesis can be reduced substantially (Costill, 1979). This is an essential prerequisite. Otherwise, competitive events lasting several days like the Tour de France would be hard to perform. The carbohydrate content should then be at least 60 en% (Schneider, 1980; Costill, 1980). One favorable point is that the activity of the enzyme glycogensynthetase varies in inverse proportion to the decrease of glycogen content. This enzyme activity appears most pronounced in the first few hours following effort. It is precisely in this period that the athlete has to contend with a suppressed appetite so that a liquid carbohydrate solution may offer the best alternative (Schneider, 1980; Brooke, 1974; Stucke et al., 1976). As in the case of orally administered food in intensive care, attention must be equally paid in competitive athletics and first-class sport to ensure that the gastro-intestinal tract needs to make as little effort as possible to digest and absorb the food administered.

It is known that liquid food has a quicker absorption so that the food taken is made available sooner to the recovery processes. X-ray examination of the gastro-intestinal motility has shown that liquid pre-game food passes out of the stomach within 2 hours and is completely digested and absorbed within 4 hours after intake. On the other hand, after a pregame meal of solid food, the food was still in the gastro-intestinal tract 4-6 hours afterwards because of the slower rate at which the stomach emptied in these conditions (Rose-Fuennng, 1960).

The consequences of such dietary measures before and after competition become evident in practice. Athletes have less gastro-intestinal problems. Those who take high-energy (carbohydrate) liquid food immediately after the cessation of sports effort say that they recover sooner and are able to settle down to a good meal earlier.

This is also confirmed by the recent findings of Ionescu (1982) and Dragan et al. (1982).
In summary it can be concluded that:
1. In periods when carbohydrate intake plays tricks on the athlete most (shortly before, during and directly after prolonged exertion) it is at its most urgent.
2. Solid food is badly tolerated just before, during and directly after strenuous exercise.
3. For adequately quick glycogen resynthesis a high carbohydrate content in the food (60 en%) is important.
4. Liquid food concentrates offer athletes the readiest option in periods when eating is difficult or impossible or can also be used as an energy supplement.
5. Athletes should be advised about their feeding habits and the composition of their food especially with regard to days of intensive training and competition.

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CHAPTER I: PART II: THE TOP ATHLETES’ MENU, NUTRIENT DENSE ENOUGH?

In a recent thesis by W.H.M. Saris (1982) we find the following statement: ‘Healthy foods do not exist’. The question is how one should interpret this for people who go in for strenuous sport, have a high energy expenditure and need a high carbohydrate proportion in their diet. Since all nutrients are interrelated one might argue that too much or too little of one particular nutrient will influence the other nutrients e.g. their absorption, utilization etc. It would in general be fair to state that meeting the daily requirement of all nutrients (in so far as this is known) comes nearest to satisfying the definition of a ‘healthy diet’. It would be nice to know this exact daily requirement for every individual but unfortunately we do not know. Most recommendations are based on a survey of large groups of people in which differentiation is made between young, old, male, female, gentle activity, heavy physical work, etc. However, a drawback to this is that feeding habits vary from country to country and that at the same time what is generally applicable for the large group will certainly not always necessarily apply to a specific target-group. Some target-groups have their own specific nutrient recommendations, such as old people, infants and pregnant women.

However, for the group of top athletes there are only very few data available. Opinions vary widely and little research has yet been done. Nevertheless, certain tendencies can be signalized which make it possible to advice athletes and their coaches based on the current status of knowledge. One of these tendencies is that deficiencies, however small they may be, can have adverse effects on the bodily functions in certain circumstances. In fact, deficiencies can result in a lowered performance, especially in periods of stress such as illness or strenuous physical exertion. That is why certain nutrient deficiencies of which it is known that they may have a drastic effect on bodily function, such as vitamins, proteins and carbohydrates, are in fact of such great importance to athletes and their coaches.

Vitamins and sport.

In almost every recent textbook on the subject of sports physiology and training theory one can find a chapter on food and particularly vitamins. The question generally considered is whether there is a need to give the athlete extra vitamins, i.e. whether intensive sports activity leads to an increased vitamin utilization or requirement. Conflicting views based upon limited evidence may confuse many readers, e.g. Nöcker (1978) states that vitamin supplementation will only bring about improved performance if the athlete has a vitamin deficiency. Well-fed athletes will have no benefit at all, an opinion shared by Vitousek (1980). However, advised intakes are often much
higher than the normal recommended daily intakes. Fox (1979), Huse-Nelson (1977) and Mashford (1973) are of the opinion that there is no real point in discussing vitamins and that the daily requirements can be amply covered by the normal diet.

Williams (1976) concluded after his extensive study of the literature that the administration of a vitamin in general has no effect upon performance. This conclusion is, however, only valid when there are no deficiencies. He goes on to say that more research is necessary on the effects of vit. B-complex, vit. C and vit. E in as much as the results and opinions obtained (up to 1976) are often contradictory.

McArdle, Katch and Katch (1981) suggest that vitamins can be used repeatedly in metabolic reactions, i.e. they are not lost, so that the requirements of athletes are no higher than those of people not in training. However this statement is in conflict with the view that the vitamin B1 requirement is directly linked to the carbohydrate proportion in the diet (National Research Council, 1980).

The conflicting opinions raise three important questions:
1. Whether or not athletes have a higher vitamin utilization, and consequently a higher vitamin requirement, than non-athletes.
2. Whether the feeding pattern, and thus the vitamin intake of athletes is indeed such as to be considered adequate.
3. Whether or not administration of a very high vitamin dosage will have an ergogenic effect.

Evidence of an increased nutrient requirement.

Many sports physicians express the opinion that the overall vitamin requirement of athletes is raised so that extra supplementation is indicated. For instance, Strauzenbarg (1978) reports that the vitamin intake of top athletes in East Germany is too low to meet the requirements (Table 1). According to Strauzenbarg the increased requirement is most pronounced in the case of endurance athletes.

Cureton (1969) states that intensive training leads to a loss of vit. B and vit. C from the body. Buskirk and Haymes (1972) suggest that training brings about an increase in the number of mitochondria and enzymes, functional proteins that play an important part in the energy metabolism. For this reason extra vitamins (cofactors) would be needed to make the enzymatic reactions possible. Prokop 1978 also supposes this.
Table 1. Vitamin requirements and intakes of East German top athletes (Strauzenberg, 1978).

<table>
<thead>
<tr>
<th>Vitamin</th>
<th>Estimated requirement</th>
<th>Daily intake</th>
</tr>
</thead>
<tbody>
<tr>
<td>A</td>
<td>13.300 IU</td>
<td>13.700 IU</td>
</tr>
<tr>
<td>B1</td>
<td>4-8 mg</td>
<td>1.9 mg</td>
</tr>
<tr>
<td>B2</td>
<td>4 mg</td>
<td>2.9 mg</td>
</tr>
<tr>
<td>Niacin</td>
<td>30 mg</td>
<td>18 mg</td>
</tr>
<tr>
<td>C</td>
<td>500 mg</td>
<td>244 mg</td>
</tr>
</tbody>
</table>

Table 2. Estimated vitamin requirements for athletes (Creff-Bérard, 1976).

<table>
<thead>
<tr>
<th>Vitamin</th>
<th>Untrained persons</th>
<th>Athletes</th>
</tr>
</thead>
<tbody>
<tr>
<td>B1</td>
<td>1.5 mg</td>
<td>5 - 10 mg</td>
</tr>
<tr>
<td>B6</td>
<td>4 mg</td>
<td>15 - 30 mg</td>
</tr>
<tr>
<td>B12</td>
<td>2- 5 mcg</td>
<td>10- 20 mcg</td>
</tr>
<tr>
<td>C</td>
<td>75-100 mg</td>
<td>150-300 mg</td>
</tr>
<tr>
<td>B2</td>
<td>2.5 mg</td>
<td>10- 15 mg</td>
</tr>
<tr>
<td>Nicotinic acid (PP)</td>
<td>20 mg</td>
<td>30- 50 mg</td>
</tr>
<tr>
<td>Pantothenic acid</td>
<td>10 mg</td>
<td>10- 20 mg</td>
</tr>
<tr>
<td>Biotin</td>
<td>5 mg</td>
<td>10- 20 mg</td>
</tr>
<tr>
<td>Folic acid</td>
<td>10- 20 mg</td>
<td>15- 40 mg</td>
</tr>
<tr>
<td>P</td>
<td>10- 20 mg</td>
<td>20- 40 mg</td>
</tr>
<tr>
<td>A</td>
<td>2000- 6000 mg</td>
<td>50000 IU</td>
</tr>
<tr>
<td>D</td>
<td>400 IU</td>
<td>400 IU</td>
</tr>
<tr>
<td>E</td>
<td>10- 30 mg</td>
<td>30- 50 mg</td>
</tr>
</tbody>
</table>
Mashford (1973) links the increased vitamin requirement to the greater substrate utilization, and so too does Troll (1978). Creff Béard (1976, 1980), Kindermann (1977), Schneider (1979) and Jakowlew (1975) suggest an increase of 200% of all vitamins except vit. B and C for which the figure is 500% (Table 2).

However, it has to be underlined that all these figures are still unsupported by solid research. In this connection it is also essential to know how one defines the term 'extra vitamin requirement':

(a) extra vitamin requirement above normal RDA, on the basis of an increased energy expenditure (if energy expenditure is doubled, the vitamin requirement is doubled too);

(b) an extra requirement over and above the adjustment mentioned in (a), e.g., energy expenditure is doubled but vitamin requirement is 5-fold.

We will consider extra vitamin requirements as described under (a).

In other publications the existence of an increased demand is denied although it is stated that, as a safety measure, a small vitamin supplement can do no harm (Ardie, Katch and Katch, 1981). Nevertheless, these authors also state that excess should be avoided as a matter of course: 'It is a great concern, however, that some athletes resort to taking megadoses or doses of at least tenfold the recommended daily allowances in the hope of improving performance'. The above mentioned safety measures play a very important part in sport since nobody can guarantee the accuracy of the normal current requirements and nobody is in a position to state the requirement of every individual athlete. Schreurs (1981) also considers this problem when stating in connection with the vitamin recommendation of the Netherlands Food Council: 'However, it seems premature and even wrong to conclude from these facts (indicated requirement and average daily intake) that every individual or the majority of Holland's population has an optimal vitamin supply'. At the same time Schreurs states that absorption disturbances and inadequate food intake are not the only reasons for vitamin deficiencies, but that these can also be caused by increased requirements arising inter alia in connection with physical exertion, periods of accelerated growth, excessive losses, etc. A reason often put forward is that athletes can have an increased vitamin loss. It has been suggested that the losses of vitamins via the sweat and urine (particularly vit. C) is increased during prolonged activity (Sanger and Israel, 1975; Nöcker, 1978; Mathews and Fox, 1979.). These suggestions are taken to heart by trainers and athletes and are fundamental for the swallowing of what are often large quantities of vitamins for fear of deficiencies and lowered performance due to loss of vitamins. However one would assume that normally such increased losses could be compensated by the larger food intake of the athlete (at least if the quality is excellent).
But is this the case?

Evidence of insufficient Intake.
The suggestion that an inadequate feeding pattern and/or poor quality food will have a bad effect on the nutrient supply is of course nothing new. But whether this applies to the hard training athlete is a matter upon which opinions differ. The following data however indicate that this may well be true. Recent studies have brought to light the existence of biochemical vitamin deficiencies among athletes, in particular of the B complex vitamins (Kindersmann, 1977; Haralambie et al., 1975; Howald and Segesser, 1975). (marginal levels of the fat soluble vitamins have until now never been found in athletes). In such situations supplementation with a vitamin mineral preparation indeed led to an increase in performance. With optimal nutrient intake this would not have been possible.

Keul et al. (1979) reported a performance improvement of 4.3%, reflected in an improved running performance and speed. The pulse rate was significantly slower and the incidence of muscle cramps was also less after supplementation. Haralambie et al. (1975) registered an improved performance chiefly in the last stage of a long lasting exercise trial. Van Dam and Waterlooh (1979) registered a better resistance to fatigue, shorter reaction times and a better movement efficiency in fencers. In a comprehensive study on physical education students by Van der Beek et al. (1981) a few biochemical deficiencies were also found but disappeared after vitamin supplementation.

These authors found no improvement in performance after supplementation. However, in a very recent follow-up study, in which a diet with a too low vitamin content (vit. B1-B2-B6 and C) was taken for several months, they found a significant decrease in performance. This like-wise vanished after supplementation (Van der Beek et al., 1984).

Bamji et al. (1982) found a large number of biochemical deficiencies (vit. B2-B6) among rural schoolchildren. After supplementation they found a significant improvement of a psychomotor test. Although the group which they studied was not representative for our European population, and certainly not for top athletes, their final conclusion is of interest to us:

'These data also suggest that a functional impact of vitamin supplements may be seen even in the absence of clearcut clinical impact as judged by established nutritional deficiency signs'.

In other words, small shortages manifested as biochemical deficiencies (without clinical deficiency symptoms) can well affect physical performance, all the more so when one reflects on the often large intake of food between meals such as cream cakes, chocolate, etc., as mentioned by De Wijn and Van Erp-Baart (1980). These foods, high in energy but low in vitamins, minerals and protein, accounted for more than 35 en% in the total energy intake of the group of oarsmen studied. A Marginal vitamin intake may then
be expected. The best advice must then be to avoid these 'empty calories' and make use of good quality between-meal foods such as muesli, dark bread, almonds and raisins etc.

However, even doing so, in the case of very strenuous sports 'deficiencies' can still arise. To quote Van Dam (1978) and Van Dam and Waterlooh (1979): 'Even dietetically composed nutrition is not always sufficient to cover the vitamin needs which can increase a great deal in active athletes'.

This agrees with the findings of Saris et al. (1986) who observed during the Tour de France (23 days cycling competition) that the vitamin B1 intake, expressed as nutrient density (the presence of nutrients in food in relation to the energy intake; example: RDA of vit. C=50 mg/day.

Average energy intake = 2000 kcal (8.4 MJ). Nutrient density of vit. C is than \( \frac{50}{2000} = 0.025 \).

When energy intake is doubled and vit. C intake remains the same, the then nutrient density decreases to \( \frac{50}{4000} = 0.013 \).

The intake of vitamin B1 was too low on successive days of the competition (0.26 mg/1000 kcal as against 0.43 mg/1000 kcal on days of rest) (the recommended daily allowance (RDA) American National Research Council Recommendations) is 0.5 mg/1000 kcal). This was caused by the high energy intake from liquid and solid carbohydrate-rich meals with a too low content of B-complex vitamins due to the refinement of the foodstuffs used. The normal meals on these competition days were of good quality so that extra supplementation appeared to be the only solution to maintain an optimal nutrient density. This in fact may justify the extra care that many trainers take in practice to ensure a 'good' vitamin intake by advising vitamin preparations.

From the foregoing it may be concluded that: (a) there are indications that vitamin requirements in athletes may be increased as a result of their increased energy turnover; (b) that vitamins may be lost from the body as a result of intensive long-lasting physical exercise; (c) that even eating 3 meals of high quality per day will not per se guarantee an optimal vitamin status and; (d) that frequent use of energy snacks high in carbohydrate but low in vitamin content may also contribute to this.

**Ergogenic effects of overdoses of vitamins.**

The question now arises as to whether megadoses vitamins stimulate performance or do not. The answer to this question is a short sharp no! Vitamin preparations only have an optimizing effect on performance when intakes are too low. There is not a single investigation up till now that can offer evidence in support of performance boosting effects when intake is
sufficient c.q. when no biochemical deficiencies can be found. Lately, vitamin E has aroused strong interest among coaches since it was suggested that this vitamin would increase the oxygen utilization in the muscle cell so that less lactate would need to be produced (Weiner and Rothschild, 1980). However, carefully controlled trials have failed to show any effect (Lawrence et al., 1975; Sharman, 1976; Talbot and Jamieson, 1977; Bell and Johnson, 1976). For overview see Shephard (1983). Another important aspect is that a megadose of a single nutrient will always interact in some way with other nutrients. These interactions can be highly undesired. The application of simple vitamins, e.g. B12 injections, shots of vit. C, etc., as often happens in sports practice, must be advised against because of possible undesirable interactions. The point is that the lack or excess of one nutrient may often affect the requirements for other nutrients. Wise (1980) and Levander and Chang (1980) report a large number of such interactions. It is known for instance that vit. C promotes the iron absorption in the gastrointestinal tract which may be important especially for endurance athletes. This is a positive interaction. One-sided interpretation however may create problems because high doses of vitamin C give rise to an inadequate absorption of vitamin B12 (Herbert et al., 1977; Newmark et al., 1976), which in its turn adversely affects the absorption of vit. B6 and B2 (Wise, 1980).

How to solve the problem?
Since even dietetically well composed meals may lead to biochemical deficiencies in periods of regular intensive training and on the other hand possible negative interactions should be avoided, two questions in the matter of measures to be taken to bridge the possible gap now appear to be relevant:

1. Which vitamins?
2. In what quantities:
   If we start by dividing vitamins into two categories: (a) fat soluble and (b) water soluble, then it is fair to say that as regards the former there is so far not a single indication that a supplementation is needed for athletes involved in strenuous sport. Athletes should be advised to take sufficient green vegetables and milk in their daily diet to ensure a sufficient content of vitamins A and D. There is no need for vit. E and K supplementation. Because of its interaction with coagulation of the blood, vitamin K may even be harmful. For water soluble vitamins the situation is rather different. Because of marginal intakes on the one hand and increased losses on the other, supplementation may not be out of place for the active athlete as far as competition and/or periods of intensive training are concerned. Primarily the intakes should be met by the choice of high quality foods. But as has been described before, this seems not always to be adequate enough. This applies particularly to the following vitamins: B1, B2 (because of their

From this it can be concluded that:
(a) There are no arguments to support increased needs for the fat soluble vitamins A, D, E and K even if daily energy turnover is very high.
(b) Intake of high doses of water soluble vitamins is not, as always assumed, entirely without undesirable side-effects.
(c) Megadoses of vitamins are not of benefit to athletes by means of improving performance.

Nutrient density.
Since 'how much is enough', 'how much is too much', 'how much is minimal' are all questions very difficult to be answered for each individual it may be important to search for an alternative which can be used safely. We have the opinion that the principle of nutrient density might be an adequate alternative, in so far as water soluble vitamin supplementation is desired (vitamin intake related to energy intake). By this means overdosing and underdosing can always be avoided and undesirable interactions precluded.
The nutrient density for vit. B1 is 0.5 mg/1000 kcal (American National Research Council, recommendations 1980). An athlete who expends 6000 kcal in one day would therefore have to have a vit. B1 intake of 6x 0.5=3 mg. If he gets 1 1/2 mg from his normal meals he will need to have a 1 1/2 mg from his energy-rich in-between-meals. If following this principle an athlete procures an energy concentrate that has to be taken say while cycling, then the vitamin content in that concentrate should be at least in accordance with the desired nutrient density. For instance, an energy concentrate containing 500 kcal should contain a minimum of 1/2x0.5 mg, i.e. 0.25 mg vit. B1. Seen from this standpoint, liquid foods ought always to be vitaminized at least following this principle in order to be of real nutritive value to the athlete. By this means an adequate nutritional intake would be possible from the normal solid meals and the food concentrates taken before and during competition. The swallowing of unnecessary large quantities of vitamin pills, to be on the safe side, would thus become superfluous. This finally leads us to the following conclusion:

The nutrient density concept gives a basis for the minimal enrichment of daily nutrition with vitamins, if necessary, in such a way that athletes using such products will never be exposed to excessive vitamin intakes, while at the same time being assured that their minimal requirement may be adequately met. The nutrient density concept is a safe guideline for vitamin supplementation, is easily understandable and can be valuable for the
Summary
Athletes involved in strenuous exercise of long duration tend to change their dietary habits depending on the daily quantitative energy expenditure. These changes may be positive, such as increased meal frequency or negative as far as composition of the diet is concerned.

This article reviews aspects of energy expenditure and its relation to dietary habits, quantitative and qualitative intake of macro- and micro-nutrients. The influence of sports activities on possibly increased requirements for vitamins is discussed and an attempt is made to relate micro-nutrient intake to daily energy needs. This results in the proposal of a (minimal) nutrient density concept for athletes involved in heavy training.

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CHAPTER II: EATING, DRINKING AND CYCLING, A CONTROLLED TOUR DE FRANCE SIMULATION STUDY, PART 1.


Introduction
Consensus exists among most nutritionists that a normal, quantitatively and qualitatively well balanced diet will be sufficient for any sportsman or woman in any situation. However, it has recently been described that athletes performing intensive long lasting exercise on a day to day basis are faced with the problem that consuming a normal well balanced diet may become quite impossible (Brouns, 1986). The reasons for this are the following: 1.- The adverse effect between physical and mental stress and appetite; 2.- Intolerance for large food volume in the exercising athlete, whenever food intake exceeds 20-25 MJ/day; 3.- The reduced number of ‘resting’ hours available for consumption and digestion.

It has been suggested that an inverse relationship exists between food intake and energy expenditure (Donath, 1975) so that it may be impossible to maintain regular meal schedules and sufficient energy intake on days that energy expenditure is very high. In order to compensate, athletes may fall into a nibbling eating pattern, more or less eating continuously throughout the day (Kirsch, 1981; v. Erp-Baart, 1988), thus avoiding a large bulk of food in the stomach, which may induce abdominal distress. These nutritional problems in endurance athletes are well known in practice. However, no controlled studies have been performed. Another widespread belief is that the ingestion of water will always be better for reasons of rehydration and that there is no reason to combine the fluid with nutrients. This in contrast to studies that show that glucose and electrolyte containing beverages restore plasma volume better than water alone, (Costill, 1973; Brandenberger, 1977; Candás, 1977) and to the increasing evidence that ingestion of plain water may lead to abnormally low levels of plasma electrolytes, whenever the volumes of fluid replaced are substantial (Noakes, 1985; Hiller, 1985, De Combaez, 1981).

From observations of professional cyclists during the Tour de France (Saris, 1988) it is known that it is possible to maintain adequate energy and fluid intakes over a prolonged period of competition, even when energy expenditure exceeds 24 MJ/day and fluid intakes (in order to compensate losses) reach a level of more than 10 l/day. However, these cyclists were only able to do so during competition - which may last up to 8 h/day - by ingestion of energy-rich foods, to a large extent as CHO- and electrolyte-containing
liquids. Those subjects who were not able to maintain energy and fluid balance frequently experienced malperformance and sometimes were forced to quit prematurely. This imbalance was commonly a result of poor food intake and/or gastrointestinal disturbances such as diarrhea causing malabsorption and excessive fluid loss.

In professional cyclists CHO intake is closely related to energy intake since the major portion of energy in the selected diet (more than 60%) comes from CHO (Saris, 1988). Because CHO is a performance-limiting substrate in endurance exercise whenever exercise intensity exceeds 50-60% of maximal oxygen uptake, it may be assumed that a negative energy balance, also leading to a negative "CHO balance", may impair performance progressively. This may happen as a result of insufficient glycogen repletion on a day-to-day basis due to insufficient CHO intake. For this reason it was decided to do a study on the effect of repeated exhaustive endurance exercise on nutritional indices in which the data obtained in cyclists ingesting a CHO-rich diet composed of normal foodstuffs (N) could be compared with those obtained after the same diet supplemented with CHO-concentrated liquids. In addition it was decided to study the daily contribution of CHO-, fat- and protein-metabolism to total energy exchange under these circumstances. The study was performed in a cross-over design with a randomized order of treatment. The part of the study in which the subjects received the normal CHO-rich diet will be described and discussed in this Chapter. The effect of dietary manipulation will be discussed in Chapter III. The aim of the study was to test the following hypotheses:

- Cyclists having ad libitum available a normal but CHO-rich diet will be able to maintain energy balance and to meet the daily CHO needs even in situations of repeated, high-intensity, sustained cycling exercise.
- Cyclists having ad libitum available ordinary and mineralized drinks throughout the exercise period will be able to compensate for the exercise-induced fluid losses.

PROCEDURES AND METHODS

Subjects
Thirteen highly trained cyclists participated in this study. Physical characteristics are presented in Table 1.
Table 1 Physical characteristics of experimental subjects

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age (yrs)</td>
<td>20.0 ± 2.1</td>
</tr>
<tr>
<td>Weight (kg)</td>
<td>73.3 ± 1.7</td>
</tr>
<tr>
<td>Height (cm)</td>
<td>179.6 ± 1.7</td>
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<td>VO₂ max (ml.kg⁻¹.min⁻¹)</td>
<td>65.1 ± 1.2</td>
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<tr>
<td>Wmax (Watt)</td>
<td>390 ± 8.0</td>
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<tr>
<td>Wmax.kg⁻¹ (Watt)</td>
<td>5.3 ± 0.4</td>
</tr>
<tr>
<td>Body fat (%)</td>
<td>12.0 ± 0.8</td>
</tr>
</tbody>
</table>

Mean values ± SEM (n=13)

Experimental design

The subjects were asked not to participate in vigorous training or competition during the two days preceding the stay in the laboratory and to ingest a normal but CHO rich diet. All subjects were informed about the nature, purpose and possible risks of the study, before giving their voluntary written consent to participate. The experiment was conducted over seven sequential days using a semi-automated respiration chamber system.

The subjects reported to the laboratory on Sunday evening (day 1) at 9:00 PM in order to get accustomed to the chamber (Fig 1). During the first day the chamber was not closed. They received a time schedule for all activities in the forthcoming week. The next morning (day 2) a by each subject selected supply of food and fluid of known quantity and composition was made available. This supply was provided each day until the end of the experiment on Saturday morning 9:00 AM six days later. Foodstuffs for breakfast and lunch were supplied at 7:30 AM. However, extras could be obtained throughout the day upon request. Dinner was served at 6:00 PM. There were no quantitative limitations. The cyclists were instructed about the importance of adequate food and fluid intake and were encouraged to eat and drink as much as desired.

Weight and volumes of foods and drinks were measured and registered in a diary allowing for analyses of 24 h intake as well as intake over specific periods of the day. The latter was done in order to analyze main meal and in-between meal energy consumption. All leftovers were re-weighed and accounted for in the final calculation of energy and nutrient intake. Energy and nitrogen content of all food items had been previously determined. Protein intake was calculated daily immediately after dinner. To assure a minimum intake of 1.2 g.kg⁻¹, subjects were supplemented with a protein con-
centrate whenever intake was calculated to be too small for that day. The reason why protein intake was set at a minimum level is discussed later.

Actual performance capacity Wmax (Watt) and maximal oxygen uptake (\(\text{VO}_2\text{max; l.min}^{-1}\)) were determined at 10:00 AM on day 2 during an incremental bicycle ergometer test.

![Graph](image)

Fig 1  Experimental schedule. On day 2 actual performance capacity (Wmax; Watt) and maximal oxygen uptake (\(\text{VO}_2\text{max; ml.min}^{-1}\)) were determined at 10:00 AM using an incremental bicycle ergometer test. Day 3 and 6 were rest days; cycling at 40% Wmax during 45 min at 10:00 AM and 2:00 PM. Day 4 and 5; exercise to exhaustion. Day 7; end of the experiment at 9:00 AM. A muscle biopsy (Chapter V) was taken on days 2, 5 and 6. Blood sampling was done at 7:00 AM, 12:00 AM and 4:00 PM on days 3 to 6.

The respiration chambers were closed at 4:00 PM on day 2 and measurements for calculation of energy expenditure were started. Days 3 and 6 represent standardized rest days during which each subject cycled for 45 min at 40% Wmax at 10:00 AM and 2:00 PM respectively. Rest days were included in the program in order to determine resting metabolism and resting food and fluid intake for comparison with those gathered from the two exhausting exercise days. On days 4 and 5, each subject exercised until exhaustion. Exercise was started at 10:00 AM with a 30 min warming-up, followed by exercise at intensities of 80% and 50% Wmax respectively. 80% and 50% Wmax were chosen in order to mimic riding ahead of the group or benefitting from wind shielding within the group respectively. At 12:00 AM exercise was interrupted for 5 min to collect blood samples, to measure body weight and to change the sweat capsules. Thereafter exercise was continued
at 50% and 60% Wmax allowing for food intake. After 3 h 44 min exercise was continued again at 80% and 50% Wmax in 3 min intervals. Finally at about 2:30 PM, the Wattload was set at 90% Wmax and the cyclists were asked to maintain pedaling frequency greater than 60 revolutions per min. 90% of Wmax - equivalent to about 80% VO₂ max - was chosen in order to simulate a finish on a mountain top (Fig 2). During exercise, fluids were available ad libitum (tea, coffee, milk, lemonade and a placebo 'sport drink' - artificially sweetened, colored, and mineralized water).

![Graph showing cycling program](image)

**Fig 2** Cycling program during days 4 and 5. There was a warming-up of 30 min at 30% Wmax, followed by 10 min 50% Wmax, followed 9 times by 6 min 80% Wmax and 6 min 50% Wmax. Thereafter exercise was shortly interrupted for collecting blood and sweat samples and was then continued at 50% Watt for 10 min, followed by 50 and 60% Wmax both for 30 min. Exercise intensity was then increased to 80% Wmax for 3 min followed by 3 min at 50% Wmax, which was repeated 8 times. Finally exercise intensity was increased to 90% Wmax which had to be performed to exhaustion.

Blood samples were drawn into EDTA evacuated tubes from a teflon catheter which was inserted into an antecubital vein at 7:00 AM and was connected with a three-way stopcock. Sweat samples were collected by absorption into pre-dried pads located in water and air tight capsules (Lamon, 1983). The sweat capsules were placed in the infraspinous fossa of the scapula and kept in place by an elastic mesh vest.

Body weight (naked) was measured daily with a digital balance accurate to 100 g, and registered at 7:00 AM, 12:00 AM, 3:00 PM and 9:00 PM respectively. 12:00 AM and 3:00 PM coincided with halfway and end of cycling exercise on days 4 and 5. Feces and urine were kept at -20°C in a
deep freezer toilet and were collected in 24 h periods.

Analyses
Energy expenditure was measured using an indirect automated calorimeter as previously described by Schoffelen et al (1986), allowing for continuous determination of VO₂ and VCO₂ and registration of environmental conditions (temperature, relative humidity and barometric pressure). Gasflows were measured with a dry gasmeter (Dort, the Netherlands), oxygen was analyzed using a Servomex® paramagnetic oxygen analyzer (Taylor, England) and carbon dioxide using an infrared CO₂ analyzer (Hartmann and Braun, Germany).

Energy expenditure was calculated according to the formula of Consolazio (1963):

$$E = 3.78 \: \dot{V}O_2 + 1.14 \: \dot{V}CO_2 - 2.98 \: N_u$$

Where E is energy expenditure (kCal), $\dot{V}O_2$ is oxygen consumption (ml.min⁻¹), $\dot{V}CO_2$ is CO₂ production (ml.min⁻¹) and Nu is the amount of nitrogen excreted in urine (mg).

Daily energy balance was calculated from energy expended and energy intake as calculated from daily food and fluid consumption. Corrections were made for energy losses from feces, urine and sweat and for the blood samples drawn. Energy content of food, fluid, feces, urine and blood was determined by bomb calorimetry (IKA Germany). Energy content from sweat was calculated from total sweat urea, assuming that the remainder was negligible.

Sweat volume collected in the capsules was determined by capsule weight changes using a Mettler® analytical balance. Total sweat loss was determined from body weight change while accounting for fluid intake, urine production, blood volume loss, and respiratory water loss (Mitchel, 1972). Sweat urea content was determined enzymatically (urease method Boehringer (396346).

Nitrogen content of feces, blood and urine was determined by the chemoluminescence method (Antek Germany). Daily nitrogen losses were then calculated.

The quantitative contribution of CHO, fat and protein to total daily energy intake was calculated from daily intake records using a UCV computer coding system for Dutch foodstuffs (Hautvast, 1975, Boeyen, 1983). CHO and fat oxidation was calculated from non-protein respiratory quotient (R).

Protein oxidation was calculated from daily nitrogen losses in urine and sweat. From the data gathered the relative contribution of CHO, fat and protein to total 24 h energy metabolism as well as the relative contribution to energy metabolism during exercise was calculated.
Plasma volume changes were calculated from hemoglobin and hematocrit values according to Dill and Costill (1974). Hematocrit was determined in all blood samples. Hemoglobin was measured with the hemoglobin-cyanide method.

Statistics
Student's paired t-test was used to compare the data of the first standardized rest day to those of the following exercise and recovery days. For all analyses, the 0.05 level was used as the minimum level of confidence for statistical significance.

RESULTS

Energy
Energy intake (EI) from food and fluid was markedly constant during the whole experimental period despite the fact that exhausting cycling exercise was executed on days 4 and 5 of the experiment (Table 2) A small but not statistically significant increase in EI was found during the recovery day.

<table>
<thead>
<tr>
<th>Table 2</th>
<th>Energy intake and expenditure</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Day</td>
</tr>
<tr>
<td>Energy</td>
<td></td>
</tr>
<tr>
<td>Intake</td>
<td></td>
</tr>
<tr>
<td>19.4±1.1</td>
<td>19.3±0.8</td>
</tr>
<tr>
<td>Expenditure</td>
<td>16.1±0.3</td>
</tr>
</tbody>
</table>

(Mean values ± SEM (MJ.day⁻¹)).
Day 3 and 6 are resting days, day 4 and 5 are exercise days.
Statistical significance of differences with respect to the values on day 3 is indicated by ***p<0.001

Energy expenditure (EE) as measured in the respiration chamber was similar on both rest days and increased by more than 60% to 25 MJ/day as a result of exhaustive cycling. This level of EE is equivalent to EE levels previously observed during the Tour de France (Saris, 1988).
As a result of the constant EI but strongly increased EE, energy balance was -8 and -10 MJ negative during the exhaustive cycling days, 4 and 5 respectively (Fig 3).
**Nutrient intake**

CHO and protein intake levels - 52.5 en% and 12.5 en% respectively - were comparable to those observed during the Tour de France (Saris, 1988). The relative contribution of CHO, protein and fat to total EI also remained constant (Figs 4 and 5), except for the recovery day when CHO intake increased significantly to 70 en%.

The absolute amount of CHO and protein intake (g) increased during the recovery day as a result of increased food intake.

---

**Fig 3** Mean daily energy balance derived from total energy intake minus total energy expenditure. Corrections were made for energy losses in urine, feces, blood samples and sweat. Day 3 and 6 are resting days; 4 and 5 are exercise days. Vertical bars indicate 1 SEM (n=13). Statistical of differences with respect to the corresponding values on day 3 is indicated by ***P<0.001."
Fig 4  Mean daily CHO intake (□ g; □ en%) and CHO oxidized (□ g). The amount of CHO oxidized was calculated from non protein R. Vertical bars indicate 1 SEM (n=13). Statistical significance of differences with respect to the corresponding values on day 3 is indicated by *p < .05; **p < .01; ***p < .001.

Fig 5  Mean daily protein intake (□ g·kg⁻¹; □ en%) and protein oxidized (□ g·kg⁻¹). The amount of protein oxidized was calculated from nitrogen excretion in urine and sweat. Vertical bars indicate 1 SEM (n=13). Statistical significance of differences with respect to the corresponding values on day 3 is indicated by *p < .01.
Nutrient oxidation
The amount of CHO and protein oxidized have also been plotted in Figs 4 and 5. CHO, fat and protein oxidation were estimated from the calculated R and the measured nitrogen excretion urine and sweat. A comparison with the intake of these nutrients reveals that CHO intake was greater than CHO oxidation on both rest days, while oxidation exceeded intake on the first exercise day, was similar to intake on the second exercise day and significantly smaller during the recovery day. Protein oxidation as calculated from N loss in sweat and urine was less than protein intake on the first rest day. During the following exercise days and also on the recovery day protein oxidation increased significantly, almost balancing protein intake. The mean levels of oxidation and intake both exceeded 1.5 g·kg⁻¹BW. The daily relative contribution of CHO, protein and fat to energy expenditure is presented in Fig 6.

Fig 6  Relative contribution of nutrients to total energy expenditure calculated from non protein R and nitrogen excretion in urine and sweat. CHO = carbohydrate; F = fat; P = protein. Statistical significance of differences with respect to the values on day 3 is indicated by *p<.05; **p<.01; ***p<0.001.

During the rest day CHO had the largest contribution to energy metabolism (58.2±6.1%). This contribution decreased gradually and significantly on days 4 and 5 (51.4±3.1% and 40.6±3.4% respectively) whereas it remained significantly lower during the recovery day (55.3±4.0%) compared to day 3. Concurrently the contribution of fat to total energy supply gradually increased.
The percentual contribution of protein decreased significantly on the first exercise day from 9.5±0.7 to 7.1±0.5% and remained unchanged on the second exercise day (8.2±0.7%). A significant increase occurred during the recovery day, (14.6±1.3%) compared with day 3. Calculation of the relative contribution of CHO, fat and protein during the sustained exercise period on day 4 and 5 revealed a highly significant increase in fat and decrease in CHO metabolism on day 5 compared to day 4 (Table 3). The quantitative relative contribution depended largely on the level of energy expenditure so that the figures obtained at rest and during exercise are quite different. This difference is the most pronounced with respect to protein.

**Food consumption pattern**

Analysis of the daily consumption pattern shows that the contribution of main meals (breakfast, lunch, dinner) to total energy intake amounted 65.7% to 69.5% on the different experimental days. There was no significant difference between rest and exercise days. The in-between meal food consumption accounted for 30.5 to 34.3% of total energy intake. This eating pattern remained markedly constant throughout the experiment (Fig 7).

**Table 3 Substrate oxidation at rest and during the entire exercise period**

<table>
<thead>
<tr>
<th></th>
<th>Day 4</th>
<th>Day 5</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Rest</td>
<td>Ex</td>
</tr>
<tr>
<td>CHO</td>
<td>35.6±4.3</td>
<td>58.8±3.2</td>
</tr>
<tr>
<td>Fat</td>
<td>47.4±4.8</td>
<td>37.9±3.1</td>
</tr>
<tr>
<td>Prot</td>
<td>17.5±1.0</td>
<td>3.3±0.3</td>
</tr>
</tbody>
</table>

Relative contribution (%) of nutrients to energy exchange as calculated from non protein R and nitrogen excretion in urine and sweat.

Statistical significance of differences with respect to the values on day 4 is indicated by *p<0.05; **p<0.001.
Fig 7  Relative contribution of main meals and in-between meals to total daily energy intake.
B = breakfast; L = lunch; D = dinner . food intake during exercise.

**Fluids**
Daily fluid intake and loss at rest and during exercise periods is presented in Table 4. As a result of fluid intake during exercise, total fluid intake increased to more than 6 l/day. Fluid intake at rest remained about the same over the whole period (3 - 4 l/day). Fluid intake during exercise exceeded 3 l/day; sweat loss and respiratory fluid loss approximated 4 l/day (Table 4). Mean urine production increased on both exercise days, compared to the rest days.
Table 4  Fluid intake/loss

<table>
<thead>
<tr>
<th>Conditions</th>
<th>Day 3</th>
<th>Day 4</th>
<th>Day 5</th>
<th>Day 6</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Intake</td>
<td>loss</td>
<td>Intake</td>
<td>loss</td>
</tr>
<tr>
<td>REST</td>
<td>4.01±0.19</td>
<td>3.14±0.22</td>
<td>3.50±0.29</td>
<td>3.70±0.25</td>
</tr>
<tr>
<td>Urine</td>
<td>1.91±0.19</td>
<td>2.81±0.32</td>
<td>2.11±0.19</td>
<td>1.68±0.11</td>
</tr>
<tr>
<td>Feces</td>
<td>0.15*</td>
<td>0.15*</td>
<td>0.15*</td>
<td>0.15*</td>
</tr>
<tr>
<td>Resp. water</td>
<td>0.40*</td>
<td>0.32*</td>
<td>0.32*</td>
<td>0.40*</td>
</tr>
<tr>
<td>Sweat</td>
<td>0.50*</td>
<td>0.40*</td>
<td>0.40*</td>
<td>0.50*</td>
</tr>
<tr>
<td>EXERCISE</td>
<td>3.18±0.16</td>
<td>3.17±0.20</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Sweat</td>
<td>3.50±0.25</td>
<td>3.38±0.22</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Resp. water</td>
<td>0.55±0.02</td>
<td>0.52±0.02</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

Mean total 4.01 2.86 6.32 7.74 6.67 6.89 3.70 2.73

Mean values ± 1 SEM (n=13). Fluid losses from sweat and respiratory water was not calculated on the rest days (3 and 6). * mean daily loss under resting conditions taken from Weltzman (1980).

On both rest days plasma volume, as a reflection of the hydration status of the blood, showed a significant increase at 12:00 and 4:00 PM compared to 7:00 AM on the first day. This increase was not observed on either of the following exercise days. The increase during the recovery day was significantly greater than during the first rest day (Fig 8).

Body weight significantly decreased during the exercise days, and increased somewhat during the following 1 1/2 day of recovery (Table 5). However, final body weight was still significantly lower than initial body weight.

Table 5  Body weight changes of experimental subjects

<table>
<thead>
<tr>
<th></th>
<th>Day</th>
<th>Time</th>
<th>3</th>
<th>4</th>
<th>5</th>
<th>6</th>
<th>7</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>7:00 AM</td>
<td>73.7±2.2</td>
<td>73.4±2.3</td>
<td>72.4±2.2</td>
<td>72.4±2.1</td>
<td>72.8±2.2</td>
</tr>
<tr>
<td></td>
<td></td>
<td>4:00 PM</td>
<td>73.7±2.3</td>
<td>72.9±2.3</td>
<td>72.2±2.1</td>
<td>72.7±2.2</td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td>9:00 PM</td>
<td>74.7±2.3</td>
<td>73.4±2.2</td>
<td>73.1±2.2</td>
<td>73.5±2.2</td>
<td></td>
</tr>
</tbody>
</table>

Mean values ±SEM (kg; n=13) Significant changes with respect to initial values on day 3 are indicated by *p<0.01; **p<0.001.
Fig 8  Percentual changes in plasma volume at rest (day 3 and 6) and during exhaustive exercise (day 4 and 5). Plasma volume on day 3 at 7:00 AM was taken as the initial value. Statistical significance of differences with respect to the corresponding values on day 3 is indicated by *p<.05; **p<.01; ***p<0.001.

Discussion
Although a normal well balanced diet is often recommended as being sufficient to meet the needs under all circumstances, we recently described that consuming such a diet may become impossible during long lasting intensive competitions when energy expenditure exceeds 20 MJ/day (Brouns, 1986).
This may be of particular importance in situations where sustained competitive exercise is executed on consecutive days over a prolonged period of time, such as cycling the Tour de France (21 days). The present study was designed to analyse the effect of repeated highly intensive cycling exercise on food consumption, energy balance and fluid status of the body, when consuming a conventional but sport adapted CHO rich diet. The cyclists were instructed to refrain from intensive training or competition during the last 2 days prior to the stay in the laboratory and to ingest a normal but CHO rich diet (dietary advise was given by a dietician). During the first day in the laboratory, which was a preparation day, food intake and amount of exercise
was controlled. At least 60 en% CHO and a minimal protein intake of 1.2 g.kg⁻¹ were assured. This was done in order to realize a complete build-up of endogenous substrate pools and to minimize inter individual differences in pre-test metabolic status, as it is known that hormonal changes and substrate mobilization and utilization during exercise are influenced by the state of the organism prior to exercise. Involved in this state are nutrition, degree of training, plasma hormone levels and level of physical activity (Galbo, 1983). An illustration of the influence of the pre-test state is the finding that subjects who have reduced CHO depots due to intake of a diet low in CHO, to previous exercise, or to a combination of both - combust more fat and protein and less CHO during exercise compared to subjects with adequate CHO intake and sufficient rest prior to the test (Galbo, 1979, 1981; Golinick, 1981; Lemon, 1981; Maughan, 1978). After the controlled preparation phase, the final study in the respiration chamber was started.

The results show that total energy intake exceeds total energy output whenever the cyclists have an active rest day. When competitive cycling comparable to a hard day in the Tour de France (Sans, 1988) was simulated, the cyclists showed a marked constancy in energy intake. Although they were all aware of the importance of adequate food intake, they were not able to ingest sufficient food to compensate for the increased energy expenditure. Because the set-up of the experiment was such that food and drink was available ad libitum within handreach throughout the day the reason for this maladaptation of energy consumption most probably is due to a suppressed appetite and/or intolerance of the gastro-intestinal tract to bear and digest large bulk loads of food. Moreover, the cyclists indicated that the exercise was too intensive to be able to ingest large amounts of food and/or were too exhausted to eat. As a result there was no change in quantitative food intake during the both exercise days compared to the foregoing standardized active rest day.

The nibbling eating pattern as described by Kirsch and Von Ameln (1981) was also present in this study. In-between meal snacking represented 30 - 34% of the total energy intake in these cyclists, the majority of which took place in the evening (Fig 7). This in-between meal energy consumption is quantitatively comparable to that found in other competitive sport events (De Wijn, 1981; Van Erp-Baart, 1988) and may be explained by a continuous hunger for small, CHO rich snacks.

Although we expected that energy consumption during the evening would be increased after intensive cycling, in order to compensate for the increased energy expenditure, we did not see such a compensation. There tended to be what might be called a 'voluntary energy depletion' which finally led to a slightly but not significantly increased energy intake and a significantly increased CHO intake (70 en%) during the following recovery day, indicating a preference for CHO rich foods. As a result, energy balance was positive on
the active rest days but significantly negative during the exercise exhaustion days (Fig 2). CHO had a large contribution to energy production on the first cycling day most probably, because exercise intensity was greater than 50% of \( \dot{V}O_2 \) max. On that day CHO intake was insufficient to meet the needs, as indicated by the difference between CHO intake and calculated CHO oxidation (Fig 4). This relatively low CHO intake can to a large extent be explained by the negative energy balance. The reason is that CHO contributed the largest fraction of total energy intake whereas on the other hand CHO also contributed the major fraction in energy expenditure during exercise. As a result of this 'CHO imbalance', endogenous CHO must have contributed considerably to energy production. In the case of endogenous CHO depletion the body will adapt with enhanced fat mobilization and gluconeogenesis (Newsholme, 1983; Rennie, 1977). It has been shown that enhanced availability of free fatty acids increases the fraction of oxidized fat and decreases CHO oxidation because both substrates act competitively at the site of transport across the muscle cell membrane and subsequent oxidation (Randia, 1963; Felber, 1964; Büber, 1968; Gomez, 1972; Rennie, 1976, 1977). This may explain the increased fat oxidation on the second exercise day and consequently the smaller CHO contribution (Table 3, Fig 6).

Protein intake per kg body weight was more than adequate according to RDA standards (1.7 vs 0.8 g kg\(^{-1}\) RDA). However, the calculated fraction of protein involved in energy exchange, indicates that during both exercise days and the following recovery day, when protein oxidation was significantly increased (Fig 5), protein intake almost equalled protein loss.

In this situation of stressful exercise and negative energy balance, protein oxidation per kg body weight was far in excess of the amount of protein intake advised by the National Research Council (RDA). The amount of protein oxidized indicates that even 1.5 g kg\(^{-1}\) in these circumstances may not be sufficient to meet the needs. (This will be discussed more in detail in Chapter IV). Increased protein degradation as an indication of energy depletion and catabolic status of the body is supported by Lemon and Mullin (1980) who showed that CHO depleted subjects had a significant increase in protein breakdown compared to CHO loaded subjects. Recently Lemon (1987) suggested a protein requirement of 1.2 to 1.6 g kg\(^{-1}\) day.

Fluid intake appeared to be sufficient to meet the requirements both during rest and exercise days. The fact that plasma volume increased during day 3 may be explained by changes in blood composition due to absorption of nutrients, electrolytes and water during the day. That this did not occur during the exercise days may be indicative of the smaller fluid overload (fluid intake - fluid loss) due to severe sweating on these days, compared to both rest days, or to fluid shifts within the body. The fact that urine volumes increased during the exercise day indicates a stimulus to water elimination from the body rather
than to retain it. Moreover, some of the subjects had to urinate during the cycling sessions. Indicative of an adequate fluid supply also during exercise is further the observation that plasma volume did not fall below the initial 'zero' value as measured at 7:00 AM after an overnight fast on the first standardized rest day. It has to be kept in mind however, that plasma volume due to changes in osmolality may not necessarily reflect the tissue hydration status. It may thus well be that tissue dehydration has taken place at cost of maintaining a normal plasma volume especially during day 4 when fluid loss exceeded fluid intake by approximately one liter. During the recovery day, plasma volume increased significantly above the values of the first rest day, which may be explained by sodium and water retention which has been shown to occur after periods of severe prolonged sweating (Costill, 1977).

In summary the results lead to the following conclusions:
- When prolonged intensive cycling increases energy expenditure to levels above a certain threshold (probably around 20 MJ), athletes are unable to consume enough conventional food to provide adequate energy to compensate for the increased energy expenditure.
- A sport adjusted CHO rich diet (>60 en% CHO) is in itself no guarantee that CHO intake will be sufficient during days of hard, long lasting exercise.
- Drinking 0.75 - 1 l.h.\(^{-1}\) is sufficient to maintain a normal plasma volume during prolonged intensive cycling under situations as in this study (ambient temperature 20 ± 2°C, relative humidity 65-75%).
- Protein requirement in the exercise circumstances described is greater than 1.5 g.kg\(^{-1}\) body weight/day.
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CHAPTER III: EATING, DRINKING AND CYCLING, A CONTROLLED TOUR DE FRANCE SIMULATION STUDY
PART 2, THE EFFECT OF DIET MANIPULATION


Introduction
Whenever energy requirement in athletes increases to levels above 20 MJ/day, the athlete is faced with the problem of how to ingest and digest such a large quantity of food when long lasting exercise has to be executed at the same time. Kirsch (1980) described a spontaneous nibbling eating pattern of athletes who, in such circumstances, tend to eat more or less continuously throughout the day. Practical experience shows that especially foods and liquids are selected that are sweet in taste, high in energy and are convenient. Consequently it is found that in-between meals composed of such foods and drinks may make up more than 30% of total energy intake (De Wijn 1980; Van Erp-Baart, 1988; this study, Chapter II). One of the main problems of athletes during competition days seems to be the intolerance of the gastro-intestinal tract to take up and digest large amounts of food whenever intensive exercise is executed at the same time. Apart from the exercise induced changes in appetite (Brouns, 1986) it is clearly indicated from practice that athletes must rely on easily digestible and energy dense foods that are low in dietary fiber, for the following reasons:
- The amount of feces produced from a normal diet would require the cyclist to defecate several times during the day and consequently to interrupt exercise.
- Large volumes of food may interfere with fluid intake which in itself may be exceptionally high during hot days (more than 10 l/day, Saris, 1988).

As a result the athlete will either undereat when only a conventional diet is available during such competition days (Chapter II) or will maintain energy balance when a combination of normal foods and carbohydrate (CHO) rich liquids is supplied as was observed during the Tour de France (Saris, 1988). In order to investigate more specifically the influence of such dietary practices on food and fluid intake, energy balance, fluid status and nutrient utilization during repeated exhausting cycling, it was decided to do a controlled laboratory study in which the effects of a normal CHO rich diet (N) (Chapter II) were compared to those of the same diet, but supplemented with a concentrated liquid CHO beverage.

The reason for CHO supplementation as diet intervention was two fold:
1) Depletion of CHO stores in the body is related to physical exhaustion (Bergström, 1967, 1967a; Sherman, 1984, Saltin, 1967) while increased CHO
intake may delay exhaustion (Coyle, 1983; Edwards, 1984; Ivy, 1983).

2) A survey among professional cyclists competing in the Tour de France showed that liquid CHO supplements had the largest relative contribution to total CHO intake (Saris, 1988).

Although there is a wide variety of CHO sources, it was specifically decided to select a solution composed of mainly maltodextrin (Mf) and a solution with a high fructose fraction (FM). Mf was selected because the use of maltodextrin may be of advantage for the athlete by means of maximizing CHO intake and absorption, while minimizing gastro-intestinal distress (Brouns, 1987). FM was selected for the following reasons. The effect of fructose on insulin secretion is minimal, which allows for a better stimulation of lipolytic activity under exercise circumstances compared to glucose alone which has a strong influence on insulin secretion. The second reason was to compose a CHO solution with a 50% contribution of both glucose and fructose but with a lower osmolality as would be derived from eucaloric sucrose.

**Experimental design, analyses and statistics**

The experimental set-up used during the diet manipulation trial is exactly the same as has been described in Chapter II for the normal diet (N) study. The same holds for the analysis of samples and the statistics used to analyze the data. Therefore only the differences due to diet manipulation, not described before, will be outlined here.

The total group of 13 subjects receiving N as described in part I, chapter II, was divided into two sub-groups for diet manipulation. Six subjects were supplemented with an experimental high maltodextrin, low fructose beverage (Perform®, Wander Ltd) (Mf). The seven other subjects were supplemented with another experimental beverage, composed of 50% free fructose and 50% maltodextrin (FM). Subjects were randomly selected for either Mf of FM treatment as was the order of treatment (normal diet or manipulated diet). The composition of the beverages used is presented in Table 1.

**Table 1 Composition of beverages**

<table>
<thead>
<tr>
<th>Beverage</th>
<th>Maltodextrin g.l⁻¹</th>
<th>Fructose g.l⁻¹</th>
<th>K⁺ mmol</th>
<th>Osmolality mosm</th>
<th>pH</th>
<th>Energy content kJ.l⁻¹</th>
</tr>
</thead>
<tbody>
<tr>
<td>Placebo</td>
<td>0</td>
<td>0</td>
<td>8.5</td>
<td>106</td>
<td>4.38</td>
<td>-</td>
</tr>
<tr>
<td>Mf</td>
<td>150</td>
<td>33</td>
<td>8.5</td>
<td>390</td>
<td>4.45</td>
<td>3158</td>
</tr>
<tr>
<td>FM</td>
<td>90</td>
<td>93</td>
<td>8.5</td>
<td>788</td>
<td>4.52</td>
<td>3158</td>
</tr>
</tbody>
</table>

*Placebo drink was supplied during N treatment in equal quantities and at the same times as the*
beverages in Mf and FM treatment. The placebo was flavored with citrus powder, sweetened with saccharin and cyclamate and artificially coloured. In all beverages the same vitamin-mineral mix was added.

The supplement was made available for intake together with normal food during cycling and during the evening, as follows: standardized resting days 500 ml during the morning, afternoon and evening (total supply 1500 ml). Exercise days during cycling ad libitum and in the evening, after dinner, 1000 ml. The cyclists in the N treatment were supplied with a placebo drink (see Table 1), in order to rule out psychological effects due to the drink.

Results

Energy

The data on energy intake (EI) and energy expenditure (EE) are presented in Table 2.

Table 2   Energy intake (EI) and expenditure (EE)

<table>
<thead>
<tr>
<th>Day</th>
<th>Treatment</th>
<th>3</th>
<th>4</th>
<th>5</th>
<th>6</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>EI Mf</td>
<td>23.5±1.0</td>
<td>29.2±0.9*</td>
<td>27.0±1.0**</td>
<td>20.3±1.6</td>
</tr>
<tr>
<td></td>
<td>EE Mf</td>
<td>16.5±0.4</td>
<td>26.6±1.1</td>
<td>25.8±1.2</td>
<td>16.2±0.5</td>
</tr>
<tr>
<td></td>
<td>EI FM</td>
<td>22.2±1.4**</td>
<td>26.3±0.9**</td>
<td>25.3±1.5**</td>
<td>18.9±1.6</td>
</tr>
<tr>
<td></td>
<td>EE FM</td>
<td>16.1±0.5</td>
<td>26.9±1.0</td>
<td>26.3±1.1</td>
<td>16.0±0.6</td>
</tr>
</tbody>
</table>

Mean values ± SEM are given in MJ. For Mf group n=6, for FM group n=7. For composition of the beverages see Table 1. EI was determined by weighed food intake, EE was determined by indirect calorimetry. Asterisks indicate a significant difference with respect to values obtained with con-ventional diet (N). (Each subject is his own control. For N values see Table 2, Chapter II), *p<0.05; **p<0.01.

When the values of diet manipulation were compared to those of N (each subject was his own control) there were large statistically significant differences for EI, while there were no differences in EE between the treatment groups. In Mf, mean EI was always greater than mean EE, despite the fact that EE increased to levels far above 20 MJ on both exercise exhaustion days. In contrast to this, in cyclists with FM treatment EI was smaller than EE on both exercise days. EI was significantly higher (p<0.05) in Mf compared to FM on the first exercise exhaustion day but not on the second exercise day. As a result of the increased EI in Mf, the subjects of this groups remained in
energy balance, whereas in the FM treatment subjects were not. In this group energy balance was negative during the sustained cycling days, however, to a lesser extent than during N (Fig 1).

**Fig 1** Mean daily energy balance ( E) derived from total energy intake minus total energy expenditure. Corrections were made for energy losses in urine, feces, blood and sweat. Day 3 and 6 are resting days, 4 and 5 are exercise days. Vertical bars indicate 1 SEM. --- conventional food group (N); n=13, --- MI group (n=8), --- FM group (n=7). Indicates statistical significance of differences compared to the initial value on day 3; *p<0.05; **p<0.01; ***p<0.001. *Indicates a statistically significant difference with respect to comparative data of groups N (each subject is his own control). **p<0.01; ***p<0.001.

**Nutrient Intake**
In contrast to N, dietary manipulation caused a significant shift in the relative contribution of macro-nutrients to total energy intake. During the exercise exhaustion days CHO intake increased significantly to a mean level of 80
en% in Mf and 77 en% in FM respectively. The CHO intake is shown in Fig 2. As a result of the diet manipulation CHO intake increased significantly on all days, reaching a mean maximum value of 1300 g and 1180 g in Mf and FM respectively. As a consequence fat intake dropped significantly to 15 en% and protein intake to 7.5 en% in both supplemented groups (Table 3).

### Table 3 Relative contribution of nutrients to energy intake

<table>
<thead>
<tr>
<th>Food component</th>
<th>Diet group</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>N</td>
</tr>
<tr>
<td>CHO</td>
<td></td>
</tr>
<tr>
<td>Day 3</td>
<td>61.8±1.7</td>
</tr>
<tr>
<td>Day 4</td>
<td>62.9±1.3</td>
</tr>
<tr>
<td>Day 5</td>
<td>62.4±1.6</td>
</tr>
<tr>
<td>Day 6</td>
<td>62.7±1.6</td>
</tr>
<tr>
<td>FAT</td>
<td></td>
</tr>
<tr>
<td>Day 3</td>
<td>25.7±1.3</td>
</tr>
<tr>
<td>Day 4</td>
<td>24.7±1.0</td>
</tr>
<tr>
<td>Day 5</td>
<td>25.6±1.3</td>
</tr>
<tr>
<td>Day 6</td>
<td>24.5±1.2</td>
</tr>
<tr>
<td>PROTEIN</td>
<td></td>
</tr>
<tr>
<td>Day 3</td>
<td>12.5±0.6</td>
</tr>
<tr>
<td>Day 4</td>
<td>12.3±0.6</td>
</tr>
<tr>
<td>Day 5</td>
<td>12.1±0.7</td>
</tr>
<tr>
<td>Day 6</td>
<td>12.8±0.4</td>
</tr>
</tbody>
</table>

Mean values ± SEM are given in en%. N = normal diet (n=13). Mf = diet supplemented with high maltodextrin/low fructose beverage (n=6). FM = diet supplemented with fructose/maltodextrin beverage (n=7). Days 3 and 6 are resting days; days 4 and 5 are exercise days. Statistical significance with respect to the corresponding initial values on day 3 is indicated by *p<0.05; **p<0.01; ***p<0.001. Statistical significance of groups Mf and FM with respect to corresponding data of group N (each subject is his own control) is indicated by "p<0.005; ""p<0.01; """"p<0.001."
Fig 2  Mean daily CHO intake (□, g) and CHO oxidized (□, g). The amount of CHO oxidized was calculated from nonprotein R. Vertical bars represent 1 SEM. For the N treatment group n=13, for the M treatment group n=6, for the FM treatment group n=7. Statistical significance with respect to the initial value on day 3 is indicated by *p<0.05; **p<0.01. Statistical significance of groups M and FM with respect to comparative data of group N (each subject is his own control) is indicated by *p<0.05; **p<0.01; ***p<0.001.

The protein intake per kg body weight, is presented in Fig 3. Although there was a significant reduction in the relative contribution of protein intake to total energy intake, it can be seen that the quantitative protein intake remained more or less constant during the different treatments. The largest intake occurred during N whereas the supplemented groups had lower intakes, but still markedly above the RDA of 0.8 g.kg\(^{-1}\)BW (mean lowest value 1.4 g.kg\(^{-1}\) on day 4 in M), and above the preset minimal intake of 1.2 g.kg\(^{-1}\)BW. There were no differences in intake in the different treatments between the first and last standardized rest day. However, when N was compared to the supplemented diets, it becomes apparent that fat and protein intake during the rest days is larger in N.
Fig 3  Mean daily protein intake (□  g.kg⁻¹) and protein oxidized (■  g.kg¹). The amount of protein oxidized was calculated from nitrogen excretion in urine and sweat. Vertical bars indicate 1 SE.M. N = conventional diet (n=13); Mf = diet supplemented with high maltodextrin/low fructose beverage (n=6); FM = diet supplemented with high fructose/maltodextrin beverage (n=7). For exact composition of diets and beverages see text. Day 3 and 6 are resting days, 4 and 5 are exercise days. Statistical significance with respect to the initial values on day 3 is indicated by * p<0.05; ** p<0.01; *** p<0.001. Statistical significance of groups Mf and FM with respect to comparative data of group N (each subject is his own control) is indicated by * p<0.05; ** p<0.01; *** p<0.001.

Nutrient oxidation
The absolute amounts of CHO, fat and protein oxidized, are also graphically presented in Fig 2 and 3. With respect to CHO it can be seen that when oxidation is compared to intake, Mf treatment resulted in a positive 'CHO balance' during the entire experiment. This is in contrast to N and FM which resulted in negative CHO balance due to increased CHO oxidation. The calculated relative contribution of the nutrients to total daily energy metabolism is presented in Fig 4.
Fig 4. Mean relative contribution of CHO, fat and protein to total energy expenditure as calculated from non protein R and nitrogen excretion. C = carbohydrate, F = fat, P = protein. The amount of C and F oxidized was calculated from non protein R. The amount of protein oxidized was calculated from nitrogen excretion in urine and sweat. Vertical bars indicate 1 SEM. Statistical significance with respect to the initial values on day 0 is indicated by ."p<0.05; "p<0.01; ""p<0.001. Statistical significance of groups Mf and FM with respect to comparative data of group N (each subject is his own control) is indicated by "p<0.05; "p<0.01; ""p<0.001.

In contrast to N the relative contribution from fat to energy metabolism in the supplemented groups decreased during the first exercise day, followed by a stepwise increase during the second exercise day and following recovery day, however, at a lower level than in N. Consequently the oxidized fraction of CHO in both groups was larger. The lowest contribution from fat to energy metabolism, together with the highest CHO contribution, occurred in the 'high' fructose group, during both rest and exercise days. Compared to N this was highly significant on all days.

Protein oxidation remained constant in Mf, despite sustained exercise. This is in contrast to N which showed a large significant increase during exercise and recovery days. In FM the amount of protein oxidized tended to increase during the first exercise day, then increased significantly during the second exercise day and almost returned to the initial level during the recovery day. Compared to N, exercise caused less protein degradation probably as a result of diet manipulation, which became significant in FM and tended towards significance in Mf. The relative contribution of protein to overall energy metabolism as calculated from nitrogen losses in sweat and urine decreased significantly during both exercise days in Mf, but in FM only during the first exercise day. Compared to N the relative contribution was smaller,
except on the first rest day. During the recovery day all 3 treatment groups showed an increased protein oxidation compared to the pre-exercise rest day. However, this increase was only significant in N. Compared to N, FM had a significantly lower value on this day while Mf tended to be significantly lower (p<.07).

**Food consumption pattern**

The food consumption pattern presented in Fig 5 shows that the supplementation of CHO rich beverages during exercise caused a significant increase in energy intake, being almost entirely responsible for the increased total energy intake compared to N. In general, energy intake derived from the three main meals remained constant.
Fig 5  Relative contribution of main meals and in-between meals to total daily energy intake. R = breakfast; L = lunch; D = dinner; # food intake during exercise. Statistical significance of groups MI and FM with respect to comparative data of group N (each subject is his own control) is indicated by *p<0.05; **p<0.01; ***p<0.001.

In FM breakfast and dinner were significantly smaller during the recovery day than during the same day in the control experiment (N). This resulted in a lower total energy intake on this day. After exhaustive exercise in between meal snacking sometimes was reduced before dinner and increased in the evening after dinner in both CHO supplemented groups.
Fluids
Fluid intakes and losses in Mf and FM are represented in Table 4.

<table>
<thead>
<tr>
<th>Table 4 Fluid intake and loss in Mf (a) and FM (b) treatment</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Condition</strong></td>
</tr>
<tr>
<td>----------------</td>
</tr>
<tr>
<td>Mf</td>
</tr>
<tr>
<td>REST</td>
</tr>
<tr>
<td>Urine</td>
</tr>
<tr>
<td>Faeces</td>
</tr>
<tr>
<td>Resp water</td>
</tr>
<tr>
<td>Sweat</td>
</tr>
<tr>
<td>EXERCISE</td>
</tr>
<tr>
<td>Sweat</td>
</tr>
<tr>
<td>Resp water</td>
</tr>
<tr>
<td>Mean total</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Condition</th>
<th><strong>3</strong></th>
<th><strong>4</strong></th>
<th><strong>5</strong></th>
<th><strong>6</strong></th>
</tr>
</thead>
<tbody>
<tr>
<td>FM</td>
<td>Intake</td>
<td>loss</td>
<td>Intake</td>
<td>loss</td>
</tr>
<tr>
<td>REST</td>
<td>3.60 ± 0.24</td>
<td>3.72 ± 0.51</td>
<td>2.55 ± 0.26</td>
<td>3.70 ± 0.37</td>
</tr>
<tr>
<td>Urine</td>
<td>1.74 ± 0.17</td>
<td>2.69 ± 0.42</td>
<td>2.35 ± 0.42</td>
<td>1.46 ± 0.16</td>
</tr>
<tr>
<td>Faeces</td>
<td>0.15*</td>
<td>0.15*</td>
<td>0.15*</td>
<td>0.15*</td>
</tr>
<tr>
<td>Resp water</td>
<td>0.40*</td>
<td>0.32*</td>
<td>0.32*</td>
<td>0.40*</td>
</tr>
<tr>
<td>Sweat</td>
<td>0.50*</td>
<td>0.40*</td>
<td>0.40*</td>
<td>0.50*</td>
</tr>
<tr>
<td>EXERCISE</td>
<td></td>
<td></td>
<td>3.03 ± 0.31</td>
<td>3.53 ± 0.24</td>
</tr>
<tr>
<td>Sweat</td>
<td></td>
<td></td>
<td>3.40 ± 0.51</td>
<td>3.54 ± 0.60</td>
</tr>
<tr>
<td>Resp water</td>
<td></td>
<td></td>
<td>0.56 ± 0.02</td>
<td>0.56 ± 0.04</td>
</tr>
<tr>
<td>Mean total</td>
<td>3.60</td>
<td>2.79</td>
<td>6.75</td>
<td>7.52</td>
</tr>
</tbody>
</table>

Mean values ± 1 SEM (Mf n=6; FM n=7)
* Mean daily loss under resting conditions, taken from Weltman (1980).
As in N, fluid intakes were larger than the calculated fluid losses during both rest days. During both sustained cycling days losses via urine, sweat and respiratory water loss exceeded fluid intake by approximately one liter. Urine losses increased during exercise days and were not different from N. Respiratory water loss during exercise was remarkably constant reaching a mean level of around 550 ml or approximately 110 ml/hr. In general plasma volume showed a similar response pattern to food and fluid intake and to exercise as described for N (Fig 6 and Fig 8 of Part I, Chapter II).

Fig 6 Relative changes in plasma volume at rest (day 3 and 6) and during exhaustive exercise days (day 4 and 5) — Mf treatment group; —— FM treatment group. Vertical bars indicate 1 SEM. There was no statistically significant difference between the treatment groups.

Although Mf subjects tended to have decreased plasma volumes below the initial level on exercise days compared to in N treatment, this was not statistically significant. Body weight remained markedly stable during the experiment in
both Mf and FM. This is in contrast to in N where a significant body weight reduction occurred which was not corrected during the studied recovery period. Although in FM as well as in Mf body weight decreased at the same measuring points during exercise, there was a tendency to significantly smaller weight changes than those measured during N. In contrast to N the subjects in Mf and FM restored their individual weight on a daily basis, by 9:00 PM. (Table 5).

### Table 5 Body weight changes of experimental subjects

<table>
<thead>
<tr>
<th>Time</th>
<th>Day</th>
<th>3</th>
<th>4</th>
<th>5</th>
<th>6</th>
</tr>
</thead>
<tbody>
<tr>
<td>7 AM</td>
<td>Mf</td>
<td>75.5±0.4</td>
<td>75.7±0.4</td>
<td>75.3±0.4*</td>
<td>75.3±0.3</td>
</tr>
<tr>
<td></td>
<td>FM</td>
<td>71.7±0.3</td>
<td>71.4±0.2</td>
<td>70.6±0.2</td>
<td>70.7±0.2</td>
</tr>
<tr>
<td>4 PM</td>
<td>Mf</td>
<td>75.5±0.4</td>
<td>74.7±0.4*</td>
<td>74.7±0.4</td>
<td>75.2±0.4</td>
</tr>
<tr>
<td></td>
<td>FM</td>
<td>71.4±0.2</td>
<td>70.9±0.2</td>
<td>70.2±0.2</td>
<td>70.9±0.2</td>
</tr>
<tr>
<td>9 PM</td>
<td>Mf</td>
<td>76.1±0.4*</td>
<td>75.8±0.4</td>
<td>76.0±0.4*</td>
<td>76.0±0.4</td>
</tr>
<tr>
<td></td>
<td>FM</td>
<td>72.1±0.2*</td>
<td>71.7±0.2</td>
<td>71.5±0.2</td>
<td>71.8±0.2</td>
</tr>
</tbody>
</table>

Mean values ± SEM (kg); Mf n=6; FM n=7.

Changes with respect to initial values on day 3 are indicated by dots, *p<0.05. Significance of changes with respect to values found when on a normal diet are indicated by asterisks, *p<0.05.

### Discussion

The present study was designed to analyse the effect of diet manipulation on nutritional indices during repeated days of highly intensive cycling and to compare these effects to those present after a supply of a normally composed CHO rich diet (N). The diet manipulation consisted of ad libitum supplementation of liquid CHO concentrates, along with the normal CHO rich diet. The parameters specifically of interest were energy intake, expenditure and balance, nutrient utilization and fluid status.

The results of this study show that the subjects of both Mf and FM increased their daily EI significantly more during days of intensive cycling than under M treatment (Table 2). However, only the Mf group was able to maintain energy balance. FM subjects did not increase their intakes sufficiently to bridge the deficit caused by exercise. The fact that Mf subjects were able to increase their EI was clearly the result of the ad libitum supplementation with a beverage high in maltodextrin and low in fructose. The same subjects were not able to reach energy balance when using a conventional CHO rich diet. Furthermore, the FM group showed that a beverage high in fructose led to lower mean intakes compared to one with a small fructose fraction. This can
partially be explained by the fact that a 20% FM solution was not always tolerated by the subjects. A high osmolality (788 mosm) together with the extreme sweetness of the drink, due to fructose, caused gastric distress and palatability. Although a reduction of the CHO concentration to 10% during exercise increased palatability, this did not necessarily lead to the intake of larger volumes to compensate for the reduced EI.

As a result of the supplementation, total CHO intake increased significantly, on the first exercise day somewhat more in Mf (mean max. intake 1302±549 g equal to 17.5±1.0 g.kg. 1 BW), than in FM (1059±608 g equal to 14.9±1.0 g.kg. 1 BW). Despite these high CHO intakes none of the cyclists reacted with (osmotic) diarrhea caused by malabsorption. This was especially surprising in FM since it has been described that free fructose solutions introducing more than 50 g of fructose in the gastro-intestinal tract cause diarrhea and gastric distress (Andersson, 1978; Dubach, 1969) also during exercise (Levine, 1983). During the day of the highest CHO intake the FM subjects derived 566±29 g from the supplement, half of which was free fructose. The discrepancy with data from literature may be caused by the additional presence of glucose or by differences in training status.

The relative contribution of CHO intake to total EI, 80 en% and 77 en% in Mf and FM respectively, is far above intakes normally advised (50-65 en%). Some studies showed that CHO intake leading to elevated blood glucose levels stimulates CHO metabolism and inhibits fat metabolism, finally resulting in increased degradation rates of endogenous CHO stores (Hargreaves, 1984; Foster, 1979). With respect to this, the question is raised whether or not CHO supply can match the increased utilization of CHO. In the present study it is shown that Mf subjects were able to remain in positive 'CHO balance'. CHO intakes were always larger than the calculated CHO oxidation, which must have led to sparing of endogenous CHO stores. This is in contrast to N where 'CHO balance' was negative.

Although FM subjects also significantly increased their CHO intake, they were not able to completely compensate for the increased oxidation. Surprisingly, the treatment with the high fructose moiety in the drink showed the largest increase in CHO oxidation and decrease in fat oxidation, as calculated from non protein R. Although fructose is insulin independent and may therefore have a smaller effect on lipolytic activity than glucose, allowing for a larger energy exchange from fat, we did not observe such an effect. This observation is supported by the recent work of Tappy et al (1987) and contradicts the general belief that fructose may lead to a sparing of endogenous CHO by enhancing fat metabolism.

Fat and protein intake, in absolute quantities and relative to total EI, decreased as a consequence of the high CHO intake; in the N group intake was remarkably constant. The reduction in protein and fat intake was entirely
caused by the intake of the protein- and fat-free energy supplement and the reduced intake of normal foodstuffs containing fat and protein, especially during the cycling hours. Interesting to note is that this reduced intake of normal foodstuffs remained present during the recovery day in contrast to the N group, where food intake increased. Probably the subjects undergoing diet intervention were less stimulated to increase food intake during recovery because the exercise induced deficits were smaller.

A value of 7.5 en% for protein intake may sound alarming. However this should be interpreted with respect to the level of daily energy expenditure. In fact the mean lowest absolute daily intake within this value of 7.5 en% still was 1.4 g.kg\(^{-1}\).

Protein utilization in energy metabolism was significantly increased in N but not in Mf. It is also known that CHO depletion leads to catabolic stress which, through a variety of biochemical changes, leads to increased protein degradation in order to supply precursors for gluconeogenesis (Lemon, 1980; Newsholme, 1981; Felig, 1971, 1971a; Lemon, 1981). The response of the N subjects being in negative energy- and ‘CHO balance’ confirms these findings. Mf subjects, remaining in positive energy- and ‘CHO balance’ did not show an increased protein oxidation which further confirms that the same type of exercise, performed with the same intensity, even with a significantly longer performance time during the final 90% Wmax load (see chapter V), causes less catabolism stress when energy intake and especially CHO intake meets the requirements.

The fact that FM subjects had a significantly increased protein oxidation on the second exercise day but not during the first may be explained as follows. The subjects under this treatment were almost in balance with respect to CHO intake and CHO oxidation. However, their energy balance was negative on both days. Initially the high CHO intake may have counter-regulated the biochemical factors inducing protein degradation but when energy balance remained negative for a longer period of time, protein catabolism must have been initiated. The fact that nitrogen losses remained significantly increased during the next 36 hours of recovery in N indicates that biochemical disturbance was more pronounced in these subjects in contrast to that in subjects with CHO supplementation. Recovery of a normal metabolic status may have been enhanced under this condition.

Some observations may indicate that fluid intakes have been sufficient to compensate for the losses during all days. Increased urine production during the exercise days indicates an increased drive to eliminate water from the body rather than to retain it.

The reason why this is so, while at the same moment plasma volumes are decreased compared to rest days, remains obscure. One possibility may be
that urine production is increased due to the potassium content of the drink. Nielsen (1986) observed a diuretic and natriuretic effect of increased plasma potassium after rehydration with a high potassium drink, in spite of a low plasma volume. They could not explain this effect. The fact that plasma volume tends to decrease more in Mf cannot be explained by a fluid shift towards the splanchnic bed caused by the osmolality of the drink (380 mosm) since osmolality of FM was higher (788 mosm). Electrolyte influences have to be ruled out also since electrolyte content in both drinks was identical. The fast restoration of body weight further indicates that fluid loss and substrate loss must have been minimal in Mf and FM compared to N subjects who, even after 36 h of recovery, did not regain their initial body weights. This finding together with a better maintenance of energy balance, CHO balance and lower protein oxidation indicate a less pronounced catabolic state and a better recovery in the diet manipulated situation as compared to the situation in which a normal CHO rich diet was fed.

Summarizing the results of this study lead to the following conclusions.

- Cyclists are able to maintain energy balance when energy expenditure levels exceed 20 MJ by using concentrated CHO solutions in addition to a normal CHO rich diet. In contrast, by using a normal CHO rich diet alone energy balance cannot be maintained in such a situation.
- The ad libitum supply of 20 and 10% CHO solutions during cycling as described does not impair fluid intake nor overall fluid status of the body compared to water intake.
- The intake of substantial amounts of a CHO solution during exercise causes an increased CHO-, a decreased fat- and a decreased protein oxidation as compared to the situation in which a normal CHO rich diet alone is eaten.
- The recommended level of protein intake for endurance athletes performing highly intensive sustained exercise, consuming a CHO rich diet and being in positive energy balance most probably is in the range of 1.5 to 1.8 g.kg⁻¹ body weight.
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CHAPTER IV: THE EFFECT OF DIET MANIPULATION AND REPEATED SUSTAINED EXERCISE ON NITROGEN BALANCE, A CONTROLLED TOUR DE FRANCE SIMULATION STUDY, PART 3.


Introduction
It is known that the dietary energy level has a direct effect on nitrogen balance. A reduction of energy intake normally is followed by increased nitrogen losses from the body (Inoue, 1963; Galloway, 1975; Garza, 1976). Starvation leads to a depletion of endogenous carbohydrate (CHO) stores, which will result in increased amino acid oxidation and enhanced gluconeogenesis from alanine (Felig, 1969, 1970; Chang, 1978). This increased protein oxidation will last until the body adapts towards enhanced utilization of fat and ketones (Felig, 1969). It has also been observed that prolonged exercise may influence nitrogen balance. A number of studies have shown that protein is used as a source of energy during sustained physical exercise (Dohm, 1980; Cerny, 1975; Decombaz, 1978; Haralambie, 1975). Recently Lemon and Mullin (1980) observed that glycogen depletion enhances the rate of protein degradation and nitrogen loss from the body during prolonged exercise. In this respect it can be stated that a certain analogy exists between the initial phase of starvation and prolonged exercise. In both situations local glycogen stores become depleted and protein degradation will be increased. Because energy balance, glycogen levels, as well as the blood glucose level (Long, 1976) may influence the extent to which protein can serve as an energy substrate, it was hypothesized that diet manipulation in the exercising athlete has a substantial influence on protein degradation during exercise. Therefore it was decided to study nitrogen balance on rest days and on days of prolonged intensive physical exercise in highly trained subjects. The main purpose of this study was to evaluate the effect of diet manipulation during exhausting physical exercise on nitrogen balance.

Experimental design, analyses and statistics
Experimental set up, analyses and statistics used are described in detail in Chapter II and III.

Nitrogen balance was calculated according to the formula

\[ N \text{ balance} = N \text{ in} - N \text{ out} \]
where \(N_{\text{in}}\) = nitrogen intake from foods and fluids
\(N_{\text{out}}\) = nitrogen loss from the body by urine, feces, sweat and blood samples taken.

**Results**

The results concerning energy intake (EI), energy expenditure (EE) and energy balance (EB) have been discussed in detail in Chapter II and III. The data of EB are summarized in Table 1. Important is the observation that the performance of exhausting endurance exercise on day 4 and 5 resulted in a negative energy balance in N and FM, whereas subjects in Mf treatment were able to maintain energy balance.

**Table 1** Mean daily energy balance on rest days (3 and 6) and exercise days (4 and 5).

<table>
<thead>
<tr>
<th>Treatment</th>
<th>3</th>
<th>4</th>
<th>5</th>
<th>6</th>
<th>Cumulative day 4+5+6</th>
</tr>
</thead>
<tbody>
<tr>
<td>N</td>
<td>1.3±1.2</td>
<td>-9.6±0.8</td>
<td>-7.9±5.1</td>
<td>2.9±0.9</td>
<td>-15.3±2.1</td>
</tr>
<tr>
<td>Mf</td>
<td>4.9±0.9</td>
<td>-0.3±1.2</td>
<td>-0.5±1.7</td>
<td>2.1±1.3</td>
<td>1.9±2.1</td>
</tr>
<tr>
<td>FM</td>
<td>3.9±1.7</td>
<td>-2.5±2.0</td>
<td>3.3±1.9</td>
<td>1.6±1.8</td>
<td>4.8±3.8</td>
</tr>
</tbody>
</table>

Mean values ± SEM (MJ).

N = conventional CHO rich diet (n=13).
Mf = N supplemented with a maltodextrin - low fructose solution (n=6).
FM = N supplemented with a 50% fructose - 50% maltodextrin solution (n=7).

Asterisks indicate a significant difference with respect to values obtained with the conventional diet (N) \(^{*}p<0.01; \, ^{*}^{*}p<0.001\), (each subject is his own control). Statistical significance with respect to the initial values on day 3 is indicated by \(^{*}p<0.05; \, ^{*}^{*}p<0.01; \, ^{*}^{*}^{*}p<0.001\).

Mean energy and nitrogen content (± SEM) of blood from a sample taken at 7:00 AM on day 3 were 5.32±0.10 kJ.ml\(^{-1}\) and 33.4±1.0 mg.ml\(^{-1}\) respectively. The total amount of blood sampled was 50 ml. day\(^{-1}\), on day 3 to 6.

Mean nitrogen excretion with urine, feces and sweat is given in Table 2. Sweat was only collected during day 4 and 5.

Mean daily nitrogen intake from food and mean daily nitrogen balance are given in Table 3 and 4.
Table 2  Mean daily nitrogen excretion on rest days (3 and 6) and exercise days (4 and 5).

<table>
<thead>
<tr>
<th>Day</th>
<th>Treatment</th>
<th>3</th>
<th>4</th>
<th>5</th>
<th>6</th>
<th>Cumulative day 4+5+6</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>2.9±0.2</td>
<td>2.8±0.5</td>
<td>2.6±0.3</td>
<td>2.6±0.3</td>
<td></td>
</tr>
<tr>
<td></td>
<td>N</td>
<td>2.6±0.2</td>
<td>3.2±0.7</td>
<td>2.2±0.5</td>
<td>1.9±0.4</td>
<td></td>
</tr>
<tr>
<td></td>
<td>MF</td>
<td>2.9±0.3</td>
<td>1.9±0.3</td>
<td>2.7±0.4</td>
<td>2.3±0.5</td>
<td></td>
</tr>
<tr>
<td></td>
<td>FM</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>URINE</td>
<td>13.0±1.0</td>
<td>15.5±1.0</td>
<td>16.2±1.1</td>
<td>19.7±1.6</td>
<td>44.7±2.5</td>
</tr>
<tr>
<td></td>
<td>N</td>
<td>14.1±0.9</td>
<td>11.5±0.6</td>
<td>11.6±1.3</td>
<td>15.5±1.2</td>
<td>38.6±2.6</td>
</tr>
<tr>
<td></td>
<td>MF</td>
<td>12.7±1.2</td>
<td>12.4±0.8</td>
<td>14.9±1.5</td>
<td>15.1±2.4</td>
<td>42.4±2.8</td>
</tr>
<tr>
<td></td>
<td>FM</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>SWEAT</td>
<td>-</td>
<td>1.9±0.3</td>
<td>1.9±0.2</td>
<td>-</td>
<td></td>
</tr>
<tr>
<td></td>
<td>N</td>
<td>-</td>
<td>1.6±0.3</td>
<td>1.5±0.2</td>
<td>-</td>
<td></td>
</tr>
<tr>
<td></td>
<td>MF</td>
<td>-</td>
<td>2.2±0.6</td>
<td>2.3±0.6</td>
<td>-</td>
<td></td>
</tr>
</tbody>
</table>

Mean values ± SEM (g).
Asterisks indicate a significant difference with respect to values obtained with the conventional diet (N). *p<0.05 (each subject is his own control). Statistical significance with respect to the initial values on day 3 is indicated by *p<0.05; **p<0.01.

Table 3  Daily nitrogen intake calculated from weighed food intake

<table>
<thead>
<tr>
<th>Day</th>
<th>Treatment</th>
<th>3</th>
<th>4</th>
<th>5</th>
<th>6</th>
<th>Cumulative day 4+5+6</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>N</td>
<td>19.3±0.9</td>
<td>18.9±1.2</td>
<td>18.6±1.1</td>
<td>21.5±1.0</td>
<td>58.9±2.6</td>
</tr>
<tr>
<td></td>
<td>MF</td>
<td>19.7±1.0</td>
<td>15.7±1.5</td>
<td>17.7±1.4</td>
<td>17.9±1.2</td>
<td>51.4±2.8</td>
</tr>
<tr>
<td></td>
<td>FM</td>
<td>19.0±1.6</td>
<td>15.4±1.3</td>
<td>15.7±1.7</td>
<td>15.9±2.4</td>
<td>47.0±4.9</td>
</tr>
</tbody>
</table>

Mean values ± SEM (g).
Nitrogen content of all foods supplied was determined previously by the che-miluminescence method.
Asterisks indicate a significant difference with respect to corresponding values obtained with conventional diet (N). *p<0.05; **p<0.01; ***p<0.001 (each subject is his own control). Statistical significance with respect to the initial values on day 3 is indicated by *p<0.05; **p<0.01.
Table 4  Mean daily nitrogen balance on rest days (3 and 6) and exercise days (4 and 5).

<table>
<thead>
<tr>
<th>Treatment</th>
<th>3</th>
<th>4</th>
<th>5</th>
<th>6</th>
<th>Cumulative day 4+5+6</th>
</tr>
</thead>
<tbody>
<tr>
<td>N</td>
<td>1.9±1.5</td>
<td>-2.6±1.3</td>
<td>-3.7±1.1</td>
<td>-2.5±1.4</td>
<td>-8.7±2.5</td>
</tr>
<tr>
<td>Mf</td>
<td>1.4±0.7</td>
<td>-2.1±0.4</td>
<td>1.2±2.2</td>
<td>-1.1±1.0</td>
<td>-2.1±2.5</td>
</tr>
<tr>
<td>FM</td>
<td>1.7±1.8</td>
<td>-2.9±1.4</td>
<td>-6.0±2.4</td>
<td>-2.5±2.9</td>
<td>-11.3±4.5</td>
</tr>
</tbody>
</table>

Mean values ±SEM (g).

In N treatment urinary nitrogen increased significantly during day 4, 5 and 6. In contrast, subjects with Mf treatment had a significant reduced nitrogen excretion in the urine on the first exercise day and tended to be reduced on the second exercise day (p=0.06) compared to the first rest day (day 3). Mf subjects reached their initial resting excretion level again on the second rest day (day 6). In FM treatment there was no difference between the first rest and exercise day. On day 5 and 6 mean urinary nitrogen excretion was increased. However, compared to N (each subject was his own control) the values in FM were significantly lower on day 4 and 6. Sweat nitrogen excretion was similar in the 3 treatments on the two exercise days.

Mean nitrogen intake from food tended to be lower in Mf and FM compared to N (Table 3). Nitrogen balance as a reflection of total daily nitrogen intake and nitrogen losses from the body was positive on day 3 and became negative on the first exercise day (day 4) in all treatments. On the second exercise day mean nitrogen balance became more negative in N and FM whereas it tended to be positive in Mf. On day 6 nitrogen balance was negative in all treatments.

**Discussion**

Energy balance was positive on the first resting day in all treatments and became negative as a result of sustained exercise on day 4 and 5 in N and FM. In contrast Mf treatment resulted in a positive energy balance over the entire exercise period (see Table 1). Notwithstanding the large contribution of CHO to total daily energy intake, in the N and MF treatment this intake was inadequate to balance the amount of CHO oxidized, most probably due to the high levels of energy expenditure during sustained exercise. In the Mf treatment CHO intake appeared to be adequate even under these circumstances (For a detailed analysis see Chapter III). Based on the observed interactions between energy balance and nitrogen losses from the body on the one hand, and CHO availability and protein degradation on the other hand it
was assumed that subjects who were able to maintain energy balance and 'CHO balance' would not develop a negative nitrogen balance, despite the prolonged exhausting exercise.

As can be seen from the data (Table 2) there was no significant difference in daily nitrogen loss by the feces. This indicates that protein digestion and absorption in the described experimental circumstances were not influenced by the heavy exercise. Urinary nitrogen content increased with exercise and showed the highest mean values for treatment N and FM, where the subjects developed a negative energy balance. In Mf urinary nitrogen excretion tended to decrease during exercise days and increased again to initial values during recovery on day 6.

These data suggest protein sparing in Mf treatment compared to N and FM. This suggestion is supported by the data of sweat urea excretion which had the lowest values for Mf. It has been observed that muscle glycogen depletion enhances the amount of nitrogen excreted in the sweat (Lemon, 1980). In the present study sweat nitrogen excretion on the first exercise day was similar to that on the second exercise day in each treatment. This may suggest that the glycogen depletion on both exercise days has been similar. The lower mean sweat nitrogen excretion in Mf compared to N and FM may be related to the observed glycogen sparing in these subjects (see Chapter V). Besides daily energy and CHO intake it is known that the amount of protein ingested also affects protein metabolism and nitrogen balance. A low protein intake may initiate a negative nitrogen balance whereas a high protein intake stimulates protein synthesis and retention (Meredith, 1982). Therefore protein intake was fixed to a minimum of 1.2 g.kg⁻¹.day⁻¹ in order to rule out a negative influence of decreased protein intake which might result from a changed appetite and food selection during the exercise days. (For a detailed description of food intake see Chapter II and III). From the data summarized in Table 3 it can be seen that in N treatment nitrogen intake from food was maintained at the initial level also during the exercise days. Mean protein intake decreased 15 to 20% in Mf and FM, most probably as a result of changed food selection due to the diet manipulation. The lowest mean daily protein intake in these groups amounted to 1.43g.kg⁻¹.day⁻¹ compared to 1.7 g.kg⁻¹ in N. Although this lower level of protein consumption was still markedly above the recommended daily allowance for protein intake of 0.8 g.kg⁻¹.day⁻¹ (NRC, 1984) it may be hypothesized that this level is too low for subjects performing sustained heavy exercise and thus may have influenced the amount of nitrogen retained and/or excreted.

At present it cannot be concluded which relative roles the negative energy balance and probably protein intake have played in the development of a negative nitrogen balance. Nitrogen balance was positive on the first rest day and became negative on the first exercise day (day 4) irrespective of energy
balance. However, on the second exercise day, nitrogen balance tended to be more negative in N and FM and was positive in Mf. On that day N and FM were in negative energy and CHO balance, whereas in Mf treatment energy and CHO balance was maintained (see Table 1 and Chapter III of this study).

An interesting observation is that the subjects in Mf who developed a negative nitrogen balance on a particular day always were on a level of protein intake lower than 1.6 g·kg⁻¹. For example, in one subject nitrogen balance was -2.14 g negative on the first exercise day. Protein intake on this day was 1.2 g·kg⁻¹ = 12.64 g nitrogen·day⁻¹. On the second exercise day nitrogen balance was 3.69 g positive and protein intake was 1.8 g·kg⁻¹ = 18.88 g nitrogen·day⁻¹. The delta change in nitrogen balance and nitrogen intake between both days was similar. This relation was present in all subjects in Mf both at rest and during exercise and suggests that the quantitative protein intake becomes critical with respect to the daily nitrogen balance under exercise circumstances in all cases where energy balance is maintained and CHO intake is sufficient. In Mf a correlation coefficient of .806, p<.001, was derived when all the day to day changes in nitrogen balance were plotted against day to day changes in nitrogen intake (see Fig 1). These data strongly suggest that athletes involved in heavy physical exercise should maintain a constant level of protein intake higher than the RDA of 0.8 g·kg⁻¹, most probably 1.5 to 1.8 g·kg⁻¹, in order to maintain nitrogen balance.

![Graph showing daily changes in nitrogen intake (g) and nitrogen balance (g)](image)

**Fig 1** Day to day changes in nitrogen intake (g) (Delta int) and nitrogen balance (g) (Delta nBal) in subjects receiving Mf treatment $R = .806, p<.001$.

During recovery (day 6), mean nitrogen balance was negative in all treatments, despite the observation that energy balance was positive. It may be assumed that this post exercise negative nitrogen balance may be caused by a persisting effect of exercise on protein synthesis (depression) and
degradation (maintenance or increase). This may have its consequences in particular for the recovery and adaptation of functional proteins involved in exercise. From the available biochemical data, including muscle glycogen (Chapter V), it cannot be concluded which factor causes this increased protein degradation during recovery. Increased post-exercise nitrogen losses have also been reported in other studies (Decombaz, 1979; Lemon, 1983, 1984).

In one study (Gontzea, 1975) it was observed that physical effort over a prolonged period of time caused a negative nitrogen balance during the first week. In the period thereafter nitrogen balance was reached again. The subjects in this study showed a training-effect, resulting in an increased work capacity. The present study was too short to study long term effects but it can be assumed that exercise to exhaustion causes always the same metabolic effects, irrespective of training adaptations, in contrast to exercise at sub-maximal work levels where exhaustion is not reached. In the latter training adaptations, leading to an enhanced work capacity, will lower the metabolic stress which may reduce protein catabolism.

In summary the following conclusions can be made:

1. Repeated heavy sustained exercise in trained athletes, who are consuming a conventional CHO rich diet, causes a negative energy balance. As a result nitrogen balance is negative, despite a mean protein intake of 1.7 g.kg⁻¹.day⁻¹.

2. Supplementation of the conventional diet with a CHO rich liquid led to a maintenance of energy balance and induces protein sparing.

3. Even under circumstances of maintaining energy and 'CHO balance', a minimal protein intake of 1.5 g.kg⁻¹.day⁻¹ was needed to reach nitrogen balance.
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CHAPTER V: METABOLIC CHANGES INDUCED BY SUSTAINED EXHAUSTIVE CYCLING AND DIET MANIPULATION, A CONTROLLED TOUR DE FRANCE SIMULATION STUDY


Introduction
The human body is able to adapt to extremely heavy workloads and as a result of long term adaptations, professional cyclists are able to increase their energy output to very high levels on a day to day basis. In one of the world's most extreme cycling competitions, the Tour de France lasting 23 days, it was shown that energy expenditure (EE) can exceed 32 MJ on a hot competition day in the mountains, while mean (EE) amounted to 24 MJ per day (Saris, 1988). The same study revealed that the cyclists were only capable of competing at such an extreme level by matching energy intake to energy expenditure. This was achieved mainly by intake of liquid carbohydrate- and energy rich meals throughout the day in addition to the normal diet. Cyclists who were not able to maintain energy balance frequently experienced malperformance and sometimes were forced to quit prematurely. Such imbalance normally resulted from diarrhea or inadequate food intake caused by changes in food tolerance and appetite, due to the extremely fatiguing exercise. Furthermore time for recovery is limited.

In contrast to this diet behavior of professional cyclists, nutritionists and physiologists generally belief that a normal well balanced diet supplying 50-60 en% carbohydrate (CHO) and 0.8-1 g of protein kg.day⁻¹ will be sufficient for any sportsman or woman in any situation. However, the question then arises how athletes can meet their daily energy and CHO needs, when both are increased to extreme levels.

Apart from problems due to the time available to eat and digest, the volume of the food and the changes in appetite (Brouns, 1986), one may question the possible interaction between changes in energy metabolism as a result of long term training and the influence of changed eating habits or diet manipulation.

It is known that liver and muscle glycogen stores limit endurance performance capacity whenever exercise intensities increase
above 60% $\dot{V}O_2$ max (Bergström, 1967; Saltin, 1967; Sherman, 1984). As a result of long-term training the body adapts towards enhanced fat metabolism in order to minimize this limitation, leading to muscle glycogen sparing and delaying the onset of fatigue (Gollnick, 1982, 1986; Saltin, 1983; Paul, 1975; Stanckiewicz, 1978).

On the other hand it has been discussed that maximal power output decreases with increasing contribution of fat in energy metabolism, such as after endogenous CHO depletion, or following a high fat diet, both of which will enhance blood fatty acid concentration and utilization. As a result, when fat is the main substrate, power output will drop to 50% of maximal capacity (Newsholme, 1983), possibly as a result of a decreased maximal energy flow from fat compared to CHO (McGilvery, 1973). Studies from the early sixties showed that glycogen stores were increased when CHO intake was increased drastically during the days preceding exercise and that this increase was associated with longer exercise times to exhaustion (Bergström, 1967; Gollnick, 1972; Saltin, 1967). The same holds for ingestion of substantial amounts of CHO during exercise, possibly delaying glycogen depletion and thereby improving performance (Coyle, 1983; Ivy, 1983; Edwards, 1984).

However, it has also been shown that fat and CHO behave as competitive substrates when passing the muscle cell membrane for oxidation (Rennie, 1977; Randle, 1963; Newsholme, 1976). Infusion of fat decreases CHO oxidation and enhances fat metabolism while CHO infusions cause the opposite effect (Felber, 1964; Büber, 1968; Gomez, 1972). As a consequence, the quantity of CHO degraded may be larger than the quantity ingested when the latter is small. Intake at rest prior to exercise, enhancing insulin secretion may also lead to enhanced rates of glycogen degradation and reduced times until exhaustion (Hargreaves, 1984; Foster, 1979). From this it may be postulated that the effectivity of diet manipulation to a large extent depends on the quantity of nutrients involved and on the moment of intake.

Apart from this it has been shown that protein degradation during exercise is increased whenever glycogen stores are depleted in order to supply precursors for gluconeogenesis (Ahlborg, 1974; Rennie, 1981a; Lemon, 1981b; Dohm, 1982) and that CHO intake and/or enhanced glycogen availability limits this protein degradation (Lemon, 1981; Rennie, 1981; Munro, 1953; Long, 1976). The fact that protein degradation is also increased at rest
whenever energy balance is negative for a prolonged period of time, such as during fasting, is also related to glycogen depletion and enhanced gluconeogenesis (Felg, 1969; Sauerwein, 1982). Taking into account these interrelationships it can be hypothesized that performance capacity can be maintained at a higher level, and that protein degradation will be limited, whenever energy balance is maintained and CHO intake matches CHO degradation.

Since cyclists competing successfully in the Tour de France were able to maintain energy balance only by using liquid CHO rich foods in addition to the normal diet and no controlled studies in the field of ultra-endurance sports supporting this practice are available, it was decided to design a cross-over study in which the nutritional and biochemical changes of cyclists ingesting a CHO rich diet composed of conventional but CHO rich food (treatment N) could be compared with those after diet manipulation, in which the same diet was supplemented with a substantial amount of CHO concentrated liquids (treatment Mf). The study was performed in a controlled laboratory set-up in which two heavy days with an energy expenditure comparable to that observed in the Tour de France were simulated.

MATERIALS AND METHODS

Subjects

Thirteen highly trained cyclists participated in this study. Physical characteristics are presented in Table 1.

Table 1 Physical characteristics of experimental subjects

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age (yrs)</td>
<td>20.0 ± 2.1</td>
</tr>
<tr>
<td>Weight (kg)</td>
<td>73.3 ± 1.7</td>
</tr>
<tr>
<td>Height (cm)</td>
<td>179.6 ± 1.7</td>
</tr>
<tr>
<td>( \dot{V}O_2 ) max (ml.kg(^{-1}).min(^{-1}))</td>
<td>65.1 ± 1.2</td>
</tr>
<tr>
<td>Wmax (Watt)</td>
<td>390 ± 8.0</td>
</tr>
<tr>
<td>Wmax (Watt.kg(^{-1}))</td>
<td>5.33 ± 0.36</td>
</tr>
<tr>
<td>Body fat (%)</td>
<td>12.0 ± 0.8</td>
</tr>
</tbody>
</table>

Mean values ± 1 SEM (n=13)
Experimental design
The subjects were asked not to participate in vigorous training or competition during the two days preceding the arrival in the laboratory and to ingest a normal but CHO rich diet (dietary advice was given by a registered dietician). All subjects were informed about the nature, purpose and possible risks of the study, before giving their voluntary written consent to participate. The experiment was conducted over seven sequential days using a semi-automated respiration chamber system.

The subjects reported to the laboratory on Sunday evening (day 1) at 9:00 PM in order to get accustomed to the chamber. They received a time schedule for all activities in the forthcoming week. The next preparation day (day 2) a by each subject selected supply of food and fluid of known quantity and composition was made available. It is known that differences in substrate mobilization and utilization may occur depending on diet composition, exercise executed and quantitave endogenous CHO stores. (Galbo, 1979; Gollnick, 1981; Lemon, 1981; Maughan, 1978). For that reason diet composition and amount of exercise was completely controlled during 24 h previous to the experiment (day 2). At least 60 en% CHO and a minimal protein intake of 1.2 g.kg\(^{-1}\) were assured in order to realize a complete build-up of endogenous substrate pools and to avoid interindividual differences in pre-test nutritional and metabolic status. Protein intake was calculated daily, immediately after dinner. To assure a minimum intake of 1.2 g.kg\(^{-1}\), subjects were supplemented with a protein concentrate whenever intake was calculated to be too small for that day. This food supply was continuously provided until the end of the experiment on Saturday morning 9:00 AM six days later. Food for breakfast, lunch and in-between meals was supplied at 7:30 AM. However, extra's could be obtained throughout the day upon request. Dinner was served at 6:00 PM. There were no quantitative limitations. The cyclists were instructed about the importance of adequate food and fluid intake and were encouraged to eat and drink as much as desired.

Weight and volumes of foods and drinks were measured and registered in a diary allowing for analysis of 24 h intake. The residual amounts were weighed and accounted for in the final calculations. Energy and nitrogen content of all available food items had been previously determined. Actual performance capacity expressed in Watt (Wmax) and maximal oxygen uptake
\( \dot{V}O_2 \text{ max; ml.kg}^{-1}.\text{min}^{-1} \) were determined at 10:00 AM, day 2, during an incremental cycle ergometer test. The respiration chambers were closed at 4:00 PM on day 2 and measurements for calculation of energy expenditure were started (Fig 1). Blood samples were taken daily at 7:00 AM immediately after waking-up, at 12:00 AM and 3:00 PM. This was done through a special designed airlock in the respiration chamber wall. The samples were drawn into EDTA evacuated tubes from a teflon catheter, which was inserted into an antecubital vein at 7:00 AM daily, and was connected with a three-way stopcock. The blood was immediately put in ice water.

Days 3 and 6 of the experiment represent standardized rest days during which each subject cycled 45 min at 40% Wmax at 10:00 AM and 2:00 PM. Rest days were included in the program in order to determine resting status for comparison with the following two exercise days. On days 4 and 5, each subject exercised to exhaustion (E).

Fig 1 Time schedule of measurements performed. On day 2 actual performance capacity (Wmax; Watt) and maximal oxygen uptake (\( \dot{V}O_2 \text{ max; ml.min}^{-1} \)) were determined at 10:00 AM using an incremental bicycle ergometer test. Day 3 and 6 were rest days; cycling at 40% Wmax during 45 min at 10:00 AM and 2:00 PM. Day 4 and 5: exercise to exhaustion. Day 7: end of the experiment at 9:00 AM. A muscle biopsy was taken on day 2, 5 and 6. Blood was sampled at 7:00 AM, 12:00 AM and 4:00 PM on day 3 to 6.

Sweat samples were collected on day 4 and 5 by absorption into predried pads located in water- and airtight capsules (Lemon, 1983). Ten min prior to exercise sweat capsules were placed in the infraspinous fossa of the scapula and kept in place by an
elastic mesh vest. Exercise was started at 10:00 AM with a 30 min warming-up, followed by exercise at intensities of 80% and 50% Wmax respectively. Eighty percent and 50% Wmax were chosen in order to mimic cycling ahead of the group or benefitting from wind shielding within the group respectively. At 12:00 AM exercise was interrupted for 5 min to measure body weight, to collect blood samples and to change the sweat capsules. Thereafter exercise was continued at 50% and 60% Wmax allowing for a food intake. After 3 h 44 min exercise was continued again at 80% and 50% Wmax in 3 min intervals. Finally at about 2:30 PM, the wattload was set at 90% Wmax and the cyclists were asked to maintain pedaling frequency greater than 60 RPM. Ninety percent of Wmax - equivalent to approximately 80% VO2 max - at the end was chosen in order to simulate a finish on a mountain top (Fig 2).

Fig 2 The cycling program was as follows: warming-up 30 min at 30% Wmax, followed by 10 min 60% Wmax, followed 9 times by 6 min 80% Wmax and 6 min 50% Wmax respectively. After this, exercise was shortly interrupted for collecting blood and sweat samples and was then continued at 50 Watt for 10 min, followed by 50 and 50% Wmax both for 30 min. Exercise intensity was then increased to 80% Wmax for 3 min followed by 3 min at 50% Wmax, which was repeated 8 times. Finally exercise intensity was increased to 90% Wmax which had to be performed until exhaustion.

During exercise, fluids were available ad libitum (tea, coffee, milk, lemonade and a placebo 'sport drink' consisting of artificially sweetened, coloured, and mineralized water). Feces
During exercise, fluids were available ad libitum (tea, coffee, milk, lemonade and a placebo 'sport drink' (consisting of artificially sweetened, coloured, and mineralized water). Feces and urine were kept at -20°C in a deep freezer toilet and were collected in 24 h periods.

A percutaneous needle biopsy sample (Evans, 1982) was taken from the m. vastus lateralis at 4:00 PM at rest on day 2, 45 min after reaching exhaustion on day 5 and after 24 h of recovery on day 6. (The biopsy could not be taken immediately after reaching exhaustion because of measurements which had to be finished before the respiration chamber could be opened). The biopsy sample was immediately frozen in liquid nitrogen for analysis of glycogen and triacylglycerol.

Diet manipulation

For diet manipulation the total group of 13 subjects was divided into two subgroups. In 6 randomly selected subjects the normal diet was supplemented with an experimental, high maltodextrin - low fructose solution (Perform®, Wander Ltd; Mf). The order of treatment, normal diet or manipulated diet, was randomized. The other seven subjects were supplemented with another experimental beverage. In this paper only the data from N treatment (n=13) and the Mf treatment (n=6) will be presented.

Although there is a wide variety of CHO sources, it was specifically decided to use a solution composed of mainly long-chain glucose polymers and a low fraction of fructose, because long-chain polymers may be of advantage for the athlete by means of maximizing CHO intake and absorption, while minimizing risks of gastrointestinal distress (Brouns, 1987). A small quantity of fructose was added for reasons of taste, in order to increase palatability. The supplement, prepared as a 20% (w/v) solution, was made available as follows: standardized resting days 500 ml during the morning, afternoon and evening (total supply 1500 ml), exercise days during cycling ad libitum and in the evening after dinner, 1000 ml. The cyclists during N treatment were supplemented with equal amounts of a placebo drink at the same times in order to rule out psychological effects due to the drink.

The composition of the drinks is presented in Table 2.

The experimental design used during the Mf trial was exactly the same as described for N. The placebo drink was supplied during N treatment in equal quantities and at the same times as the
beverage in Mf and FM treatment.

The placebo was flavored with citruspowder, sweetened with saccharin and cyclamate and artificially colored.

Table 2 Composition of beverages

<table>
<thead>
<tr>
<th>Beverage</th>
<th>Maltodextrin</th>
<th>Fructose</th>
<th>K⁺</th>
<th>Osmolarity</th>
<th>pH</th>
<th>Energy content</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>g. l⁻¹</td>
<td>g. l⁻¹</td>
<td>mmol</td>
<td>mosm</td>
<td></td>
<td>kJ. l⁻¹</td>
</tr>
<tr>
<td>Placebo</td>
<td>0</td>
<td>0</td>
<td>8.5</td>
<td>106</td>
<td>4.38</td>
<td>-</td>
</tr>
<tr>
<td>Mf</td>
<td>150</td>
<td>33</td>
<td>8.5</td>
<td>390</td>
<td>4.45</td>
<td>3158</td>
</tr>
</tbody>
</table>

Analyses

Energy expenditure was determined using an indirect semi-automated calorimeter as previously described by Schoffelen et al. (1986). Gasflows were measured with a dry gasmeter (Dort, the Netherlands), oxygen was analyzed using a Servomex® paramagnetic oxygen analyzer (Taylor, England) and carbon dioxide using infrared CO₂ analyzer (Hartmann and Braun, Germany).

Energy expenditure was calculated according to the formula of Consolazio (1963):

\[ E = 3.78 \dot{V}O_2 + 1.14 \dot{V}CO_2 - 2.98 \text{Nu} \]

Where E = energy expenditure (kcal.min⁻¹), \( \dot{V}O_2 \) = oxygen consumption (ml.min⁻¹), \( \dot{V}CO_2 \) = CO₂ production urine (ml.min⁻¹) and Nu is the amount of nitrogen excreted in (mg.).

Daily energy balance was calculated from energy expended and energy intake as calculated from daily food and fluid consumption. Corrections were made for energy losses from feces, urine and sweat and for the blood samples drawn. Energy content of food, fluid, feces, urine and blood was determined by bombcalorimetry (IKA Germany). Energy content from sweat was calculated from total sweat urea, assuming that the remainder is negligible. Total sweat loss was determined from body weight change while accounting for fluid intake, urine production, blood volume loss,
and respiratory water loss (Mitchel, 1972). The sweat volume collected in the capsules was determined by capsule weight changes using a analytical balance (Mettler). Sweat urea content was calculated enzymatically (urease method Boehringer 396346). Nitrogen content of blood and urine was determined by the chemiluminescence method (Antek, Germany). Daily nitrogen losses were then calculated.

CHO and fat oxidation was calculated from non-protein respiratory quotient (R). Protein oxidation was calculated from daily nitrogen losses in urine and sweat. From the data gathered the daily contribution of CHO, fat and protein to total 24 h energy exchange as well as the relative contribution during exercise was calculated.

Muscle glycogen content in the biopsy sample, which was freeze-dried (Leybold Hereaus, GT2, Germany), was determined fluorimetrically after HCL hydrolysis (Passonneau, 1974) and expressed as mmol of glycogen units . kg⁻¹ dry weight. Muscle triacylglycerol content was assessed as described elsewhere (Van der Vusse, 1982). In short freeze-dried tissue specimen were extracted with chloroform-methanol (2: 1 by vol). The various lipid classes were separated using one dimensional thin layer chromatography. The fatty acids in the triacylglycerol spot were transmethylated with BF₃ in methanol. The fatty acid methyl esters were quantitated by gas liquid chromatography.

Plasma volume changes were calculated from hemoglobin and hematocrit values according to Dill and Costill (1974). Hematocrit was determined in all blood samples. Hemoglobin was measured with the hemoglobin cyanide method.

Blood chemistry

The following methods have been applied for determinations in blood:

Blood glucose - glucose oxidase method (Boehringer 124036).
Lactate - oxidimetric analysis (Roche analyzer 640),
Acetoacetate and 3-hydroxybutyrate - enzymatically with 3-hydroxybutyrate dehydrogenase (Bergmeyer, 1965),
Glycerol - glycerol kinase UV test (Boehringer 297771),
Free fatty acids - enzymatic and fluorometrically, Transcon 102 FNR analyzer (OY Elomit, Helsinki, Finland, Härkönens and Adlercreutz 1981).
Blood ammonia - glutamate dehydrogenase UV test (Boehringer 125857),
Urea - urease method (Boehringer 396346).
Catecholamines - HPLC, electrochemical detection system (Smedes, 1982).
Insulin, glucagon and cortisol - Radio-immuno-assay (Medgenix Belgium; DPC Kit, USA).

Statistics
Wilcoxon signed rank test was used to compare the data of the first standardized rest day to those of the following exercise and recovery days as well as for comparison of differences between the two treatments N (n=13) and Mf (n=6). For the latter each subject served as his own control. For all analyses, the 0.05 level was used as the minimum level of confidence for statistical significance.

RESULTS
The results of the study dealing with food consumption, energy-, fluid- and nitrogen balance, as well as relative contribution of CHO and fat in energy metabolism have been described and discussed extensively in other papers (Brouns, 1988, 1988a, 1988b see Chapter II, III and IV). The results most important with respect to the studied biochemical parameters are summarized in Table 3 and Fig 3.

Plasma Volume
On both rest days plasma volume in M and Mf showed a significant increase at 12:00 AM and 4:00 PM compared to 7:00 AM on the first day. All though Mf subjects tended to have decreased plasma volumes below the initial level on exercise days compared to N treatment, this was not statistically significant.
Table 3 Summary of observations with respect to nutritional indices and performance time.

<table>
<thead>
<tr>
<th></th>
<th>Day 3</th>
<th>4</th>
<th>5</th>
<th>6</th>
<th>Cumulative (day 4+5+6)</th>
</tr>
</thead>
<tbody>
<tr>
<td>CHO intake (g)</td>
<td>N 61.8 ±1.7</td>
<td>62.9 ±1.3</td>
<td>62.4 ±1.6</td>
<td>62.7 ±1.6*</td>
<td>-</td>
</tr>
<tr>
<td>(en%)</td>
<td>Mf 66.8 ±2.7</td>
<td>80.5 ±1.5**</td>
<td>79.7 ±1.3**</td>
<td>67.5 ±2.6</td>
<td></td>
</tr>
<tr>
<td>Nitrogen intake (g)</td>
<td>N 19.3 ±0.9</td>
<td>18.9 ±1.2</td>
<td>18.6 ±1.1</td>
<td>21.5 ±1.0</td>
<td>58.9 ±2.6</td>
</tr>
<tr>
<td>Energy Balance (MJ)</td>
<td>Mf 19.7 ±1.0</td>
<td>15.7 ±1.5</td>
<td>17.7 ±1.4</td>
<td>17.9 ±1.4***</td>
<td>51.3 ±2.8</td>
</tr>
<tr>
<td>Nitrogen balance (gN)</td>
<td>N 1.18±1.20</td>
<td>-9.78±0.82***</td>
<td>-8.12±0.88***</td>
<td>2.65±0.92*</td>
<td>15.28±2.05</td>
</tr>
<tr>
<td>Final performance at 90 % Wmax, (min) over day 4 and 5.</td>
<td>N 9.9±2.6</td>
<td>Mf 22.4±7.9*</td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

For complete description of the nutritional data see Brouns 1988, 1988a, 1988b. Mean values ± SEM (for N, n=13; Mf, n=6). CHO intake was determined by weighed food intake procedure using a computerized food table for analysis. Nitrogen intake was determined by weighed food intake after previous determination of the nitrogen content of all food items by the chemiluminescence method. Energy balance was determined by indirect calorimetry and weighed food intake after previous determination of energy content of all food items by bomb calorimetry. Nitrogen balance was calculated from total nitrogen intake and nitrogen losses in urine, sweat, feces and blood. Statistical significance with respect to the initial value on day 3 is indicated by *p<0.05; **p<0.01; ***p<0.001. Statistical significance of Mf with respect to N (each subject is his own control) is indicated by *p<0.05; **p<0.01; ***p<0.001.
Fig 3  Relative changes in plasma volume at rest (day 3 and 6) and during exhaustive exercise days (day 4 and 5). Vertical bars indicate 1 SEM. There was no statistically significant difference between the treatment groups. Statistical significance with respect to the initial value on day 3 is indicated by *p<0.05; **p<0.01; ***p<0.001 ; ---N, --- = Mf.

Blood glucose changes are represented in Fig 4. During N treatment blood glucose remained almost the same throughout the entire test. As a result of exercise there was a small but significant elevation on day 4 and 5. During Mf treatment blood glucose values were significantly more elevated on the exercise days than during N treatment. There was no difference between the two rest days.
Blood lactate was significantly elevated after 2 h of exercise and at the moment of exhaustion. This elevation was the same for both N and Mf. Mean maximal lactate values did not exceed 4 mmol.l⁻¹, despite the fact that workload was increased to 90% Wmax before reaching the moment of exhaustion (Table 4).

**Table 4 Blood lactate concentration**

<table>
<thead>
<tr>
<th></th>
<th>7:00 AM</th>
<th>12:00 AM</th>
<th>4:00 PM</th>
</tr>
</thead>
<tbody>
<tr>
<td>Day 3</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>N</td>
<td>1.5±0.1</td>
<td>2.8±0.3</td>
<td>3.8±0.5</td>
</tr>
<tr>
<td>Mf</td>
<td>1.3±0.1</td>
<td>2.6±0.4</td>
<td>3.6±0.5</td>
</tr>
</tbody>
</table>

Day 4  

<table>
<thead>
<tr>
<th></th>
<th>7:00 AM</th>
<th>12:00 AM</th>
<th>4:00 PM</th>
</tr>
</thead>
<tbody>
<tr>
<td>N</td>
<td>1.3±0.1</td>
<td>2.6±0.2</td>
<td>3.0±0.4</td>
</tr>
<tr>
<td>Mf</td>
<td>1.5±0.1 *</td>
<td>2.4±0.5</td>
<td>3.0±0.4</td>
</tr>
</tbody>
</table>

Mean blood lactate values ± 1 SEM (mmol.l⁻¹), corrected for changes in plasma volume on exercise days 4 and 5. Blood samples were taken at 7:00 AM immediately after waking up, at 12:00 AM immediately after cycling at 80% Wmax and at exhaustion after having cycled at 90% Wmax. Statistical significance with respect to the initial value on day 3 is indicated by *p<0.05; **p<0.01. Statistical significance of Mf with respect to N (each subject is his own control) is indicated by "p<0.05.

Plasma glycerol was stable at rest; post-exercise resting levels (day 6) were not different from pre-exercise resting levels (day 3). During exercise glycerol was significantly elevated in both treatments. However, this elevation was significantly greater during N treatment than during Mf treatment at the end of exercise (Fig 4).

Plasma free fatty acids. During the first resting day FFA levels were significantly lower than at 7:00 AM, after an overnight fast (7.00 AM of day 4 serves as reference value). This in contrast to the post exercise rest day where morning levels were elevated. Both treatments showed similar responses during the rest days. During exercise days however, FFA increased significantly more in N than in Mf. This increase remained present during the post exercise night. In both treatments FFA increased more on the second exercise day than on the first (Fig 4).
Plasma ketones. Acetoacetate at rest in N was significantly elevated at 4:00 PM compared to 7:00 AM of the first rest day. As a result of exercise the level increased significantly, remained elevated during the night, increased further during the second exercise day and remained significantly elevated throughout the post exercise recovery day. In Mf plasma acetoacetate increased less and tended to be significantly lower throughout the experiment (Fig 4). Beta-hydroxybutyric acid response was similar to that of acetoacetate, except that it returned somewhat faster to basal levels (Fig 4).

![Graph of changes in protein metabolism](image)

Fig 5 Changes in protein metabolism as measured by ammonia and urea levels in whole blood and blood plasma, corrected for changes in plasma volume. Statistical significance with respect to the initial value on day 3 is indicated by \( *p<0.05; \quad \cdot\cdot*p<0.01; \quad \cdot\cdot\cdot p<0.001 \). Statistical significance of Mf with respect to N (each subject is his own control) is indicated by \( *p<0.05. \quad \text{---} = \text{Mf.} \quad \boxed{\text{= exercise}} \)
Plasma urea was stable in N during day 3 and increased significantly lower during both exercise days, reaching the highest level at the moment of exhaustion. In Mf plasma urea was significantly on lower day 4. There was no significant difference in response between N and Mf on day 5 and 6 (Fig 5). On day 6 plasma urea remained elevated and increased to a level comparable to day 4 and 5.

Plasma ammonia responses during both treatments were equal. During exercise days plasma ammonia increased significantly and reached the highest values at exhaustion. The values at the moment of exhaustion on the second exercise day tended to be lower than on the first exercise day (Fig 5).

Plasma insulin levels in N and Mf were equal at 7:00 AM of the first rest day (Fig 6). During this day insulin increased significantly. This increase was more pronounced in Mf. During the following exercise days insulin in N was significantly lowered at all measuring times. The food induced insulin response during the day remained entirely absent during the second exercise day. The post-exercise recovery day showed a normal response again. However, the insulin level at 7:00 AM and 12:00 AM remained significantly lower compared to the pre-exercise rest day. The subjects under Mf treatment responded in a similar manner. However, their insulin levels were significantly elevated compared to N during the exercise days, whereas food induced insulin increase on the second exercise day was not observed at all.

Plasma glucagon responses in N and Mf were not different. Plasma glucagon was significantly elevated at the end of both exercise days compared to the 3:30 PM sample of the pre-exercise rest day. The overnight fasting level between both exercise days was not different from that following the first rest day and from that post-exercise. (Fig 6).

Plasma epinephrine and norepinephrine responded to exercise in a similar way. It should be mentioned, however, that the catecholamines were only determined at the moment of exhaustion and at the same measurement time during day 2 and 6. Interestingly norepinephrine tended to increase more in Mf than in N, whereas the opposite was the case for epinephrine (Fig 6).

Plasma cortisol decreased during the day on both rest days. This decrease was absent on both exercise days resulting in a significant elevation half-way through exercise and at the end of exercise. The response in N and Mf was the same (Fig 6).
Fig 4  Changes in CHO and fat metabolism as measured by substrate levels in blood. Blood glucose and FFA are presented as absolute values. Ketones and glycerol are corrected for changes in plasma volume. Statistical significance with respect to the initial value on day 3 is indicated by *p<0.05; **p<0.01; ***p<0.001. For FFA the initial value is day 4, 7:00 AM. Statistical significance of Mf with respect to N (each subject is his own control) is indicated by *p<0.05. Mf = exercise
Fig 6 Changes in plasma hormone levels. --- = N, --- = Mf. Statistical significance with respect to the initial value on day 3 is indicated by $p<0.05$; $^{**}p<0.01$; $^{***}p<0.001$. Because of the influence of diurnal rhythm, comparison in time was made with respect to the same times on day 3. For glucagon day 4; 7:00 AM was taken as reference morning value. Statistical significance of Mf with respect to N (each subject is his own control) is indicated by $p<0.05$. 

--- exercise
Muscle glycogen on rest day 2 was not different among treatments. As a result of exercise, glycogen decreased significantly in both N and Mf, however Mf post-exercise levels were significantly higher than the values of the same subjects under N treatment. After 24 h of recovery N subjects did not restore their muscle glycogen to initial levels, whereas in Mf subjects glycogen was restored showed supercompensation (Fig 7).

Fig 7 Mean muscle glycogen content at rest as determined from a biopsy sample on day 2, 45 min after exhaustion on day 5, and after 24 h recovery on day 6. Statistical significance with respect to the initial value on day 3 is indicated by *p<0.01; **p<0.001. Statistical significance of Mf with respect to N (each subject is his own control) is indicated by *p<0.05. Vertical bars indicate 1 SEM (for N, n=13; Mf, n=6). □ = N, □□ = Mf.
**Muscle triacylglycerol** (TG) was equal among treatments in all sample times. Intramuscular triacylglycerol decreased, as a result of exhaustive exercise, significantly in Mf but not in N. However, it should be mentioned that values in N contained two extreme values (higher value post exercise) most probably caused by subcutaneous fat accompanying the biopsy. Significance was reached in N when these two extreme values were not taken into account. Interestingly there was no difference among the treatments in decrease of intramuscular TG levels as a result of exercise. Twenty four hours after finishing exercise, muscle TG content was not restored to initial levels (Fig 8).

![Graph showing muscle TG content over time](image)

Fig 8 Mean muscle triglyceride content as determined from a biopsy sample, before exercise on day 2, 45 min after exhaustion on day 5, and after 24 h recovery on day 6. Statistical significance with respect to the initial value on day 3 is indicated by $p < 0.05$ and $p < 0.01$. Vertical bars indicate 1 SEM (for N, n=13; Mf, n=6).

**Discussion**

The present study was designed to analyze the effect of repeated long lasting exercise on nutritional indices and metabolic changes and to compare these effects to those present after supplementing
the same diet with a maltodextrin-low fructose beverage while performing the same exercise program. A survey among professional cyclists competing in the Tour de France showed that liquid CHO supplements provided the major part of total CHO intake. Moreover, the study indicated that the athletes were only able to maintain energy balance by using these supplements in addition to the normal conventional meals (Saris, 1988). During a part of this competition the cyclists are competing in the Alps. At this stage finishing on sequential days takes place on the top of a mountain and daily energy expenditure did exceed 35 MJ. It is especially during this extremely intensive part of the competition that the athletes have to cope with exhaustion.

It is known that higher exercise intensities will deplete CHO stores of the body to a low level when performance lasts longer than 45 min and that this is related to a decrease in power output and finally physical exhaustion (Bergström, 1967, 1967a; Sherman, 1984; Saltin, 1967; Newsholme 1983). Cyclists in practice indicate that the only way to prevent this total exhaustion is to ingest large amounts of CHO during the competition. Meanwhile a number of studies showed that time to exhaustion is increased when substantial amounts of CHO are ingested (Coyle, 1983; Ivy, 1983, Edwards, 1984) and that training may enhance the capacity to take up blood borne glucose during exercise (Krzentowski, 1984). In contrast to this it is shown that ingestion of small amounts of CHO may have no effect on performance (Felig, 1982), which suggests that the amount of nutrients ingested may be critical with respect to possible benefits. None of the available studies however has dealt with exhausting exercise over a prolonged period of time during sequential days in highly trained subjects.

The amateur cyclists in the present study were of international level. The athletes cycled 4.5 h per day while simulating two intensive competition days. During this exercise approximately 1 h 20 min were performed at an intensity of 80% Wmax and final exercise to exhaustion was performed at 90% Wmax.

The results of the study show that the subjects ingesting the conventional CHO rich diet at rest and during exercise were not able to maintain energy balance during days of exhaustive physical work in contrast to when the diet was supplemented with Mf (Table 3).

As a result of the supplementation total CHO intake increased significantly as did the relative contribution of CHO to total daily energy intake. From these observations it becomes apparent that
the two main factors influencing energy exchange processes and performance capacity - energy balance and CHO availability - were significantly different between the two treatments and must have had their influence on metabolic regulation and substrate availability. Although the subjects receiving the conventional CHO rich diet were in negative energy balance on day 4 and 5 and the amount of CHO oxidized was substantially greater than the amount of CHO intake, especially on day 4 (for a detailed description see Chapter II and III (Brouns 1988a, 1988b), they did not develop hypoglycemia (Fig 4). A possible explanation may be that the ad libitum food intake during exercise conserved sufficient liver glycogen and/or supplied adequate amounts of CHO to the blood to avoid a fall in blood glucose. A second explanation may be that highly trained individuals have developed an enhanced capacity to synthesize glucose from lactate, glycerol and alanine during exercise.

A contribution of gluconeogenesis to the maintenance of the blood glucose level is supported by the observed increase of hormones that favor gluconeogenesis in the liver and by the observation that blood lactate was significantly increased halfway during exercise and at exhaustion. The fact that maximal blood lactate did not increase above 3.8 mmol.l⁻¹ despite the final exercise intensity of 90% Wmax, indicates that lactate clearance in these subjects must have been substantial. Recently it was shown that lactate turnover is quantitatively larger during continuous exercise than glucose turnover (Brooks, 1986), whereas trained subjects have an enhanced lactate clearance capacity rather than a change in production (Donovan and Brooks 1983). Although it may be assumed that lactate production in the exercising muscle is related to the availability of CHO, such that in a state of CHO depletion less lactate will be produced, no difference could be detected in blood lactate between the two treatments. This occurred despite the large difference in CHO availability (Table 3). Blood glucose was significantly increased during day 4 and 5 in Mf as a result of the high CHO intake whereas it was maintained in a normal physiological range in N. The differences between N and Mf were statistically significant except at the moment of exhaustion on day 4. These differences most probably are due to the ad libitum intake of the CHO beverage started at the onset of exercise, which must have led to a continuous supply of CHO from the gastro-intestinal tract to the blood. The intake of 300 g CHO from the beverage supplemented
during the rest days did hardly influence the blood glucose level. Oxidation and/or storage in endogenous energy depots at same rate as entry in the blood may have been the reason for this. It has been shown that an increase of the CHO fraction in the diet enhances CHO metabolism at rest and during exercise (Yoshida, 1984; Gollnick, 1972) and that blood glucose uptake at the site of the muscle membrane increases with both increasing exercise intensity and blood glucose levels (Saltin, 1973; Berger, 1975, Wahren, 1971, 1971a; Felig, 1975).

Insulin is not required for glucose uptake in the muscle cell during exercise, because of an insulin like factor having the same effect on glucose transport through the muscle cell membrane (Holloszy, 1984). However, it is known that insulin in the presence of muscle contractions has an additive effect on glucose uptake (Ivy, 1981; Ploug, 1984, 1985; Constable, 1985). Therefore it may be assumed that during the present study where both blood glucose and insulin were increased in Mf (compared to N), substantial amounts of CHO will have been available for oxidation in the muscle cell and consequently must have induced glycogen sparing. In studies in which labeled glucose was infused or orally ingested, the observation was made that exogenous CHO is highly oxidized during exercise leading to a sparing of endogenous CHO. It appeared that the amount of CHO sparing seems is related to the dose of the load infused or ingested, i.e. greater loads induce more elevated blood glucose levels and subsequently an enhanced oxidation thereby reducing the degradation of local glycogen pools (Mosora, 1981; Pallikarakis, 1986; Pirnay, 1977, 1977a; Decombaz, 1985)

Sparing of glycogen may take place in the liver and the muscle. Although liver glycogen was not measured it may be assumed that there was no reason to increase liver CHO output from liver stores because of the high amount of CHO ingested. Glucose infusion studies have shown that liver glucose output drops to a low level or is almost entirely blocked when 2 mg.kg\(^{-1}\).min\(^{-1}\) is infused. Although exercise seems to reverse this inhibition, it was observed that during exercise splanchnic glucose output of glucose and saline infused subjects was identical. This indicates that infusion limits the magnitude of rise in glucose output to an amount equal to infusion rate (Felig, 1979). In the present study CHO supply to the vena portae must have been substantially greater in the Mf treatment than the above mentioned infusion rate, both at rest and during exercise.
A statistically significant difference in post exercise muscle glycogen level was observed after exercise on day 5 between N and Mf. Subjects supplemented with Mf showed a significant glycogen sparing, although cycling time to exhaustion during the final 90% Wmax load lasted significantly longer in this group (Table 3). Since the muscle biopsy was taken at approximately 45 min after exercise it can be hypothesized that glycogen resynthesis will have taken place during these 45 min. It has been observed that post exercise glycogen resynthesis can occur from the conversion of lactate (Hermansen 1977; Hultman, 1986). However, blood lactate levels at exhaustion were equal in both N and Mf so that this cannot account for the observed difference. Although a substantial amount of CHO may still have been present in the gastro-intestinal tract in Mf subjects, this can only account for a small part of the observed difference in muscle glycogen. From a recent study (Kuipers, 1987), in which the same beverage was supplied, it was calculated that the average maximal rate of post exercise glycogen synthesis in the trained cyclist amounted to 37 umol.g⁻¹ dry weight.h⁻¹ (range 27-52). This rate was comparable to maximal synthesis rates after exercise as calculated from data of other investigators (Keizer, 1986; McDougal, 1977; Piehl, 1974). Assuming an average maximal post exercise synthesis rate of 37umol.g⁻¹ dry weight only 20% of the observed difference (141 umol.g⁻¹ dry weight) in the post exercise glycogen can be explained by de novo synthesis.

Taking these findings into account it can be concluded that the majority of the glycogen sparing observed must have been net glycogen sparing during exercise. In this respect it is an interesting observation that exhaustion under Mf treatment, which was delayed significantly as a result of CHO supplementation, was not caused by a low level of muscle glycogen. Studies in which glycogen depletion in time was related to the state of fatigue indicate that exhaustion occurs when muscle glycogen drops to a low level (Sherman, 1984; Bergström, 1967; Saltin 1971). In the present study mean muscle glycogen at exhaustion in the Mf group was greater than 250 umol.g⁻¹ dry weight. Thus other factors than glycogen must have played a role in inducing the state of exhaustion.

As discussed earlier, the availability of fatty acids is of great importance for energy metabolism in the endurance athlete because of it's sparing effect on endogenous CHO stores. A high
CHO intake makes a high lipolytic activity superfluous and a low CHO intake or glycogen depletion enhance lipolytic rate via a number of hormones and mediating substrates (Newsholme, 1976, Rennie, 1977). Therefore, the difference in CHO availability between N and Mf must have had its impact on overall fat metabolism.

Glucagon, catecholamines, growth hormone and sympathetic activity all raise lipolysis whereas insulin has an inhibiting effect (Bagby, 1978; Boyd, 1979; Issekutz, 1975).

The hormonal changes in the present study (Fig 6) were all in favor of an enhanced lipolytic activity especially during N treatment where energy balance became significantly negative on both exercise days. Plasma levels of glycerol, fatty acids and ketones increased significantly in this group as a result of the exercise.

The presented values of all hormones and of blood glucose and FFA are absolute values (not corrected for changes in plasma volume) in contrast to the values of metabolic intermediates and end products such as lactate, glycerol, ketones, urea and ammonia which are given as corrected values. The reason is that it is the absolute level of the first mentioned parameters which determines the metabolic effects. With respect to the last mentioned parameters it is particularly important to consider the rate of mobilization or production. Therefore a change in those parameters should be corrected for changes in plasma volume.

Since the uptake of FFA is directly related to the plasma fatty acid concentration (Eaton, 1961; Fritz, 1957) and observed R values declined as a result of exercise, it can be concluded that fatty acids have contributed substantially to energy exchange in N.

The significant increase in plasma ketones may directly be related to the enhanced fat metabolism since a high plasma fatty acid and a low insulin level enhance the rate of ketogenesis (Newsholme, 1976). In Mf fat metabolism was suppressed through the high CHO intake and the related metabolic changes. Although glycerol and FFA increased also significantly in this group, the magnitude of this increase was significantly smaller than that observed in N. An exception to this was the plasma glycerol level at the end of the second exercise day. The fact that a significant increase in lipolysis occurred in Mf, despite the high blood glucose and insulin level, can only be explained by the mutual action of factors that exert inhibiting and/or stimulating effects on lipolytic activity. The inhibiting effects of increased blood glucose and insulin levels may at the very moment have been overruled by the effect of an
increase in catecholamines, glucagon and cortisol. It may be assumed that sympathetic nerve activity, while performing the same exercise protocol, will also have been the same. Interesting is the observation that the plasma glycerol level at exhaustion on day 5 is not significantly different between N and Mf. This coincides with disappearance of the significant differences between N and Mf in blood glucose and insulin despite the large CHO intake from the supplement in the Mf group. Plasma fatty acids on the other hand remained significantly lower under Mf treatment but it has to be kept in mind that plasma fatty acids are a reflection of both fatty acid release and uptake so that it is difficult to conclude about lipolytic activity and/or fatty acid oxidation using this parameter. Despite a small plasma pool, fatty acids may have a rapid turnover rate supplying substantial portions of substrate (Dole, 1956). Glycerol is a better indicator because its conversion to glucose by gluconeogenesis is believed to be slow so that it's accumulation in blood will better reflect overall lipolytic activity (Newsholme, 1983). So far it can be concluded that blood borne fatty acids have contributed significantly to energy exchange despite the high level of CHO intake during exercise.

Intramuscular TG content, as determined from the biopsy samples decreased due to exercise but was not influenced by the diet treatment. Enhanced uptake of blood borne fatty acids and CHO may lead to a sparing of local glycogen (Newsholme, 1976; Jansson, 1984; this study) but this sparing effect was not observed with respect to muscle triglycerides in the present study. In the trained muscle it may be that the release of intracellular fatty acids from local triacylglycerol during exercise depends on a local regulatory factor and that the amount released is always smaller than the oxidative capacity. Fatty acids entering the muscle cell during exercise may then are also be oxidized immediately. This additional fat metabolism may therefore lead to (additional) glycogen sparing but not to a decrease of intracellular lipolytic rate and sparing of TG. The reason why elevated FFA have no sparing effect on intramuscular TG (Stanckiewicz, 1978, the present study) may be partially explained by this, i.e. what is offered is oxidized.

Several studies have indicated that intramuscular fat contributes to a significant extent to overall fat metabolism in the working muscles particularly in the first stage of endurance exercise. Havel (1967) and Paul (1975) calculated in their exercise experiments that approximately 50% of the fat oxidized was
derived from local stores. The observation that trained individuals have a greater capacity to rely on intracellular fat (Saltin, 1983; Holloszy, 1984) and have an increased fat storage in the slow twitch muscle fibers in droplets located near the mitochondria (Hoppeier, 1986) while the fractional extraction of FFA does not increase during exercise induced increased bloodflow suggests that maximal fatty acid transport across the muscle cell membrane in some way exposes a limiting factor for fat metabolism during endurance exercise. There may thus be a physiological need to store more intracellular TG in muscle cells involved in endurance exercise. From the data obtained it may be hypothesized that intramuscular fat degradation during exercise is under direct control of sympathetic nerve activity and that circulating hormones, blood borne substrates and local glycogen levels may only have an additional influence. Some indirect evidence may support this hypothesis.

- Insulin, a strong lipogenic hormone was significantly elevated in Mf compared to N.
- In Mf muscle glycogen was higher than in N.
- In Mf plasma fatty acids were lower than in N.

Nevertheless, muscle TG depletion was similar. Moreover, elevated plasma FFA were shown to have no sparing effect on intramuscular TG (Starckiewicz, 1978), the major part of fat oxidized in the first stage of prolonged exercise is derived from intramuscular stores (Paul, 1975; Carlson, 1971), and finally, if moderate continuous exercise or intensive interval exercise with the same mean power output per hour is performed, muscle TG degradation is found to be the same (Essen, 1977).

**Protein metabolism** is influenced by the concerted action of anabolic and catabolic regulators which are activated or inhibited by physical activity, energy balance, CHO availability and the level of daily protein intake.

It is known that protein turnover is quite sensitive to the immediate energy supply (Waterlow, 1981) and that the most stable turnover rates appear to be present during a state of stable hormone levels and constant supply of nutrients in a fixed ratio such as during the supply of an enteral feeding formula by nasogastric tube at rest (Golden, 1979). Low energy intakes lead to a negative nitrogen balance, whereas high energy intakes have a positive effect on nitrogen balance. (Inoue, 1973; Galloway, 1975; Garza, 1976; Rennie, 1981b). An almost linear relationship
between the level of protein intake and protein synthesis rate has been observed at low levels of protein intake (Meredith, 1982). Das (1972) observed in rats that change from a high- to a low protein diet or vice versa caused an immediate change in urinary nitrogen output which then reached a new constant level after approximately 30 h. This change occurred simultaneously with an adaptation in the activity of urea cycle enzymes. Because exercise induces a negative nitrogen balance by suppressing muscle protein synthesis and increasing protein degradation most probably in the liver and muscle (Rennie, 1981a; Dohm, 1978, 1982; Kasperek, 1980) which may further be influenced by the availability of CHO (Lemon, 1981a, Long, 1976), it was assumed that an accurate estimation of the changes in protein metabolism due to exercise and diet manipulation could only be made if the pre-exercise nutritional and metabolic status were controlled as much as possible. Based on this discussion it was decided to standardize a minimum level of protein intake of 1.2 g. kg\(^{-1}\).day\(^{-1}\) during day 2 and to maintain at least this minimum intake during the entire experimental period. According to Waterlow (1981), in blood the only end products of protein indicating actual changes in protein degradation are urea and ammonia. But for an overall indication of net protein breakdown in the body, resulting in a net nitrogen loss, an accurate analysis of nitrogen excretion in urine and sweat should be performed. During exercise urinary nitrogen excretion falls and plasma urea concentration rises (Rennie, 1981a) most probably because of both a reduced urine production and decreased renal excretion. This leads to an alternative route for elimination of the protein waste products via enhanced secretion in the sweat (Lemon, 1981a). A measurement of both routes is thus essential. The results of the present study show that plasma urea increased significantly, as a result of exercise and negative energy balance, in the subjects with N treatment. In contrast plasma urea remained on a more or less stable level in Mf and tended only to rise at the point of exhaustion on the second exercise day. This difference in urea response is comparable to that described by Lemon and Mullin (1981a) who observed that endogeneous CHO depletion led to a significant increase in urea production during exercise.

Plasma ammonia was not different between the two treatments. The reason for this is not clear. Ammonia may be derived from the deamination of amino acids or from the intracellular adenylate pool. The latter takes place during exercise of high intensity when
immediate energy requirements exceed the amount of energy released from substrate degradation. Rises in plasma ammonia during exercise may be influenced by the availability of local glycogen stores as is observed in McArdles disease (Wilkerson, 1977; Coleman, 1986; Babij, 1983). In the present experiment the subjects cycled until complete exhaustion. It may thus be that AMP deamination has played a role, apart from the involvement of amino acid degradation for reasons of gluconeogenesis, in ammonia production.

Nitrogen losses were affected by exercise, leading to a negative nitrogen balance in N treatment, also during the recovery day. The mean cumulative nitrogen balance over 2 exercise days and the following recovery day was -8.69±2.50 g. In the Mf group there was a tendency to nitrogen sparing despite the significantly longer performance time at 90% Wmax (-2.09±2.50 g). However, most probably due to the large variations this effect did not reach significance. This difference in nitrogen balance between the treatments may entirely be explained by the effect of a negative energy balance and 'CHO balance'. A complete picture of all variables regarding protein metabolism studied in the present experiment will be presented in separate papers (Brouns, 1988c; chapter IV). From the data presented it may be concluded that protein degradation and nitrogen loss from the body is increased in athletes during intense sustained exercise and that this increase can be reduced by measures leading to a maintenance of energy balance and enhancement of CHO intake.

**Hormone levels** were greatly influenced by the exercise. Insulin increased as a result of food intake on day 3 but was significantly lowered by exercise on day 4 and 5. Interesting is the observation that this effect was more pronounced on the second exercise day than on the first. This was not only the case in N treatment but also in Mf. However in N the food induced increase disappeared completely on day 5 indicating a strong inhibition of insulin secretion whereas in Mf food stimulation still remained present. This further decrease on day 5 cannot be explained by the inhibiting effect of catecholamines on insulin secretion (Porte, 1966, 1966a; 1967; Brouns, 1988), since the level of epinephrine and norepinephrine was not different between the two exercise days. In this respect it is tempting to speculate that local factors such as relative glycogen availability indirectly influence insulin secretion. It may be that low glycogen levels in liver and/or muscle initiate an inhibition of insulin release so that the
insulin/glucagon ratio decreases to a low level, in order to enhance lipolytic activity as much as possible in order to spare endogeneous CHO. Plasma glucagon as well as cortisol did not differ between the two exercise days nor between the treatments so that the insulin/glucagon ratio decreased as a result of exercise, to the largest extent on day 5. These findings are surprising in the light of the large difference in CHO availability between the two treatments.

Glucagon, in contrast to insulin has a strong lipolytic effect (Gerich, 1976), increases hepatic glucose output (Felig, 1972; Ahlborg, 1974) and stimulates hepatic gluconeogenesis (Exton, 1972;Clarck, 1974; Garcia, 1966). It is well documented that glucagon levels will rise whenever blood glucose levels fall or whenever local CHO stores become depleted. Several authors have described that an increase in blood glucose due to CHO ingestion during or prior to exercise inhibits glucagon secretion (Ahlborg, 1976, 1977; Luykx, 1978; Felig, 1979). However, in the present study such an effect was not observed.

Cortisol also stimulates hepatic gluconeogenesis (Schrage, 1963; Exton, 1972) and appears to become increased indirectly by adrenergic hypothalamic mechanisms via ACTH secretion (Toivola, 1972). Although it has been described that submaximal exercise may not be associated with increments in plasma cortisol (Schrage, 1963; Exton, 1972) we did observe a significant increase. In general it is known that cortisol release during exercise depends largely on exercise intensity, duration and training status (Keibel, 1974; Bloom, 1976). The increase observed may thus be explained by the submaximal exercise over a prolonged period of time and the related adrenergic activity.

Also catecholamines stimulate liver glycogenolysis, gluconeogenesis and overall lipolytic activity (Landsberg, 1990). Catecholamine levels in plasma increase exponentially with work intensity and are most correlated to the relative exercise intensity. However, physiological deterioration by means of hypoglycemia, hyperthermia or hypoxia may further enhance catecholamine release, especially of norepinephrine (Galbo, 1983; Garber, 1976). As a consequence catecholamine increase is observed to be depressed during exercise trials in which glucose was ingested compared to control (water) trials ( Björkman, 1984; Felig, 1982; Galbo, 1983). The subjects under N treatment, although being in negative energy balance did not develop hypoglycemia.
Mf subjects had significantly increased blood glucose levels but nevertheless showed the same response as N subjects. From these data it may be concluded that hormones which enhance gluconeogenesis, liver glucose output and lipolytic activity, are released during highly intensive and exhausting endurance exercise irrespective of CHO availability. These findings contradict other studies where during less intensive exercise of shorter duration effects due to substrate availability were observed (Galbo, 1983; Viru, 1985) suggesting that a high level of sympathetic activity over a prolonged period of time in man may be dominant with regard to metabolic regulation of substrate mobilization and availability.

Cycling performance quantitated by the time that the subjects were able to perform at 80% and finally 90% Wmax, was substantially effected by the two treatments. The subjects increased their mean final 90% Wmax performance over two days from 9.9±2.6 to 22.4±7.9 min when being under Mf treatment as compared to N treatment. The intramuscular glycogen content, a performance limiting factor as discussed before, may have been responsible for this difference. The subjects in N were not able to restore their muscle glycogen levels within 24 h of recovery (Fig 7). Assuming a similar recovery pattern, glycogen resynthesis during the 18 h of rest between day 4 and 5 will have amounted approximately 75% of the measured value on day 6. This incomplete glycogen recovery may be related to the decreased performance of subjects in N on the second exercise day. In contrast, in Mf performance on day 5 was improved in 5 of the 6 subjects. One subject could not be motivated to sustain the high workloads already at 80% Wmax and quitted prematurely (Fig 9). Of particular interest is the observation that subjects in Mf were able to perform significantly longer but became exhausted without any relation to metabolic changes which normally are related to fatigue such as hypoglycemia, high blood lactate, severe dehydration associated with hyperthermia and glycogen depletion. It might be that after the diet manipulation described a shift occurs from glycogen, as limiting factor (subjects in N), to another unknown factor (Subjects in Mf). It has been suggested that ammonia plays a role in processes of central fatigue (Mutch, 1983) and it is known that ammonia influences a number of steps in the metabolic pathways of energy release (Buono, 1984; Much, 1983, Lowenstein, 1972).
The plasma ammonia levels observed were similar in the two treatments after reaching the point of exhaustion.

![Graph](image)

Fig 9 Individual cycling performance in min at 80% and 90% Wmax on day 4 and day 5.

In summary, the results described lead to the following conclusions:

- The supplementation of a conventional CHO rich diet with a 20% maltodextrin-fructose beverage leads to a marked increase in CHO availability which induces sparing of intracellular glycogen stores and increases exercise time until exhaustion.
- Glycogen supercompensation can be achieved within 24 h post exercise.
- A high CHO intake during sustained exercise reduces protein degradation.
- Local glycogen stores may influence food induced insulin secretion during exercise.
- Intramuscular TG degradation during exercise is not influenced by substrate availability from the blood, nor by intramuscular glycogen content.
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CHAPTER VI THE EFFECT OF CARBOHYDRATE INTAKE DURING WARMING-UP ON THE REGULATION OF BLOOD GLUCOSE DURING EXERCISE


Introduction
It has generally been accepted that the maintenance of a fairly constant blood glucose level is one of the pre-requisites for the ability to perform longlasting intense exercise. It has been shown that performance impairment is often associated with low blood glucose levels and hypoglycemia has been suggested as one of the causes of central fatigue and exhaustion. In 1924 Levine, Gordon and Derick (Levine, 1983) were the first authors reporting that in runners at the finish of a marathon a close relationship was found between blood glucose level and grade of well being; hypoglycemic runners having the most problems. They therefore suggested that ingestion of carbohydrate prior to and during exercise would be of considerable benefit in preventing hypoglycemia.

A few years later however it was observed that instead of maintenance of a desired blood glucose level, a reactive hypoglycemia could occur as a result of pre-exercise carbohydrate feedings (Beje, 1940; Christensen, 1939). Since then many studies have been performed in which the effect of carbohydrate ingestion on substrate utilization has been investigated. The outcome of these studies showed considerable disagreement about the effect and value of carbohydrate ingestion in relation to endurance exercise. This has led to confusion among athletes and their advisors. Analysis of the available literature leads to several conclusions:

1. Intake of carbohydrate drinks after an overnight fast in resting conditions may induce hypoglycemia by a rebound mechanism involving insulin secretion (Bums, 1965; Cahill, 1974) especially during subsequent exercise (Beje, 1940; Christensen, 1939; Costill, 1977; Köivistö, 1987).

2. On the contrary, exercise studies which were carried out in the fed state did not show hypoglycemia after CHO intake at rest (Boné, 1980; Keller, 1984; Keul, 1986; Levine, 1983).

3. Studies in which carbohydrate solutions were ingested immediately prior to exercise showed increased blood glucose and depressed insulin values and no rebound mechanism (Boné, 1980-1981; Keul, 1973).

4. Intake of carbohydrate solutions during exercise enhances blood glucose values while insulin remains low. In (Water) control experiments blood glucose values always tend to be lower (Björkm, 1984; Boné, 1975; Brooke, 1975; Felig, 1983-1982; Flynn, 1987; Luyckx, 1987; Palikarakis,
5. Insulin secretion during exercise is inhibited by catecholamines (Bloom, 1976; Malaisse, 1969; Porte, 1966a-1966b) and/or sympathetic pancreas innervation (Richter, 1984).

It is apparent in the available studies that the methodological set up of a number of studies doesn't simulate the actual situation in sports practice. First of all, the idea to compete after an overnight fast does not seem to be logical compared to the advice generally given that athletes should ingest a light digestible carbohydrate rich breakfast in order to restore liver glycogen. Secondly, there is no reason to ingest carbohydrate at rest prior to exercise because muscle glycogen levels should be high as a result of a high carbohydrate intake during the day(s) prior to the competition. With respect to the last point, the question may be raised why drinks should be ingested at all prior to exercise.

There may be one important reason to do so. During most competitions, especially long distance running (Rehrer, 1988), athletes drink far less during exercise than the amount of fluid that is lost by sweating, leading to dehydration and performance impairment. Therefore, the measure of prehydration becomes very important. After fluid intake there will be a period of overhydration. Whenever this period coincides with the exercise period during which urine production and insulin secretion may be decreased, one may hypothesize that the extra fluid will become available for sweating while the carbohydrates may not elicit an insulin response sufficient enough to induce hypoglycemia, but rather may enhance or maintain blood glucose level.

Because initial gastric emptying and absorption of carbohydrate containing drinks may take place within approximately 7 min (Costill, 1973) the advice should then be to ingest a drink immediately prior to competition or during warming-up, when competition follows immediately thereafter. We therefore decided to study the effect of prehydration with carbohydrate containing drinks in competition-like circumstances in order to test the following two hypotheses:

1. Warming-up induced catecholamines secretion will be insufficient to inhibit insulin secretion after intake of carbohydrate containing drinks.
2. Ingestion of selected CHO containing drinks during warming-up in trained athletes, in the fed state, leads to a reactive hypoglycemia and results in subjective feelings of fatigue.

Methods
Subjects. Eighteen highly trained male amateur cyclists participated in this study. Their mean physical characteristics are presented in Table 1. The maximal oxygen uptake and the maximal achieved watt load were deter-
mined during an incremental bicycle ergometer test in the week prior to the first test session.

**Table 1**  
**Subject characteristics (mean ± SEM)**  
n = 18

<p>| | | |</p>
<table>
<thead>
<tr>
<th></th>
<th></th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td>Age</td>
<td>(yrs)</td>
<td>21.7 ± 4.5</td>
</tr>
<tr>
<td>Weight</td>
<td>(kg)</td>
<td>74.2 ± 9.3</td>
</tr>
<tr>
<td>VO2 max</td>
<td>(ml.kg⁻¹.min⁻¹)</td>
<td>63.2 ± 4.6</td>
</tr>
<tr>
<td>Wmax</td>
<td>(Watt)</td>
<td>390.0 ± 30</td>
</tr>
<tr>
<td>Wmax</td>
<td>(Watt.kg⁻¹)</td>
<td>5.4 ± 0.7</td>
</tr>
</tbody>
</table>

The subjects were asked not to participate in vigorous training or competition on the day prior to the test and to ingest a normal CHO rich diet. All subjects were informed of the nature, purpose, and possible risks involved in the study, before giving their voluntary written consent to participate. The study was done in a cross-over set-up with each subject being his own control.

**Procedure**

The subjects were divided into three groups of six subjects. Each group was studied after at random ingestion of a placebo drink (control) or a selected carbohydrate drink (Table 2).

**Table 2**  
**CARBOHYDRATE COMPOSITION OF SELECTED DRINKS (g/100 ml)**

<table>
<thead>
<tr>
<th></th>
<th>Malto-dextrin</th>
<th>Sucrose</th>
<th>Fructose</th>
<th>Glucose</th>
</tr>
</thead>
<tbody>
<tr>
<td>1 CONTROL</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>2 &quot;FRUC&quot;</td>
<td>-</td>
<td>-</td>
<td>3</td>
<td>-</td>
</tr>
<tr>
<td>3 &quot;SUC&quot;</td>
<td>1.3</td>
<td>6.0</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>4 &quot;MALT&quot;</td>
<td>15.2</td>
<td>-</td>
<td>2.8</td>
<td>-</td>
</tr>
<tr>
<td>5 &quot;GLUC&quot;</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>15.2</td>
</tr>
</tbody>
</table>
The drinks were: a low fructose drink (FRUC) (Isostar- Light®), a sucrose-low maltodextrin drink (SUC) (Isostar®), and a highly concentrated maltodextrin-low fructose drink (MALT) (Perform®). Six at random selected subjects performed one extra test in which a free glucose solution was ingested (GLUC). The set up of the treatments was such that information could be gathered about the effect of different commercially available carbohydrate sport drinks with different carbohydrate sources on the regulation of blood glucose. The carbohydrate composition of the drinks is listed in Table 2.

In SUC the fructose content, derived from the sucrose, is equal to that in FRUC (3g). In GLUC the amount of glucose units is kept equal to the amount present in MALT. This was done in order to compare the effects induced by a glucose polymer - to those induced by a free glucose solution with an equal amount of glucose units.

The subjects came to the laboratory at 7:30 AM after an overnight fast. At 8:00 AM each subject received a standard low fat, high carbohydrate and protein breakfast of normal caloric content (2076 Joules, 13.2 en% fat, 23.8 en% protein, 62.9 en% carbohydrate). The breakfast consisted of solid food (bread without butter or anything else) and a complete nutritional liquid (Powerplay®). Bread was given as 1 g.kg⁻¹/BW and the liquid as 5 ml.kg⁻¹. This was done in order to standardize both food intake and quantitative contribution of nutrients that may exert an effect on insulin secretion. At 9:00 AM a teflon catheter was inserted percutaneously into an antecubital vein. Patency of catheter was maintained by continuous drip of saline solution between insertion and blood sampling.

The subjects remained in resting position on a couch for the whole preparation period. At 10:00 AM exercise warming-up procedure was started on the bicycle ergometer. The warming-up consisted of five min bicycling at 1.5 Watt.kg⁻¹ followed by 20 min at 2.5 Watt.kg⁻¹. At 15, 17 and 19 min each subject performed an acceleration sprint during 10 sec, watt load 5 Watt.kg⁻¹, simulating normal warming-up procedures prior to competition.

All drinks were ingested at 10 min (50% of the drink) and 20 min. Treatments (Table 2) were performed in random order. Warming-up was followed by a resting period of 7 min (preparation time for start and line-up according to competition practice), after which 45 min of exercise was performed at an intensity such that a heart rate of 150 min⁻¹ was maintained continuously (Fig. 1).

Heart rate was monitored throughout the test (Sporttester®, Polar-Electric, Kempele, Finland).
Fig 1  TIME-EXERCISE SCHEDULE
Open arrows indicate blood sampling. Solid arrows represent drink ingestion.
Black triangles represent acceleration sprint. The heart frequency curve of one typical subject is represented.

Blood samples were drawn at rest, before - and immediately after warming-up, immediately before the final exercise period, and at several times during exercise. The Borg scale score of perceived exertion (Borg, 1973) was obtained during the last minute of warming-up and three times during the final exercise bout. In order to establish a possible CHO dose related response the whole procedure was performed twice with ingestion of 300 ml and 600 ml of drink respectively.

Analyses: Glucose was analyzed in plasma (G.O.D.-perid method). Insulin was analyzed by radio-immuno assay (RIA Kit, Medgenix BV, Nederland), catecholamines were analyzed on a HPLC electro chemical detection system (Snedes Kraek, 1982). Perceived exertion was analyzed using the Borg scale.

The data in the text, tables and figures are given as means ± SEM of the delta changes relative to time zero (rest value) for each individual within each treatment group. These delta values were also used for further analyses.

Standard statistical methods were employed using the paired t-test with each subject's control values being compared with values obtained (from the same subject) after ingestion of a selected carbohydrate drink.

A regression analysis was performed to determine possible relationships between insulin on the one hand and epinephrine, norepinephrine or dopamine on the other hand in the different treatments. A stepwise regression was performed to see if the best single correlation could be improved by including
one or both of the other independent variables.

Results

Hormones
Blood insulin levels decreased in control trials as a result of warming-up (Fig 2a and c). At the same time the catecholamine levels were increased (Fig 3a). The ingestion of the 300 ml SUC and FRUC did not influence this blood insulin fall during warming-up. However, when the volume of the drink was doubled a marked reduction of this decrease was observed with the glucose and maltodextrin drink but not with the fructose drink (Fig 2b, 2d).

As a result of the seven min break between warming-up and final exercise, insulin levels increased in all treatments including the control trials. The increase of insulin was more pronounced with the more concentrated carbohydrate solutions (Fig 2b), whereas doubling the dose further potentiated the insulin response (Fig 2d).

We could not detect a quantitative difference in insulin response between the GLUC and MALT trial when 600 ml was drunk (Fig 2d), although insulin response in the glucose trial was initially somewhat faster and significantly increased at all times (relative to the control drink), whereas increase in the MALT trial was only significant at the onset of exercise. Norepinephrine levels rapidly decreased during the seven min break to the same extent in control as in carbohydrate trials (Fig 3a, c). In the control trial epeniphrine showed the same response, but in all carbohydrate trials the epinephrine increase was markedly blunted as a result of carbohydrate intake except for the glucose trial (Fig 3c). Linear regression analysis performed over all times showed that a good correlation existed between response of insulin and norepinephrine but less between insulin and epinephrine (Table 3).

When stepwise regression analysis was performed it was found that, with inclusion of both norepinephrine and epinephrine, the correlation coefficient decreased to 0.71 and 0.62 in the maltodextrin- and free glucose drink trial respectively. During the first part of the heavy exercise period both insulin and catecholamines showed the same response as during warming-up. However, after 20 min of exercise norepinephrine reached a plateau level. From that moment on insulin levels did not further fall. Epinephrine continued to rise throughout the experiment.
The data presented in the graphs represent changes in plasma insulin levels in response to exercise and carbohydrate intake. Values are expressed as the mean ± SEM. Statistical significance is indicated by asterisks: *p < 0.05, **p < 0.01, ***p < 0.001.
Fig 3 PLASMA CATECHOLAMINE CHANGES as result of exercise and carbohydrate intake.
Values as mean ± SEM. Control n=8. Carbohydrate treatments n=6. Control values are given as absolute values. Levels of significance indicate a significant in- or decrease relative to the first value at time zero. The delta changes of the treatments are expressed relative to the first value. Levels of significance indicate the difference relative to the control value of the same subjects.
Fig 4  BLOOD GLUCOSE CHANGES as result of exercise and carbohydrate intake. Values as mean ± SEM.
Control n=8. Carbohydrate treatments n=6. Control values are given as absolute values. Levels of significance indicate an in- or decrease relative to the value at time zero. The delta changes of the treatments are expressed relative to the first value. Levels of significance indicate the difference relative to the control value of the same subjects.
Blood glucose

Blood glucose remained constant during warming-up in both control trials. There was a small rise during the seven min break followed by a decrease below baseline values during the following exercise task (Fig 4a, 4c). In the 300 ml carbohydrate trials blood glucose increased significantly following the SUC and MALT ingestion but not following the FRUC ingestion (Fig). However, when the amount of FRUC was doubled to 600 ml, blood-glucose values were significantly higher than during the control (Fig 4d). In the 300 ml carbohydrate drink trials blood glucose decreased to the values of the control trial after approximately 20 min of exercise and remained stable thereafter. However, after ingestion of 600 ml of the highly concentrated carbohydrate drinks blood glucose values remained above the control values for a longer period of time (Fig 4d).

Hypoglycemia (< 2.5 mmol.l⁻¹) did not occur at all. In the 36 control trials blood glucose fell 7 times below 3 mmol.l⁻¹. In the 42 carbohydrate drink trials blood glucose fell 3 times below 3 mmol.l⁻¹. In both cases the lowest measured blood glucose value was 2.7 mmol.l⁻¹. In the fructose trials blood glucose always remained above 3 mmol.l⁻¹.

The Borg scale score for subjective fatigue did not reveal any difference between the trials.

Discussion

The intake of CHO containing drinks has been promoted in order to maintain or enhance blood glucose values and to spare muscle glycogen so that performance can be maintained at a high level and for a longer period of time. However, intake of carbohydrate solutions in resting conditions after an overnight fast may induce a reactive hypoglycemia (Beje, 1940; Christensen, 1935; Costill, 1977; Koivistio, 1981) which may be mediated by enhanced insulin secretion (Koivistio, 1981).

Such a hypoglycemic reaction does not seem to occur when the exercise is performed in the fed state (Bonen, 1980; Keller, 1984; Keul, 1986; Levine, 1983) or when carbohydrates are ingested immediately prior to exercise (Bonen, 1980-1981; Keul, 1973) or during exercise (Björkman, 1984; Bonen, 1981; Brooke, 1975; Felig, 1983-1982; Flynn 1987; Luyckx, 1978; Pallikarakis, 1986; Pirnay, 1977). The differences may be explained by differences in feeding status, gastric emptying and substraterceptor regulation.

In the fed state gastric emptying may be decreased compared to the fasting state because of receptor feed back from the duodenum due to food substances present (Brouns, 1987). As a result of this and also because of mixing with food remnants in the stomach, gastric emptying of the initial bolus of the drink may be quantitatively less, which will have its consequences for the
magnitude of receptor activation and in turn will affect insulin secretion. Another factor which may induce differences in the regulation of blood glucose may be the difference in the insulin status between the fed and fasted state, as well as the difference in liver glycogen level, which will be substantially reduced after an overnight fast (Hultman, 1981).

Differences in pre-exercise insulin status can affect blood glucose levels during exercise by modification of insulin receptor binding (Berger, 1978). It has been shown that insulin can down or up-regulate its receptor number depending upon previous food intake and related blood glucose and insulin levels (Garrel, 1984; Gorden, 1980; Grundegger, 1982). As a result of this the insulin receptor number at the cell surface will be increased after an overnight fast while insulin levels are low. If carbohydrate is then introduced in the gut and blood glucose increases rapidly it may be assumed that ‘massive’ insulin binding takes place leading to a strongly enhanced blood glucose withdrawal while at the same time glucose output from the liver is largely inhibited by an increase of the insulin glucagon ratio (Felig, 1982; Rizza, 1981; Wahren, 1971). In fact it has been shown that a close correlation exists between fasted pre-exercise blood glucose and insulin levels and the magnitude of blood glucose decrease at the beginning of exercise (Koivisto, 1981).

Whenever blood glucose increases during exercise there will be no strong insulin response because insulin secretion is inhibited as a result of increased catecholamine levels (Bloom, 1976; Malaisse, 1969; Porte, 1966a-1966b). In our study we wanted to test the hypothesis that warming-up prior to exercise is an insufficient stress to cause a catecholamine increase large enough to inhibit insulin secretion. We also wanted to test if ingestion of carbohydrate containing drinks (with differing concentration and CHO sources) in the fed state, during warming-up induces a reactive hypo-glycemia and results in feelings of fatigue during the following endurance exercise.

After analysis of the data we had to reject these null hypotheses. The results demonstrated that the catecholamines increase as a result of warming-up and continued (strenuous) exercise and that this increase is accompanied by a decrease in insulin concentration. By their nature catecholamines are involved in the regulation of rapid adjustments to a changing environment such as homeostatic disturbance. Accordingly, their effects are induced rapidly and dissipated quickly (Landsberg, 1980). From the data it can be seen that a warming-up procedure of 20 min with a heart rate of approximately 130-140 min and including three acceleration sprints is sufficient to cause catecholamines to rise and insulin to fall. During the 7 min break and following strenuous exercise this pattern was subsequently and quickly reversed. The fall in insulin concentration may be explained by the inhibiting effects of both epi-nephrine and norepinephrine on insulin secretion in combination with insulin binding at the site of muscle tissue (Berger, 1978;
Dohm, 1985; Le Blanc, 1979), rather than by changes in insulin clearance which does not seem to be increased as a result of exercise (Franckson, 1971a; Galbo, 1983a).

Innervation of sympathetic nerve endings in the pancreas may also be involved (Richter, 1984). Norepinephrine appears to inhibit insulin secretion in a manner similar to epinephrine. However, when equivalent doses are infused the effects of norepinephrine are quantitatively less (Porth, 1966).

From this finding one may conclude that epinephrine is more important with respect to modifying liver glucose production and blood insulin levels. However, norepinephrine levels increase quicker and reach higher levels than epinephrine which only seems to increase significantly later during exercise when blood glucose levels tend to fall (Galbo, 1975; Harty, 1972; Manhem, 1976). In this study we found the best correlation between the norepinephrine and insulin response (Table 3). This relation is further underlined by the fact that insulin did not fall to lower levels after norepinephrine reached a plateau level after approximately 20 minutes of exercise, at a time that epinephrine further increased.

**Table 3**  Correlation coefficients of insulin and catecholamines in the different treatment groups with insulin as dependent variable.

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Epinephrine</th>
<th>Norepinephrine</th>
<th>Dopamine</th>
</tr>
</thead>
<tbody>
<tr>
<td>Fructose dr.</td>
<td>0.21</td>
<td>0.06</td>
<td>0.19</td>
</tr>
<tr>
<td>Sucrose dr.</td>
<td>-0.30</td>
<td>-0.60 p&lt;0.01</td>
<td>0.06</td>
</tr>
<tr>
<td>Maltodextrin dr.</td>
<td>-0.47</td>
<td>-0.62 p&lt;0.01</td>
<td>-0.15</td>
</tr>
<tr>
<td>Free glucose dr.</td>
<td>-0.33</td>
<td>-0.76 p&lt;0.01</td>
<td>-0.27</td>
</tr>
</tbody>
</table>

This is in line with the findings of Galbo et al (Galbo, 1975) who found that early in exercise insulin levels decreased at the same time as norepinephrine increased and epinephrine remained stable and that variations in epinephrine concentrations during exercise are not accompanied by insulin variations (Galbo, 1977). The fact that the insulin decrease was suppressed after ingestion of the more concentrated carbohydrate solutions may be explained by the fast increase in blood glucose which may have had a stronger effect on insulin secretion than the counter-active hormone during warming-up of this intensity.

During the more intense final exercise this effect was not present anymore. No apparent difference was observed in blood insulin level between the fructose drink and the control supporting the findings that fructose exerts only a
very small effect on insulin secretion (Bohannon, 1980; Crapo, 1980; Kolivisto, 1978).
Although a quick increase in blood insulin occurred during the break after
warming-up, especially in the concentrated carbohydrate trials, we did not
find a reactive hypoglycemia as described by Kolivisto (Kolivisto, 1981).
In all cases insulin fell below baseline levels again within 10-15 minutes of
exercise, whereas blood glucose levels remained elevated for a prolonged
time. The fact that blood glucose remained elevated for a longer period of
time after ingestion of 600 ml boi can be explained by the fact that a phased
gastric emptying takes place so that carbohydrates are introduced into the
gut over a prolonged period of time, especially if gastric emptying rate is
further depressed by higher carbohydrate concentrations (Brouns, 1987).
The fact that hypoglycemia did not occur and that lowered blood glucose va-
values occurred more frequently during the control trials than during the carbo-
hydrate trials underlines the assumption that the intake of carbohydrate con-
taining drinks during warming-up in athletes who are not in the fasted state, is
not detrimental with respect to the regulation of blood glucose and enhances
blood glucose levels as long as carbohydrate absorption from the gut takes
place. Although the cyclists consumed only one bolus of drink during war-
m ing-up it can be assumed that whenever the ingestion of CHO containing
drinks is continued throughout the duration of the exercise period, the fall in
blood glucose levels to baseline levels, as seen after 15-20 min with the low
concentrated drinks supplied in 300 ml boi, will not occur. The validity of this
assumption is underlined by a number of studies (Björkman, 1984; Bonen,
1981; Brooke, 1975; Felig, 1982; Flynn, 1987; Pallikarakis, 1986).

Summary
The effect of carbohydrate drink ingestion on blood glucose regulation
during exercise was tested in eighteen highly trained amateur cyclists.
Ingestion of the drinks took place during warming-up after having consumed
a solid-liquid breakfast.
In contrast to other studies where drinks were ingested after an overnight
fast, in a resting condition, no negative effects on the blood glucose level
were observed.
A close relationship was found between the changes in insulin and the cate-
cholamines, norepinephrine being the most important inhibiting hormone.
Catecholamines increased as a result of exercise while insulin decreased.
The study shows that prehydration with carbohydrate containing drinks du-
ring warming up in the fed state does not affect glucoregulation negatively,
nor subjective fatigue during the following endurance exercise.
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Glucose metabolism during leg exercise in man.
CHAPTER VII  ABDOMINAL COMPLAINTS AND GASTRO-INTESTINAL FUNCTION DURING LONG LASTING EXERCISE

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Abstract.
Over the last several years there has been an increased popularity of endurance sport, with a concomitant rise in ailments which hinder optimal performance. The "ultra" distances, running, bicycling, swimming, skating, and combinations of these, are popular at an elite-competitive, as well as recreational level. Among the problems which are associated with these activities, gastro-intestinal disturbance is one of the most common and one of the most disruptive. Gastro-intestinal distress, at the very least, decreases the pleasure experienced during training or competition and may decrease performance. Disturbances range from mild to severe, from abdominal discomfort to bloody diarrhea. Physical condition, degree of dehydration, and food ingestion prior to or during competition may play a role in the development of these complications. This article deals with the etiology of gastro-intestinal problems in sport and the increased incidence associated with activities where the vertical component of movement is large (i.e. running) in comparison to "gliding" sports such as bicycling, skating, and swimming, where the movement is mainly horizontal and there is minimal jolting. Gastric acid reflux and regurgitation appear to be related to ingestion of food and drink. Bloody diarrhea may occur independent of food and drink and is caused by physiological and morphological changes in the intestinal tract during exercise with dehydration. Suggestions for further research include a systematic study of the changes in the gastro intestinal system resulting from intense endurance-sport activity.

1. Introduction
During the last decade sports physicians have emphasized the intake of liquid during long lasting exercise in order to overcome the problems of severe dehydration and subsequent heat stroke. Coaches and physiologists have
promoted the regimes of carbohydrate (CHO)-loading prior to, and CHO intake during long lasting exercise in order to avoid early glycogen depletion and subsequent exhaustion. Furthermore the occurrence of muscle cramps in the fatigued athlete has led to the advice that body salts that are lost with profuse sweating be replaced. Abdominal cramps or diarrhea often impels the athlete not to eat for 3-5 hours prior to exercise or to ingest complete nutritional liquids. Referring to dehydration, glycogen depletion and loss of electrolytes, Seiple, Vivian and Fox (105) wrote:

"The ideal beverage for athletes
1) should have a rapid gastric emptying rate to provide fast rehydration,
2) should provide minerals to replace those lost in sweat and,
3) should be a substantial energy source."

Taking these points as a beginning for further discussion we are faced with the following questions:
re 1. Gastric emptying is influenced by a large number of factors and fast supply of water and nutrient requires fast absorption in the gut. How will this be achieved, especially during exercise?
re 2. How large is sweat loss and which minerals are lost in sweat in a quantity large enough to consider replacement?
re 3. How much energy is optimal and which energy source is the best for the athlete in which circumstance?

The practical problem is whether or not the intake of nutrients during exercise may lead to abdominal complaints which may hinder the athlete performing optimally. The following discussion tries to answer questions dealing with the occurrence of abdominal complaints, possibly as a consequence of the effect of exercise on the functioning of the intra-abdominal organs.

2. PROBLEM ORIENTATION
2.1 Abdominal complaints
During the last decade long distance running has become very popular for both adults and children. Large numbers of trained and more or less untrained people participate in competitions of 20 km and longer and physicians and first aid teams have frequently become aware of abdominal problems or abnormal facies and urine associated with this type of exercise. Although these complaints are often a matter of discussion and several publications in the sport medical literature have given attention to this topic, it must be stated that research in this field is only fragmentary. The stitch in the side has been known for a long time. However, the process causing the stitch is still open for discussion (114) and needs further research. Post-exercise proteinuria may
last 48 hours and may be related to renal ischemia (4). Nowadays this is not considered to be pathological but is seen as a normal consequence of sports activity. The presence of blood in the urine is of more serious character and may be due to superficial bladder trauma (13). From the foregoing it becomes clear that caution is needed when suggesting causal mechanisms. The presence of hæmaturia may initiate discussions about what a normal consequence of intensive exercise is, as may the statement by Derek Clayton after setting his world record on the marathon in 1979. "Two hours later the elation had worn off; I was urinating large dots of blood and I was vomiting black mucus and had a lot of black diarrhea. I don't think too many people can understand what I went through for the next 48 hours" (Runners World, May 1979: p 72).

Systematic information was gathered by Sullivan (116) who interviewed 57 runners, (running 29-160 km/week). He reports the following results:
- while running, 30% occasionally or frequently had the urge to defecate
- 25% had abdominal cramps or diarrhea during or after competition
- 6% had severe nausea or retching

In 1969 Dancaster described two well trained distance runners who suffered acute tubular necrosis. Both runners had diarrhea during or immediately after the race. It was speculated by Forgores (48) that it is especially the combination of diarrhea and severe dehydration that might be extremely dangerous. The resulting combination: dehydration + hypokalemia + hyperthermia was suspected to cause rhabdomyolysis and acute necroses. Forgores (48) described two cases of long distance runners suffering from loose bowel movements and bloody diarrhea. These problems occurred in periods of very intensive training and competitions. This was suggested to be caused by relative gut ischemia. In one athlete the combination with hyperthermia was present and the result was a bloody diarrhea. Bloody stools have also been reported by Cantwell (24) and Keeffe (75). Hyperthermia due to dehydration has often been reported and may reach values of more than 40 degrees Celsius (32). Such high temperatures might predispose one to ischemic necrosis. Lately Keeffe et al (75) carried out a questionnaire survey on 1700 participants of a marathon. (Response n=707). The results were:
- Lower gastro-intestinal (G.I.) symptoms were more commonly associated with running than upper G.I. complaints.
- The urge to defecate was the most common symptom experienced by runners (36.4% at moderate intensity (M.I.) to 38.6% at high intensity (H.I.) ) and appeared both during and immediately after running.
- Bowel movements (34.9% or diarrhea (19.2%) were relatively frequent immediately after running.
- Runners needed to interrupt runs for bowel movements - 18.4% (M.I.) to 16% (H.I.) - or diarrhea - 8.2% (M.I.) to 10% (H.I.).
- 1.2 (M.I.) 2.4% (H.I.) of runners had bloody bowel movements.
- All lower G.I. symptoms were noted more commonly by women than by men.
- Some symptoms were more reported by younger than older runners.
- Nausea - (11.6% (M.I.) to 12.7% (H.I.) - and vomiting (1.8%) were more troublesome during hard runs or after running.

From these figures it becomes clear that the number of complaints among those athletes is considerable. However, the authors discuss the fact that it cannot be disregarded that primarily those who suffer the most from G.I. problems may have responded. Total response was 40%. The complaints that are related to the intra-abdominal organs may be explained by a combination of two or more of the following factors:
- type of exercise and exercise intensity
- dietary habits (qualitative and quantitative)
- alterations in blood flow and oxygen supply
- alterations in absorption
- hormonal changes
- gastric content and gastric emptying (G.E.)

A few other observations from daily sports practice may be interesting.
1. In sports events where the body is relatively stable, such as in cycling, swimming, speed skating, or cross country skiing the number of abdominal complaints is far less than in running, irrespective of nutritional intake.
2. Training seems to decrease the occurrence of G.I. disturbances.

It is difficult to explain these observations from available scientific data. The pathophysiology of G.I. disturbances associated with running has not been studied and remains speculative (75). In spite of a resurgence of interest in physiologic investigations of marathon running, the G.I. tract has been neglected (80). The lack of knowledge underlines the need for research (48,116,117,114). The athlete, the coach, and the physician are left with a number of questions, which may be summarized as follows:

A. Is there a real need to ingest solid or liquid nutrition during the exercise, if so, what and when?
B. Is it possible to eliminate abdominal problems by ingesting water only or by modulation of the food and drink composition?

With regard to the first question it can be said that there is extensive literature available emphasizing the importance of taking measures for maintaining water, electrolyte and energy (CHO) balance in endurance athletes in order to maintain optimal performance capacity and to reduce the risk of early
exhaustion (For review see: 10,14,66,35,44,102). How, when and which measures should be taken to achieve this deserves further discussion, especially because abdominal complaints may interfere with nutrient intake and absorption. For example the intake of a concentrated glucose solution during exercise may inhibit gastric emptying and induce a water flux into the gastric lumen (this will be explained later). This leads to the feeling of having an overfilled stomach and refrain from drinking. Regurgitating of food and the reflux of gastric juice often occurring in this situation are very unpleasant events. An overview of the main problems of food intake such as quantity, quality, time, etc. related to exercise is presented elsewhere (19). The second question initiates a discussion about the logic of the advice not to eat and drink prior to and during the exercise, or about possibilities to modify the composition of the food and/or the fluid to be taken in such a way that the risk of causing problems will be minimized. Indications for possible modifications may be derived from the available literature in the field of gastro-intestinal, splanchnic, and renal physiology. A brief discussion of the importance of water and/or carbohydrate intake in relation to endurance exercise will follow.

2.2 The need for fluid intake
Between 1931 and 1966 twenty-six football players died from hyperthermia and dehydration (113). In the last decade numerous cases of heat stroke due to dehydration have been reported during long distance running. Costill (32) reported a rectal temperature of a non-finisher, following a marathon in the heat, at 41.3 degrees Celsius. During a 2 hour laboratory running trial, with a workload of 70% VO2 max and no water ingestion, the weight loss in a runner was 4.02 kg and rectal temperature increased to 40.6°C. During the Olympic marathon trial in 1968 Costill measured a mean body weight loss of 6.1 kg. Such body weight losses are not only present in elite athletes setting their top performances but occur in general whenever long lasting exercise is executed at an intensity that is high for the individual in question. Dressendorfer (41) reported a mean sweat loss of 3 liters in five trained coronary patients running a marathon. Depending on temperature, humidity, sunshine, clothing, wind and altitude the fluids lost can be substantially greater. Saltin reported a sweat loss of 7 liters during exercise lasting 3.5 hours (103). It is obvious that such fluid losses have to be replaced in time. Practical experience has shown that this is possible during cycling. Saris et al. (104) measured fluid intakes of more than 10 liters on a hot competition day in the 1984 Tour de France. Fluid intake was closely related to external temperature.
Fig. 1  Water intake during the Tour de France.

Fig. 2  Water intake in relation to environmental temperature.
During running, however, drinking seems to be problematic. Running initiates movements in a vertical plane, which makes drinking from a cup difficult. Breathing frequency, which is normally related to stride frequency (some fixed ratio e.g. 3 strides inhaling - 3 strides exhaling) is also disturbed, which is unpleasant. As a result total fluid intake per hour during running is much less than during cycling which initiates dehydration and heatstress more frequently. In this context it is difficult to understand how elite marathon runners sometimes perform so well with very little fluid intakes (less than 200 ml), while others run into serious problems.

2.3 The need for carbohydrates
Controlled studies have shown that muscle glycogen availability is a strong limiting factor for long lasting exercise of high intensity (10,35). The higher the intensity, the more CHO metabolism is stimulated. Energy flow from CHO is faster than from fat (90). Thus the prerequisite for maximal, endurance exercise is the availability of CHO since energy flow from fat only enables 50-60% of maximal power output (94). The problem of CHO depletion and subsequent exhaustion can be avoided or delayed by taking the following measures:

1. Enhancement of CHO intake 3 to 4 days prior to exercise in combination with a training volume reduction, so called "glycogen loading during tapering off"
2. Intake of CHO during exercise
3. Adequate training in order to increase exercise induced fat metabolism

Point 1 may lead to a maximal glycogen content in liver and muscles at the onset of exercise, whereas 2 and 3 may lead to glycogen sparing. In general it can be stated that the intake of CHO during exercise will be beneficial. However, fluids and nutrients have to be tolerated and it should be realized that they first become available after passage of the stomach. For better understanding it will therefore be necessary to discuss some details of normal G.I. functioning and regulation first. The influence of exercise on the G.I. tract and the possible relation to the development of disturbances and abdominal complaints will be discussed after that.

3. GASTRO INTESTINAL FUNCTIONING AND REGULATION AT REST
3.1 Gastric emptying (G.E.)
The pylorus is opened most of the time (letting liquid pass through unhindered and closes only in cooperation with the terminal antrum and some duodena contractions (42,115). This is evidenced by the finding that the passage o
liquids remains the same after pylorectomy or placing a transpyloric cannule (40,115). It seems that the tone of the proximal part of the stomach determines to a large extent the moments for emptying of liquid and half solid substances (61) and that the motor activity of the distal segment determines the passage of solid particles (84). The complex of contraction and relaxation of proximal and distal gastric compartments in close cooperation with pyloric action is regulated to a large extent by a number of stimuli resulting from:

- osmolality
- caloric density
- type of carbohydrate
- fatty acids
- amino acids
- acidity (pH)
- particle size
- meal volume
- meal temperature
- dietary fiber
- hormones
- other factors

A summary of research in which these effects have been studied is given in tables 4-8 at the end of this review. In the following paragraphs the most important factors will be discussed.

3.1.1 Intra-gastric regulation of gastric emptying
Particle size, meal volume, and meal temperature exert an intra-gastric effect on G.E. These factors will be described first.

3.1.2 Particle size
Using labeled meals, Moore (88) measured the G.E. of solid and liquid parts. The solid parts were emptied at a rate of 2 gm/min. Chicken liver given as large cubes emptied more slowly than small cubes due to "grinding down" and "liquefying" work done by the stomach (64). Bernier (11) measured emptying of two identically composed meals, one solid, one homogenized. The homogenized meal emptied faster (4 hours vs 6 hours). It seems that the pylorus refuses particles larger than 1mm to pass (11,84). Fink (46) showed that complete liquid meals (12.4-14% protein, 31.4-37.4% fat, 49.8-54.7% carbohydrate) emptied by 70% within the first hour and almost completely within two hours. The liquid meal emptied substantially faster than the solid meal. (This might lead one to question the value of ingesting solid nutrition in order to enhance energy intake during long lasting exercise, especially because the ability to chew and to swallow may be impaired during higher exercise intensities and onset of food availability to the intestines will last several hours). However, in the fasted subject, G.E. of both solids and liquids is not immediately inhibited after ingestion (when duodenal receptor control is still inactivated). During this phase rapid G.E. occurs and leads to the escape of larger nutrient particles (11) after which G.E. becomes inhibited and stomach digestive activities are stimulated. Thus, regulation of G.E. seems to
function in a bi-phasic response in which particle size control is initiated subsequent to duodenal receptor control. At the onset of food intake, the gastro-duodenal cooperation can be considered as an "open loop" in which the initial effluent depends primarily on gastric activity and little upon any inhibitory influence of the duodenum. This means that a bolus will be emptied immediately after the initial intake (rapid emptying phase). Once received in the duodenum the food will influence local processes such as hydrolysis and receptor activation or inhibition. Once the receptors are activated they will exert their effect on G.E. resulting in a phased emptying (18).

![Graph showing gastric emptying of homogenized meal](image1)

![Graph showing gastric emptying of solid meal](image2)

*(Homogenized)*

*(Solid)*

Fig. 3 Gastric emptying of identical solid and homogenized meals.
Adapted from Bernier (1985).
Testing procedures
From this information one may speculate about the legitimacy of analyzing G.E. by serial measurements in between which the stomach is washed with water or saline and gastric residue is aspirated by a G.I. tube. It can be assumed that this procedure has its limitations, since it is primarily the composition of the duodenal content that determines gastric emptying of liquids and not the composition of the gastric content. It would therefore be more appropriate to have a longer time between the trials in order to "wash out" the duodenum before the next trial is performed. Unfortunately most studies done have not taken into account this fact. Probably the best procedure is to do a post-trial emptying test of saline and to start the next trial at the moment that saline empties in a normal rate again, suggesting that post pyloric inhibition is reduced to a desired "standard" level. Another point of interest is that a recent study (2) indicated that the presence of a gastro-intestinal tube delays gastric emptying and accelerates bowel transit. The development of non-invasive techniques thus seems the most appropriate way to study G.I. function in the future.

![Flowchart]

Fig. 4 Dynamic closed loop feedback system.
Fluid ingestion

OPEN LOOP

I. Rapid emptying phase

- volume
- contractile characteristics of stomach
- particle size

Bolus size dependent on

Duodenal receival + feedback control

II. CLOSED LOOP
Phased bolus emptying

Fig. 5  Fasted subject - stomach and duodenum are empty.

3.1.3 Meal volume
Volume effects of G.E. Hunt (67) showed that the rate of G.E. increased 3.3 ml for every 100 ml increase in gastric content. Costill (33) found a value of 3.7 ml for each 100 ml increase with a maximum at a volume of 600 ml. It is hypothesized that the increased stomach wall pressure leads to wall distention which stimulates gastric motility and subsequent G.E. This might be an exponential function (71).

3.1.4 Meal temperature
The studies on the effect of meal temperature on G.E. have all been done with liquids. Gershon-Cohen (55) showed that decreased thermal temperatures are associated with an increase in gastric motility and a rapid flow through the jejunum. As a result of this cold drinks tend to leave the stomach earlier than do warm drinks (38). Lower temperatures enhance G.E. When a drink of 5°C was ingested, the gastric rest volume after 15 min was 50%. However, when the temperature of the same drink was 35°C the rest volume was 73% (33). It may be that these effects on G.E. can be counteracted by changes in duodenal receptor activity. Other factors exerting an effect on the regulation of G.E., are located in the gut or are of more central character such as hormones.
3.2 Gut regulation of Gastric Emptying

The most important site in the regulation of G.E. is the duodenum which contains a large number of receptors sensitive for osmolality, acidity, fatty acids, mono- and di-glycerides, amino acids and carbohydrates (9,31). Stimulation of these receptors delays G.E.

3.2.1 Osmoreceptors

Hunt and Pathak (68) hypothesized that solutions of glucose isosmotic with plasma are thought to make the duodenal osmoreceptor permeable to sodium. This leads to receptor swelling (penetration of solute + sodium), a signal to allow G.E. Whenever the concentration of solutes in the duodenum is too high (and thus is hypertonic) sodium and water will be drawn from the receptor vesicle and subsequent shrinking will take place, a signal for inhibition of G.E. If the latter takes place, the duodenal content will first be adjusted to isotonicity, resulting in subsequent increase in receptor permeability, before a new bolus may leave the stomach. Meeroff (82) stated very clearly: “The rate of emptying is not influenced by non isotonic solutions in stomach or jejunum as long as duodenal contents remain isotonic”. Hunt (68,70) reported that isotonic saline solutions empty more rapidly than water. The findings of Hunt suggest that pure water entering the duodenum will be adjusted to isotonicity too by electrolyte fluxes (from hypo-isotonicity) or that slow water diffusion may be the stimulus for decreased G.E. compared to saline. Ruppin (101) found that the osmolality of a liquid meal correlated with its G.E. rate (higher osmolalities - slower G.E. rates), as does fluid and nutrient absorption (20,109). Foods always leave the duodenum with an osmolality of 300 m.osmol/l (11). It is suggested that the two most important ways to realize this adjustment to isotonicity is a synchronized process of water secretion and nutrient absorption. Water secretion also may take place in the stomach but even though gastric secretion may dilute the gastric content, the emptying rate of a hypertonic solution remains the same because of direct duodenal control (18).

From this it is concluded that osmoreceptors are not present in the stomach or intestine beyond Treitz ligament. Since isotonic drinks have been promoted for rapid fluid replenishment there have been a lot of controversial discussions about their effectiveness and value. There is no doubt, however, that in the gut water together with nutrients is absorbed as isotonic solution and that after liquid intake, there will be an adjustment towards isotonicity which subsequently will influence G.E. Water alone can enter the blood stream by simple diffusion. In the jejunum the solute absorbed is isotonic (112). In experiments in which constant perfusion with a triple lumen catheter was used it was found that the osmolality of the fluids down the jejunal test segment was unaltered despite different flow rates of test solutions. Water absorption took place together with solute in isotonic proportions. Also the human colon
absorbs water and sodium from an isotonic medium (79). However, literature on absorption of nutrients is conflicting. Single nutrients can be absorbed from a hypertonic medium in contrast to what is believed for water absorption (see also section 3.8). If water absorption is the primary factor for rehydration in the athlete then adjustment to isotonicity may play a limiting role. Theoretically, there are three ways to realize this:

1. secretion of water and/or electrolytes into the G.I. lumen
2. absorption of nutrients
3. duodenal escape of nutrients

Apart from effects exerted by osmoreceptors, there is evidence that receptors responding to specific nutrients may exert the strongest effect on the regulation of G.E. Glucose, fatty acids and amino acids may be important to consider in this respect. Another subject for discussion may be the absorption of these nutrients in the duodenum and jejunum. Because glucose, fructose and amino acids as single nutrients have been tauted as important for sports performance, many athletes ingest these nutrients in a purified form. The question then arises whether glucose taken as a concentrate is absorbed with a maximal speed and if not, whether for example sodium has to be added to the nutrient to increase absorption. Some of these interactions will be described more in detail in section 3.8.

3.3 Water secretion
Water secretion as a result of increasing osmotic gradient between the G.I. lumen and the blood has been described for the individual G.I. sections.

3.3.1 Stomach
Foods that are hypertonic at the onset of ingestion will leave the stomach partly as iso- to hypotonic bolii at the end of the digestive process (101). A similar situation has been described in the canine stomach (5). Hypertonic solutions increase gastric secretion (18,46). Costill (33) suggested that, despite the movement of a hypertonic solution out of the stomach at the onset of ingestion, gastric residue may increase due to gastric secretion. Glucose polymers (having a lower osmolality) induced a smaller secretion than free glucose (33).

3.3.2 Duodenum and jejunum
Sessions (106) found that infusion of 300 ml hyperosmolar (50%) glucose solutions directly into the duodenum led to the so-called "dumping syndrome". This was associated with a plasma volume decrease and a large intestinal water increase in order to reduce intestinal osmotic pressure (106, 21). These changes in the upper intestine did not take place if the solution was infused into the stomach. Launiala (77) described that unsplit di-saccharides led to
water secretion in the gut. The amount of water secreted depends on the amount of undigested disaccharides.

3.3.3 Colon
Increased water movement into the G.I. lumen due to an osmotic gradient has also been described for the colon (12). With hypertonic solute perfusion the water absorption ceased when osmotic pressure of the solute was 350 mOsm/kg. In that occasion water was secreted. This was also the case with rising osmolalities of infused mannitol solutions.

3.4 Nutrient absorption
The second proposed way to adjust to isotonicity is via the absorption of solutes (25). After ingestion of a labeled diet (\(^{3}\)H\(_{2}\)O and U\(^{14}\)C Sucrose) rates of tracer appearance in blood plasma suggested that both water and carbohydrate absorption were accelerated with increasing osmolality. By 30 minutes apparent \(^{3}\)H\(_{2}\)O absorption from a 700 mosm/kg diet was three times greater than that absorbed from a 250 mosm diet. Water absorption is closely coupled to that of total solute since glucose absorption can stimulate water absorption directly (20,49,51,111). Water absorption is also observed with fructose (51) and leucine (1). This absorption does not exclude the possibility that water secretion can take place at the same moment so that the end result of inward and outward H\(_{2}\)O flux is a net water increase in the gut. A more detailed description of the effects of CHO absorption on gut regulation of G.E. is given in section 3.7.

3.5 Duodenal "escape" of nutrients
If the duodenum is the prime site of regulation then it is clear that foods after leaving this section loose their influence on the receptors located in the duodenal lumen. Ruppin (101) performed an experiment in which 400 ml test solutions were infused into the stomach. The solutions used were 1. polycose (a glucose polymer), 2. po-lycose + sucrose, 3. sucrose. All had an identical caloric content. Osmolality, however, was different. The volume emptied from solution 1. was significantly larger during the first 80 minutes. Osmolality of duodenal contents was equal for all meals at Treitz ligament. From these findings they suggested that the glucose polymer was less completely hydrolyzed than the sucrose at the site of duodenal receptors and more in the proximal jejunum. Mollison (87) described that patients with severe deficiency of pancreatic amylase had normal inhibition of gastric emptying after a glucose load. However, boiled starch solutions left the stomach nearly as quickly as water. Presumably the starch passed the osmoreceptors in the duodenum and exerted no large osmotic effect because hydrolysis and subsequent osmolality increase did not take place at this regulation site. The use of polymers in
nutritional liquids may thus be of advantage compared to the use of nutrients in their mono-form because reduced osmolality may minimize dumping syndrome-like reactions (81,110).

Recently this principle has been applied to the composition of sportdrinks. Studies have been performed to test the effect of glucose polymer ingestion on G.E. Since hydrolysis of glucose polymers is not rate limited and the duodenum has the capacity for rapid absorption (15,101), it is assumed that ingestion of a glucose polymer might induce a larger quantity of carbohydrate to be emptied from the stomach and thus lead to a larger energy contribution to the body. Two studies support this assumption. Seiple (105) showed that G.E. of a 7% glucose polymer solution was not significantly different from cold water. Foster (52) found that ingestion of a 5% glucose polymer solution resulted in 69% more fluid and 33% more carbohydrate deliverance to the small intestine than from a 5% free glucose solution. The underlying factor making this possible might be the earlier proposed "escape" of a part of the polymers to the upper jejunum. However, the available studies should be interpreted with care since the tests have been carried out at rest and not during intensive exercise.

Summarizing the foregoing we can conclude that the bio-availability of nutrients and liquids to the intestinal system is dependent on the rate of gastric emptying which in itself is regulated to a large extent by the chemical composition of the duodenal luminal content.

3.6 Acid receptors

pH receptors are assumed to be present at the site of G.E. regulation, the duodenum. Foods leave the stomach with a pH between 5 and 2 depending on the type of food ingested. At the end of the duodenum pH is systematically 7 pH neutralization is a consequence of the absorption of H+ ions and alkaline secretions from the pancreas and duodenum (11). Adjustment to pH 7 takes time and therefore G.E. will be inhibited by pH deviations of the duodenal content (107).

3.7 Carbohydrate receptors

The type of CHO ingested may act on the regulative site of G.E in two ways:
1. specific single nutrient effect on the receptors.
2. indirect osmolality effect due to rapid hydrolysis.

Specific effects
Especially glucose tends to be a very potent inhibitor once present in the duodenum. In a study of Coyle (36) plain water emptied 39% faster than a preparation containing 4.6 g dl-1 of free glucose. This is in line with earlier
studies (49,68,70), in which water was compared to glucose-saline solutions. Elias (43) found that glucose is more effective in G.E. inhibition than galactose or fructose. Subjects showed little slowing of G.E. with concentrations of fructose up to 200 mosmol compared to glucose. At higher concentrations this difference remained although to a lesser extent. From these findings it was concluded that fructose is relatively ineffective in slowing G.E. This confirms the findings of McHugh and Moran (89).

![Graph](image)

**Fig. 6** Gastric emptying of 150 ml meals, 0.05 g/ml fructose and 0.9% NaCl. Moran, T. and McHugh, P.R. (1981): Distinction among three sugars in their effects on gastric emptying and satiety. Am. J. Physiol. 241: R25-R30.
Table 1 Gastric emptying rates on test meals

<table>
<thead>
<tr>
<th>Test meal</th>
<th>Conc (g/ml)</th>
<th>N</th>
<th>mosm</th>
<th>ml/min</th>
<th>Emptying rate g/min</th>
<th>Emptying rate kcal/min</th>
</tr>
</thead>
<tbody>
<tr>
<td>Glucose</td>
<td>0.05</td>
<td>9</td>
<td>291</td>
<td>1.76 ± 0.27</td>
<td>0.088 ± 0.01</td>
<td>0.352 ± 0.05</td>
</tr>
<tr>
<td></td>
<td>0.25</td>
<td>16</td>
<td>1.425</td>
<td>0.381 ± 0.04</td>
<td>0.095 ± 0.01</td>
<td>0.377 ± 0.03</td>
</tr>
<tr>
<td>Fructose*</td>
<td>0.05</td>
<td>20</td>
<td>293</td>
<td>3.52 ± 0.42</td>
<td>0.18 ± 0.02</td>
<td>0.72 ± 0.08</td>
</tr>
<tr>
<td></td>
<td>0.125</td>
<td>24</td>
<td>721</td>
<td>1.36 ± 0.14</td>
<td>0.17 ± 0.02</td>
<td>0.68 ± 0.08</td>
</tr>
<tr>
<td></td>
<td>0.25</td>
<td>12</td>
<td>1.426</td>
<td>0.679 ± 0.08</td>
<td>0.17 ± 0.02</td>
<td>0.68 ± 0.08</td>
</tr>
<tr>
<td>NaCl*</td>
<td>0.009</td>
<td>20</td>
<td>290</td>
<td>3.86 ± 0.62</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

N. No. of test meals from which the regression was derived. * emptying rates although exponential are expressed as average over time.


Table 2 Volume of 400 ml meal emptied from the stomach in 5 min.

<table>
<thead>
<tr>
<th></th>
<th>Saline 0.9% (290 mosm/l)</th>
<th>Meal constitution Glucose 0.2 kcal/ml (290 mosm/l)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Subject 1</td>
<td>259 ml</td>
<td>69 ml</td>
</tr>
<tr>
<td>Subject 2</td>
<td>150 ml</td>
<td>80 ml</td>
</tr>
</tbody>
</table>

Brener et. al. (1983): Regulation of the gastric emptying of glucose. Gastroenterology 85: 76-82.

It has been suggested that in particular glucose loads quickly initiate the phased G.E. by closed-loop control (18). The delivery of glucose seems to be subject to a specific tight regulation and depends on the amount received by the duodenum directly. The resulting "steady state" of G.E. then is a balanced outcome of two reciprocally interdependent functions: gastric delivery and its post-pyloric inhibition by glucose. Di- saccharides (once present in the duodenum being hydrolyzed very rapidly) are approximately twice as effective per mole as mono-saccharides, assuming that the molar concentration at the moment of ingestion is the same (43). After CHO ingestion intraluminal
enzymes are secreted in excessive amounts to hydrolyse starch (7,37,47), or maltose or sucrose (56). The breakdown of carbohydrate is not the rate limiting step for glucose uptake (119), hydrolysis of CHO is usually more rapidly than absorption (124). Thus the inhibiting effects by receptor activation may be maintained to a larger extent by a relatively slow absorption which may differ depending on the CHO source. For example, the absorption of lactose is rate limiting (57) which may explain the complaints of diarrhea always present in cases of lactose intolerance or with high lactose intakes. Glucose is absorbed actively and fructose passively and there is no competitive inhibition between glucose and fructose in their absorption (63). Passive fructose absorption is believed to be slow. In the jejunum it is half as quick as glucose. (57).

Andersson Nygren (6) performed a study in which 50 grammes of fructose was given orally either in 10% or 20% solutions. Ten out of 14 subjects showed incomplete absorption (71%) with the 20% solution and 6 out of 10 with the 10% solution (60%). Complaints during this period of malabsorption were gas, abdominal cramps, and diarrhea. When the dose was reduced to 37.5 gm only 2 subjects out of 14 had malabsorption with the 20% solution and there was no malabsorption with the 10% solution. After ingestion of 25 gm no problems were reported. There may thus be an upper limit of approximately 30 gm. Some studies indicate that glucose absorption from sucrose may be enhanced by the presence of fructose, a so called di-saccharide effect (118). This is further underlined by the fact that glucose and fructose ingested in the monomer form raise plasma triglycerides significantly less than an equivalent amount of sucrose. With equimolar intake of starch and sucrose the blood glucose after starch feeding was the lowest (95). The speed of absorption may play a central role in exerting osmotic effects at the site of regulation. If absorption is inadequate osmotic diarrhea may follow. At the same time G.E. will be inhibited.

It may be concluded that carbohydrates exert their duodenal mediated effect on G.E by:

- the type of carbohydrate ingested
- the concentration

This effect may be mediated by:
- factors underlying hydrolysis - enzymes
- kinetic differences
- active/passive processes
- competitive actions
- stimulating interactions
- water and electrolyte fluxes
3.8 Electrolyte Interactions
With increasing osmolalities, water and electrolytes are secreted into the G.I. lumen. Sodium, potassium, calcium and magnesium are found to be increased in gastric residues in such conditions (36,105). Electrolytes are necessary for hydrolysis and absorption. The hydrolysis of sucrose is sodium dependent (26). The translocation of glucose by a bifunctional carrier, with two binding sites, one for Na\(^+\) and the other for glucologues (D-glucose of D-galactose), also requires sodium. A one-way or two-way interaction between water and sodium as being coupled to glucose absorption has been discussed. Glucose absorption stimulates water absorption and water movement across the mucosa stimulates passive absorption of Na\(^+\) by solvent drag (50). Sladen-Dawson (111) suggested that glucose stimulates both water and sodium absorption by the jejunum and that water absorptions from isotonic solutions is ordinarily coupled to that of total solute. Water follows passively in isomotic proportions. Brown (20), on the other hand, suggested that glucose can stimulate water absorption directly without mediation of sodium and that glucose follows at a rate which maintains isotonicity. Fordtran (51) hypothesized that these two mechanisms may act at the same time:
1. Glucose stimulates active sodium transport, water follows
2. Glucose stimulates water transport, sodium follows
Summarizing this information it may be postulated that water movement, as coupled to glucose, may have an optimum at a certain concentration and will cease if glucose absorption ceases or if electrolyte fluxes are inadequate. This information may play a significant role in the discussion about optimal intake and absorption of water, water plus electrolytes, or water, electrolytes and CHO, depending on the athletes immediate needs determined by the sport event and climatological circumstances. One factor seems to be of special interest here: active transport is dependent on the integrity of the sodium-potassium pump. This pump requires energy (ATP) in order to work. Thus depriving cells of energy (i.e. anoxia, energy depletion) or inhibition of the sodium-potassium pump will inhibit glucose transport (26). If transport ceases then G.E. may be inhibited and osmotic diarrhea may follow. The question thus arises whether or not oxygen and energy available to the epithelial gut cell is adequate during higher exercise intensities and different levels of dehydration.

3.9 Caloric or Energy Density
Another proposed and some what "confusing" mechanism for the regulation of gastric emptying is the caloric density of the food leaving the stomach (91). Starch and glucose in isocaloric solutions leave the stomach at the same rate (68,72), although the osmolalities of the solution are different. A progressive slowing of G.E. correlated with increasing calories is found. This was not influenced by the fat, CHO, or protein content, or by the initial volume of the
meal ingested. Moore (88) showed that G.E. of mixed solid and liquid meals differ, dependent upon the weight and caloric content. Meals with larger weight and caloric content had slower emptying rates for both solids and liquids. Isoaloric casein, mol-oil, and glucose empty at the same rate, independent of the changes in osmolality at onset of ingestion (91). Regulation is thought to be direct via the closed loop feed-back system from the duodenum.

If meals composed of solids and liquids are homogenized then the rates of emptying are similar (61). There are, however, indications that caloric control may be different for pure solids and liquids. Solids may be primarily under caloric and particle size (84) control and liquids primarily under combination of other factors such as acidity, osmolality and single nutrient composition. Bernier (11) found that t1/2 for G.E. of liquids is around 40-50 min after ingestion, although the caloric content varied from 0-550 calories. On the other hand isotonic NaCl drinks had a t1/2 of 14 min compared to milk (also about isotonic) with a t1/2 of 62 min.
Moran (89) showed in Rhesus monkeys that glucose and D-xylose empty linearly and more slowly with increasing concentration. Delivery to the duodenum is constant at 0.1g/min or 0.4 Kcal/min, regardless of concentrations, osmolality, or volume of the intragastric meal. Fructose, however, emptied exponentially and more rapidly than the other sugars: 0.2g/min or 0.8 Kcal/min. Brener (18) compared the effect of saline solutions and glucose solutions on G.E. Even the most dilute glucose solution (0.2 Kcal/ml), comparable to the osmolality of saline, only emptied 63% of 400 ml in 20 minutes, compared to 84% of saline (glucose receptor or energy density control?). However, when the same quantity of glucose calories (with widely different osmolarities) was introduced into the duodenum, then the inhibition times of G.E. was similar. The number of glucose calories passed per unit time remained the same over a five fold concentration range (2.13 Kcal/min). These findings are in line with McHugh (92) and Moore (88) who found 4.6 Kcal/min passed for large meals in humans. The quantitative figures are different, possibly because of differences in the meal consistency (i.e. liquid, half solid, solid) and differences in species. From the discussions about osmotic- and energy density control one may raise the question about the effectiveness of the duodenum to reduce the activating signals to absorb nutrients quickly. The faster the absorption is the less the influence will be of nutrients that induce osmolality or energy density stimuli. One might also discuss the validity of an energy density concept, since increasing energy density also means an increased content of nutrients that will exert an inhibiting effect on the receptors located in the duodenum. From this it may be hypothesized that the energy density effects found are mainly the result of a cumulative effect of
different specific nutrient receptor activities.

4. THE EFFECTS OF EXERCISE ON GASTRO-INTESTINAL REGULATION

4.1 Oxygen and energy availability in the G.I. tract
Tissues which may undergo large changes in activity and energy expenditure are for their energy supply to a large degree dependent upon blood flow, substrate and oxygen availability. It is hypothesized that under local hypoxic circumstances energy liberation in the G.I. mucosa cells may be impaired.

4.2 Blood flow
Exercise causes a quantitative redistribution of tissue blood flow. The working tissues (skeletal muscles) undergo a vasodilatation to increase their blood flow, while the G.I. tract undergoes a vasoconstriction. As a result, splanchnic and gastric blood flow will be diminished (16,53,76,121). Clausen (29) reported that during maximal exercise in both trained and untrained people, bloodflow to the gut is reduced as much as 80%. Rowell (100) found that splanchnic blood flow during workloads of 70% $\dot{VO}_2$ max is only 30-40% of the resting value. However, at submaximal loads splanchnic blood flow in the trained is better than in the untrained, which may be explained by metabolic adaptation of the body in such a way the same sub-maximal exercise induces less stress. This also becomes evident from the lower heart rate during submaximal exercise after a period of training. Studies on CHO absorption in dogs showed that absorption was diminished when blood flow decreased (120,123). If this happens, G.E. will be inhibited due to malabsorption and subsequent increase of osmotic pressure in the duodenum. From these findings it may be hypothesized that the problems of diarrhea present after carbohydrate intake during high intensity work may be related to malabsorption. The cause of diarrhea during long distance running, independent of prior food and/or fluid intake, may be a change of the epithelial tissue in the gut due to necrosis caused by hypoxia. Anoxia or hypoxia may lead to lesions of the inner surface of the gut. These lesions may also explain the occurrence of blood or metabolites from blood in the feces of long distance runners. Schaub (108) described a case of a 33 year old marathon runner with complaints of nausea, urge to defecate, and diarrhea (sometimes bloody) during high intensity training sessions or competitions. Colonoscopic inspection showed epithelial surface changes such as are known to occur during gut ischemia. These lesions were predominantly present in the caecum and colon. Because bleeding, pain, and diarrhea are the normal symptoms in a situation of gut ischemia, it was concluded that hypoxia of the lower G.I. sections may induce the problems.
4.3 Exercise and hormonal influences

An important factor in the regulation of gut tissue activity and metabolism at rest is hormonal control. Hormonal regulation at rest is strongly influenced by the activity of the sympathetic and parasympathetic nervous system. A summary of hormonal influences in the G.I. tract in normal resting conditions is presented in Table 3.

<table>
<thead>
<tr>
<th>Hormone</th>
<th>Secretion</th>
<th>Motility</th>
</tr>
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<tbody>
<tr>
<td></td>
<td>H⁺</td>
<td>pancreas enzymes</td>
</tr>
<tr>
<td>gastrin</td>
<td>↑</td>
<td>↑</td>
</tr>
<tr>
<td>CCK-Pz</td>
<td>↑↑</td>
<td>↑↑</td>
</tr>
<tr>
<td>secretin</td>
<td>↓↓</td>
<td>↑</td>
</tr>
<tr>
<td>G.I.P.</td>
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<td>0</td>
</tr>
<tr>
<td>V.I.P.</td>
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<td>↑</td>
</tr>
<tr>
<td>motilin</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>glucagon</td>
<td>↓</td>
<td>↓</td>
</tr>
</tbody>
</table>

[↑ ↑] slight increase
↑ increase
[↓ ↓] slight decrease
↓ decrease
0 no effect


However, the influence of quantitative changes of G.I. hormones during exercise is poorly understood. Sympatho-adrenal activity changes during long lasting exercise. Enhanced concentrations of catecholamines have been found during dynamic exercise. Higher workloads, induce larger increases (exponential). The concentration of norepinephrine in plasma varies inversely with the oxygen saturation of mixed venous blood. Hyperthermia and hypoglycemia are related to enhanced catecholamine concentrations. All these factors play a role during long lasting exercise in the untrained, as well as in the trained individual during maximal exercise. (For a complete review see the extensive work of Galbo, 54). Catecholamines, by their classical actions, would delay G.E. and prolong transit time (73,99). However, this is not in line with the increased G.E. of solid meals during prolonged exercise of
moderate intensity in untrained man (22). It can be hypothesized, however, that this inhibiting influence of increased catecholamines may be counter-effected by an enhanced influence of other hormones such as the gastro-entero-pancreatic hormones. Feldman (45) found that serum gastrin levels after a steak meal were significantly enhanced by exercise. He suggested that this might be explained by the exercise induced catecholamine release, since these are capable of increasing serum gastrin levels in humans (28). This gastrin increase was also confirmed by other studies (17,96). However, in Feldman’s (45) study the increased gastrin levels did not influence secretion. Perhaps this was counter effected by a release of substances that inhibit gastric acid secretion such as somatostatin and vaso intestinal peptide (60,117). Plasma motilin levels also increase during long lasting exercise and this may account for the regular bowel function and enhanced gut transit of food which is assumed to be the result of light endurance exercise (116,117). The motilin levels raise to a level which is normally seen after ingestion of a mixed meal and which that stimulates gastric emptying, and gut motility (117). The reason why these increases occur during exercise remains unknown. It has been hypothesized however, that the decreased splanchnic bloodflow during exercise might lead to a decreased hepatic and renal clearance of the peptides with a resulting plasma level increase.

The effect of hormones on G.I. function during exercise remains obscure.

4.4 Endorphins
Endogenous opiate-like agents may influence G.I function. As a result of endurance exercise endorphins are released into the peripheral blood (3,58). They may interact with opiate receptors present in the gut (97) to influence gastro-intestinal motility. In general however, opiates tend to delay gastric emptying (27), so that the conclusion that endorphins will influence G.I. motility remains very speculative.

5. EXERCISE RELATED STUDIES
Most studies on exercise and G.I. function have been done with liquids. Only a few studies deal with solid or homogenized food. An overview of these studies is presented in table 9. Fordtran-Saltin (49) studied four men and one woman during one hour of treadmill running at an exercise intensity of 70% \( \dot{V}O_2 \) max. They found no effect on G.E. of a 13.3% glucose, 0.3% sodium chloride solution, and a slight inhibition when pure water was ingested. Intestinal absorption of water in both jejunum and ileum was studied with poly-ethylene glycol (a non absorbable marker) which was supplied together with the test liquid by constant infusion. There was no difference in absorption of water, glucose, and electrolytes, compared to the resting situation. The luminal
content of these substances was measured at the beginning and at the end of the gut test segment by triple lumen catheter. Costill (32) performed a two hour treadmill test at the same intensity (70% \( \dot{V}\text{O}_2 \) max) with four highly trained marathon runners. Total fluid ingestion was 2000 ml, consisting of either a glucose-electrolyte solution (glucose 4.37 g/dl, potassium 96 mg/l, sodium 460 mg/l, chloride 536 mg/l) or water. No post-exercise gastric residue differences were found. Feldman (45) studied G.E. of a blended steak meal in untrained persons after 45 min of exercise at either 50 or 70% of their maximal workload G.E. after a period of two hours (including the exercise trial) was 87%, compared to 88% for the control group (no exercise). Assuming that the rise in serum triglyceride concentration after a fat containing meal is a reflection of lipid absorption, he concluded that fat absorption was unimpaired. Cammack (22) studied G.E. and gut transit time of solid meals labelled with technetium. G.E. was measured at 10 min intervals by determining gastric radioactivity. Gut transit of the "head" of the meal was defined as the time that passed between ingestion of the meal and the first consistent increase in breathing hydrogen concentration (caused by CHO fermentation in the gut). Exercise (cycling) significantly accelerated G.E. but had no effect on transit time. Exercise intensity however was low: heart frequency 117/min, pedaling rate 33/min. In terms of \( \dot{V}\text{O}_2 \) max this is equivalent to approximately 30% \( \dot{V}\text{O}_2 \) max. G.E. time (t1/2) of the control group was 1.5 ± 0.1 hours, for the exercise group 1.2 ± 0.1 hours. Possibilities for the reduction in of G.E. proposed were:

1. relative dominance in parasympathetic tone
2. increase in endogenous opiates
3. decreased gastric acid secretion (23)

Points 2 and 3 are questionable, since endorphins may inhibit G.E. as discussed earlier, and gastric acid secretion was found to be unchanged in a more recent study (49).

Very recently the effect of a 6 weeks running programme (3 times 30 min at 70-80% \( \dot{V}\text{O}_2 \) max/week) on bowel transit time was determined (30). The authors found a 22.8% reduction in transit time but could not define the causative mechanism. It was hypothesized that an exercise-induced increase in sympathetic nervous system activity may cause a relaxation of the G.I. tract which may facilitate the passage of contents from colon to rectum, in particular during running exercise which has an additional mechanical effect due to up and down motion. From the studies available it may be concluded that exercise intensities up to 70% \( \dot{V}\text{O}_2 \) max do not influence G.E. or absorption in the gut and that G.E. and bowel transit may be enhanced, compared to testing conditions. The fact that some studies show contradictory results may be due to differences in training status, pre-exercise feeding status or order of serial testing (section 3.1.2).
6. CONCLUSIONS AND SUMMARY

If the studies cited are analyzed critically one may conclude that the experimental situation (exercise intensity and duration), and degree of dehydration and hyperthermia, never reflected the situation in sports practice in which G.I. disturbances and complaints occur most frequently (see introduction). In Fordtran’s study the exercise time was only 1 hour, so that the combination of dehydration, hyperthermia and high intensity exercise which is expected to cause G.I. problems, was not present. In Costill’s study, however, this combination was present, but energy density (CHO content) and hence the possible inhibition of G.E. by duodenum, was minimal. In Feldman’s study (45) exercise duration was only 45 minutes. In the study of Cammack (22) the subjects performed a 3 hour cycling test as part of the total test trial which lasted 6 hours. However, exercise intensity was very low as discussed before. Thus, the available research does not answer the question: to what extent will absorption of a single nutrient such as CHO be changed in a situation of dehydration, hyperthermia and minimal G.I. bloodflow, as has been demonstrated in dogs (120,123)? If we summarize the sections described here we may conclude that G.I. complaints occur in relation to long lasting, exhaustive exercise.

Further, that complaints seem to be present to a larger extent during running than during the more “gliding” sports such as cycling, swimming and cross country skiing. From the available data we have the impression that symptoms such as regurgitation, gastric acid reflux and vomiting may be related to the composition of the gastric content together with changes in motility. However symptoms such as diarrhea or the occurrence of blood in the feces may be related to functional and morphological changes in the gut tissue primarily. To what extent mechanical factors (type of exercise) play an important role has to be elucidated. Much data is available on G.I. function at rest, however, G.I. functioning during long lasting, high intensity exercise has only been studied incidentally. The influence of exercise induced hormonal changes, especially of the G.I. hormones, cannot be explained at the moment.
Suggestions for sports practice

- Solid food should be avoided during the last 3 hours prior to exercise.
- Liquid foods can be taken as pre-competition meal and also during exercise, however, fat and protein should not be included.
- Whenever fluid intake is of first priority, drinks should be low in CHO.
- When maximal intake rates of both CHO and water is desired the optimal concentration should be in the range of isotonicity.
- When CHO intake is the first priority drinks may be concentrated and hypertonic. A combination of glucose polymers with fructose may be advantageous.
- Athletes suffering frequently from diarrhea or the urge to defecate may benefit from complete nutritional liquids (low in fiber content) during the last day preceding the competition, so that gastro-intestinal contents will be minimized.
- Drinking during exercise should be part of the training programme.

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Table 4 Observed effects of gastric content on gastric emptying

<table>
<thead>
<tr>
<th>Variable</th>
<th>G.E. rate</th>
<th>Author</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>Increase ↑</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Decrease ↓</td>
</tr>
<tr>
<td>Particle size</td>
<td>↑</td>
<td>Holt 1982, Meyer 1980</td>
</tr>
<tr>
<td>Temperature</td>
<td>↓</td>
<td>Gershon-Cohen 1940, Costilli 1974</td>
</tr>
<tr>
<td>Volume</td>
<td>↑</td>
<td>Hunt 1951, 1968, Costilli 1974</td>
</tr>
<tr>
<td>Weight [bulk] (solids)</td>
<td>↑</td>
<td>Moore 1981</td>
</tr>
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Table 5 Possible effects of duodenal content on gastric emptying

<table>
<thead>
<tr>
<th>Variable</th>
<th>G.E. rate Increase ‡</th>
<th>G.E. rate Decrease ‡</th>
<th>Author</th>
</tr>
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<tbody>
<tr>
<td>Osmolality</td>
<td>‡</td>
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<td>Barker 1974, Simko 1976</td>
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<td></td>
<td></td>
<td>‡</td>
<td>Meerof 1975, Cooke 1977</td>
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<tr>
<td>Glucose content</td>
<td>‡</td>
<td>‡</td>
<td>Hunt-Pathak 1960, Hunt' 1961</td>
</tr>
<tr>
<td></td>
<td></td>
<td>‡</td>
<td>Fordtran 1967, Elias 1968</td>
</tr>
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<td></td>
<td></td>
<td></td>
<td>Coyle 1978, Brener 1983</td>
</tr>
<tr>
<td>Amino acid content</td>
<td>‡</td>
<td>‡</td>
<td>Barker 1974</td>
</tr>
<tr>
<td>Fatty acid content</td>
<td>‡</td>
<td>‡</td>
<td>Cooke 1977, Bernier 1985</td>
</tr>
<tr>
<td>Acidity</td>
<td>‡</td>
<td>‡</td>
<td>Shapiro 1950, Bernier 1985</td>
</tr>
<tr>
<td>Caloric density</td>
<td>‡</td>
<td>‡</td>
<td>Hunt 1960, Hunt-Stubs 1975,</td>
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<td></td>
<td></td>
<td></td>
<td>McHugh-Moran 1979, Moore 1981,</td>
</tr>
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<td></td>
<td></td>
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<td>Brener 1983</td>
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<tr>
<td>Sodium content</td>
<td>‡</td>
<td>‡</td>
<td>Hunt' 1960, Barker 1970</td>
</tr>
<tr>
<td>Potassium content</td>
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Table 6 Possible effects of duodenal processes on gastric emptying

<table>
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<th>Factor</th>
<th>G.E. rate</th>
<th>Author</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Increase↑</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Decrease↓</td>
<td></td>
</tr>
<tr>
<td>Enzyme availability</td>
<td>↑</td>
<td>Mollison 1968</td>
</tr>
<tr>
<td>Absorption rate</td>
<td>↓</td>
<td>Gray 1975</td>
</tr>
<tr>
<td>Absorption capacity</td>
<td>↑</td>
<td>Andersson Nygren 1978</td>
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</table>

Table 7 Possible factors influencing duodenal absorption

<table>
<thead>
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<th>Factor</th>
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</tr>
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<td></td>
</tr>
<tr>
<td></td>
<td>Decrease↓</td>
<td></td>
</tr>
<tr>
<td>Sodium</td>
<td>(carbohydrate)</td>
<td>Hirschhorn 1980</td>
</tr>
<tr>
<td></td>
<td>(sodium)</td>
<td></td>
</tr>
<tr>
<td>Splanchnic bloodflow</td>
<td>↓</td>
<td>Williams 1964, Varro 1975</td>
</tr>
<tr>
<td>Sodium pump inhibition</td>
<td>↓ (glucose)</td>
<td>Caspary 1984</td>
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</table>
### Table 8 Some hormonal factors influencing gastric emptying

<table>
<thead>
<tr>
<th>Hormone</th>
<th>G.E. rate</th>
<th>Author</th>
</tr>
</thead>
<tbody>
<tr>
<td>Catecholamines</td>
<td>↑ ↓</td>
<td>Jenkinson 1967, Rees 1980</td>
</tr>
<tr>
<td>Gastrin</td>
<td>↑ ↑</td>
<td>Feldman 1982</td>
</tr>
<tr>
<td>Motilin</td>
<td>↑ ↑</td>
<td>Sullivan 1984</td>
</tr>
<tr>
<td>Endorphins</td>
<td>↑ ↓ ?</td>
<td>Chapman 1950</td>
</tr>
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</table>

### Table 9 Effect of exercise on gastric emptying

<table>
<thead>
<tr>
<th>Liquids During</th>
<th>Gastric Emptying Method</th>
<th>Author</th>
</tr>
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<tbody>
<tr>
<td>light exercise</td>
<td>unchanged</td>
<td>Ramsbottom 1974</td>
</tr>
<tr>
<td>moderate exercise</td>
<td>unchanged</td>
<td>Costill 1974</td>
</tr>
<tr>
<td>moderate exercise</td>
<td>accelerated</td>
<td>Campbell 1928</td>
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<tr>
<td>moderate exercise</td>
<td>delayed</td>
<td>Hellenbrandt 1934</td>
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<tr>
<td>moderate exercise</td>
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<td>Neuer 1986</td>
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<td>moderate exercise</td>
<td></td>
<td>Ramsbottom 1974</td>
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<tr>
<td>severe exercise</td>
<td>unchanged</td>
<td>Fordtran 1967</td>
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<tr>
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<td>delayed</td>
<td>Costill 1970</td>
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<tr>
<td>severe exercise</td>
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<td>Campbell 1928</td>
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<td>severe exercise</td>
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<tr>
<td>severe exercise</td>
<td></td>
<td>Costill 1974</td>
</tr>
<tr>
<td>Solids during moderate exercise</td>
<td>unchanged</td>
<td>Cammack 1982</td>
</tr>
<tr>
<td>&quot;halfsolids&quot; during moderate and severe exercise</td>
<td>unchanged</td>
<td>Feldman 1982</td>
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<tr>
<td>&quot;halfsolids&quot; during moderate and severe exercise</td>
<td></td>
<td>serum T.G. measurements</td>
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CHAPTER VIII: GENERAL DISCUSSION AND CONCLUSIONS

'You are what you eat' and 'eat to win' are popular statements from the athletic world which have had a large influence on the sporting population during the last decade.

Nutrition has become important with respect to performance at the time that athletes have reached their limits in training volume and -intensity. This has led to a renewed interest among athletes, coaches and exercise physiologists in the role of nutrition and the influence of gastro-intestinal problems on physical performance and well being. Clear cut nutritional counseling, however, is often difficult for a number of reasons. First of all there is no generally accepted nutritional recommendation for athletes involved in heavy physical training. In contrast, most countries have accepted standards for the recommended daily allowance (RDA) of nutrients, given for different age groups, sexes, as well as different levels of physical activity. The RDA for people involved in heavy physical work is normally derived from studies done among miners and lumberjacks. Information about the daily requirements of athletes, who with respect to the relative work intensities differ from miners and lumberjacks, is lacking.

Another reason for the absence of clear nutritional advises for athletes is the diverse outcome and conflicting results present in the large number of studies that have been done on the influence of specific nutrients on metabolism and performance. Although such differences often can be explained by the wide variation in age, training status, duration, intensity and type of exercise as well as the dose of nutrients given, it makes generalization difficult.

A third reason is the lack of information from controlled studies about the causes of gastro-intestinal problems during exercise. This situation leads to a diversity of opinions often based on subjective experience and/or misinterpretation of available studies.

In the present work an attempt was made to analyze the dietary problems that occur in the case of strenuous exertion. The two main goals were:

1) To gather information about diet and nutrition problems in the elite athlete under field circumstances.

2) To perform experiments in which the effects of diet manipulation on nutritional and metabolic parameters could be studied in a laboratory setting as close as possible to the practical situation.

The results of the available literature show that athletes have altered eating patterns and have an increased number of in between meals on days when training is intensive and resting hours are limited. As a result, foodstuffs are selected that are often energy rich but low in micronutrients. The conse-
quence may be a decreased nutrient density, with the risk of long term
marginal intakes of micronutrients, whereas fat intake may be too high (see
Chapter I). Therefore it should be advised to teach the athletes about the
selection of high quality foods. A prerequisite for adequate food selection
however is the availability of the foods desired in the immediate surrounding.
Especially sport canteens at training centers should therefore adjust the foods
items offered toward a variety of carbohydrate (CHO) and dietary fiber rich
foods among which full grain products, fresh fruits and vegetables.

Apart from this problem it has been observed that an adequate energy intake
from conventional CHO rich foods may become impossible on days at which
energy expenditure is increased above 20 MJ and athletes cope with
physical exhaustion. The results from the present work (Chapter II) give
evidence that a high CHO content of the diet per se is not a guarantee for an
adequate CHO and energy intake during days of hard sustained physical
exercise. In this case the addition of liquid foods to the normal meals during
long lasting exercise has proved to be efficient with respect to maintaining
energy balance and improving performance (Chapter II).

The observation that athletes who consumed a conventional CHO rich diet,
with a mean protein intake of 1.7 g.kg\(^{-1}\).day\(^{-1}\), were not able to maintain
nitrogen balance, questions the validity of an extrapolation of the RDA for
protein intake, derived from studies on 'normal' populations, towards athletes
working at the upper limits of human performance.

The fact that diet manipulation on the one hand led to a maintenance of
energy and CHO balance but on the other hand did not lead to a
maintenance of nitrogen balance whenever protein intake was less than 1.5
g.kg\(^{-1}\).day\(^{-1}\) further provides evidence that the daily protein requirements of
athletes involved in heavy sustained exercise are higher than the RDA value
for normally active people and will be in the range of 1.5 to 1.8 g.kg\(^{-1}\)
(Chapter III and IV). However the present data do not elucidate whether a
possible adaptation to a 'relative' low protein intake occurs when exhausting
exercise is repeated over a prolonged period of time. Such an adaptation
was observed, as a result of training, when non exhaustive exercise was
performed (Gontzea, 1975).

Measures to increase CHO intake are beneficial for performance. The relation
of quantitative CHO intake to the amount of CHO stored and physical
performance capacity has been proven in a large number of studies.

In the present work it was shown that performance was significantly increased
by the addition of a CHO rich liquid to an already CHO rich conventional diet
(Chapter V). It was further demonstrated that additional CHO intake leads to
protein sparing and enhances recovery of the glycogen stores in the first 24 h
after exercise.
An increased consumption of mono- and di-saccharides in the daily diet has been a matter of discussion during the last 20 years as being a risk factor for the development of cardiovascular disease and of insulin resistance with subsequent diabetes (Davidson, 1985). The advice given to athletes to ingest CHO concentrated liquids in addition to the normal diet or as a substitute for normal food during exercise may therefore appear conflicting. There are however a number of reasons to suggest that the supplementation of CHO beverages in the exercising athlete is not harmful in this respect. It is observed that a high CHO intake during exercise, especially mono- and di-saccharides, does not lead to a large increase in plasma insulin (chapter V1). From this observation and from the studies that showed that exogenous CHO is readily oxidized during exercise it is hypothesized that a high CHO intake during exercise is not harmful with respect to the development of insulin resistance. Extra CHO intake at rest, if desired, should be limited to supplementation at the end of a normal meal in order to realize a smooth gastric emptying and to avoid large fluctuations in plasma insulin. High plasma insulin levels after intake of CHO solutions at rest, in the fasting state, as well as a decreased physical performance whenever exercise is started at the moment of hyperinsulinemia, have been observed in several studies. However, the situation as well as the nutritional status in which the athletes were tested in these studies differed with respect to the situation in sports practice. (Chapter VI). The present work provides evidence that the ingestion of CHO containing liquids in the fed state during warming-up has no harmful effect on the blood glucose level nor on subjective fatigue during exercise. In contrast, blood glucose can be maintained at levels significantly above control values when ingestion is done properly. With respect to dietary counseling of athletes it should therefore be stressed that CHO containing beverages should only be ingested at the end of warming-up whenever this is followed by exercise or 5-3 min prior to the start, when warming-up is not possible. Fructose as an alternative CHO source has been suggested as being of advantage because of its insulin independency. The argument is that fructose does not trigger insulin secretion and thus allows maximal lipolytic activity while at the same time supplying CHO. In the present work however it was shown that supplementation with a CHO rich liquid, 50% of which consisted of free fructose, resulted in a significantly increased CHO metabolism and decreased fat metabolism (Chapter III). Moreover, the subjects receiving this dietary supplement had the largest protein degradation as measured from nitrogen excretion. These results, supported by the recent work of Tappy et al (1986), contradict the hypothesis of enhanced fat metabolism after fructose intake compared to glucose intake. To what extent food and fluid intake during exercise leads to the development
of abdominal problems is not entirely clear (Chapter VII). It is generally accepted that exercising with a full stomach leads to gastric distress. The advice given therefore is that the last meal should be taken at least 3 h prior to exercise. However with respect to the abdominal problems occurring during exercise this may be too short because it has been observed that a normal meal has not passed the stomach within 5-7 h depending on its composition. The finding that homogenization of a meal causes a marked reduction in gastric emptying time and that liquid meals poor in dietary fiber and fat have a high emptying rate is of importance with respect to the ingestion of liquid meals prior to exercise or during exercise.

It has been observed that gastric emptying and food absorption in the gut is not changed with exercise intensities up to 70% \( \dot{V}O_2 \) max. However most of the studies have been done during cycling exercise and data from running experiments are lacking. It has been suggested by coaches that the occurrence of abdominal complaints and diarrhea in runners may be related to poor absorption. However, the observation that this occurs mainly in runners and not in cyclists suggests that the type of movement may be an important factor in the etiology of gastro-intestinal disturbances. It can be hypothesized that changes in gastro-intestinal motility due to increased retox activity of the intestinal wall, caused by the pounding actions during running, may trigger the occurrence of intestinal cramps and that ischemic necrosis due to severe dehydration may cause the observed bloody diarrhea in the runner. Until now no controlled studies dealing with this problem are available, which indicates a need for the initiation of research in this field.

The results of the work presented here deal mainly with dietary problems related to energy consumption and energy exchange. Although the role of micronutrients has briefly been discussed in Chapter I, it cannot be concluded from the data presented to what extent micronutrients may have played a metabolic role in the exercise situations described, nor what the daily requirements under these circumstances are.

In conclusion the most important observations in the present work can be summarized as follows:

- elite athletes have changed eating patterns and consume 30-40 % of total energy intake as small in-between meals consisting mainly of energy dense foodstuffs.
- with increasing energy expenditures there is an increased selection of foods and liquids that are high in energy, sweet, and convenient;
- there is an energy threshold of approximately 20 MJ per day, above which athletes are unable to consume enough normal food in order to maintain energy balance.
- This threshold occurs mainly through a decreased appetite and a limited tolerance of the stomach to a large food intake under exercise circumstances.
- For the degree of exercise intensity described, the normally recommended CHO intake of 60 en% is not sufficient to maintain "CHO balance".
- The supplementation of a CHO containing beverage in addition to the normal meals and during exercise is shown to be an effective measure to maintain energy - and "CHO balance" at higher energy expenditure levels.
- Muscle glycogen supercompensation can be obtained within 24 h after exhausting exercise by the supplementation of a CHO beverage to the normal diet.
- Under the exercise circumstances described a protein requirement of > 1.5 g.kg⁻¹.day⁻¹ is required to maintain nitrogen balance.
- When energy balance is negative, 1.7 g of protein .kg⁻¹.day⁻¹ is insufficient to meet the needs.
- An increased CHO intake induces protein sparing.
- Despite a high CHO intake during exercise and the resulting significantly increased blood glucose levels, lipolysis is stimulated by the execution of exhausting exercise.
- From the data gathered and the literature presented it is tempting to speculate that fructose ingestion during exercise causes some change in intermediate metabolism leading to a stimulation of CHO metabolism and inhibition of fat oxidation.

It is suggested that for future research in the field of nutrition and exercise the study of elite athletes should be conducted under controlled practical situations.

There is a need to gather data about the daily requirements of nutrients for this specific population. However, the participation of these highly gifted men and women in research will probably be one of the most limiting factors for the attainment of data

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HOOFDSTUK 8 Samenvatting en konclusies

"You are what you eat" en "eat to win" zijn populaire uitspraken afkomstig uit de wereld van de topsport welke gedurende de laatste 20 jaar een flinke invloed hebben gehad op de naar fitheid strevenende mens. De voeding is een belangrijk aspect bij het presteren geworden nu atleten met betrekking tot trainingsvolume en -intensiteit hun limiet hebben bereikt. Dit heeft bij trainers, coaches en inspanningsfysiologen geleid tot een hernieuwde interesse in de rol van voeding op het lichamelijk presteren en welzijn.
Het verstrekken van uniforme voedingsrichtlijnen is om een aantal redenen echter vaak moeilijk.
Op de eerste plaats bestaat er geen algemeen aanvaarde voedingsaanbeveling voor atleten die aan zware lichamelijke training onderhevig zijn. Daarentegen hebben de meeste landen erkende tabellen met dagelijkse aanbevolen voedingskwantiteiten voor verschillende leeftijdsgroepen, sexe en niveaus van lichamelijke inspanning. De aanbevelingen voor mensen met zware lichamelijke arbeid zijn meestal verkregen uit onderzoeken welke uitgevoerd zijn bij mijnwerkers en havenarbeiders. Informatie over de behoeften van atleten, die met betrekking tot de relatif lage arbeidsintensiteit verschillen van mijnwerkers en havenarbeiders ontbreekt.
Een andere reden is dat verschillen, en soms de strijdigheid tussen, de vele resultaten die verkregen zijn uit onderzoeken naar de invloed van specifieke voedingsstoffen op de stofwisseling en het prestatievermogen. Alhoewel dergelijke verschillen vaak verklaard kunnen worden door de grote variatie in leeftijd, trainingsstatus, duur, intensiteit en type van de inspanning alsmede de hoeveelheid voedingsstoffen welke ingenomen is, is generalisatie op grond van deze resultaten moeilijk.
Een derde reden is het gebrek aan informatie uit goed gecontroleerd onderzoek omtrent de oorzaken van maagdarmproblemen bij lichamelijke inspanning.
Deze situatie leidt tot een diversiteit van opinies, vaak gebaseerd op subjectieve ervaring en/of foutieve interpretaties van de beschikbare gegevens en onderzoeken.

In het voorliggende werk is een poging ondernomen om de voedingsproblemen, welke inherent zijn aan lichamelijke inspanning te analyseren.
De twee belangrijkste doelen waren daarbij:

1. Het verzamelen van informatie over de voedingsgewoonten en -problemen bij goed getrainde atleten in hun dagelijkse situatie.

2. Het uitvoeren van experimenten, waarbij het effect van voedingsmanipulatie op voedings en metabole parameters kan worden bestudeerd in laboratoriumomstandigheden welke de praktijksituatie zoveel mogelijk benaderen.

De beschikbare resultaten uit de literatuur tonen aan dat atleten andere eetgewoonten hebben en een toegenomen aantal tussenmaaltijden gebruiken op dagen dat er zeer intensief getraind wordt en de tijd voor rust en herstel beperkt is. Omdat voor dergelijke tussenmaaltijden vealal vette en zoete voedingsmiddelen gekozen worden, met een laag gehalte aan micronutriënten, is een achteruitgang van de "nutrient density" (de aanwezigheid van vitaminen, mineralen en sporenelementen in relatie tot het energiegehalte), met het risico van marginale micro-nutriënten inneming aanwezig, terwijl anderzijds de vetconsumptie hoger is dan wenselijk (zie hoofdstuk I).

Het is daarom aan te bevelen om sportmensen te adviseren over de dagelijkse keuze van de juiste voedingsmiddelen. Een voorwaarde voor zo'n keuze is echter de beschikbaarheid van de betreffende voedingsmiddelen in de onmiddellijke omgeving. Daarom moeten in het bijzonder de sportkantines en de trainingscentra erop toe zien dat er een gevarieerde keuze mogelijkheid bestaat voor KH- en voedingsvezelrijke voedingsmiddelen zoals volkoren produkten, vers fruit en groenten.

Naast dit probleem toont de praktijk dat een adekwate energie inneming door middel van normale voedingsmiddelen onmogelijk kan worden gedurende dagen waarop het energieverbruik meer dan 20 MJ (5000 kcal) bedraagt en de sporter blootstaat aan lichamelijke uitputting. De resultaten van het voorliggende onderzoek (hoofdstuk II) bewijzen dat een hoog koolhydraatgehalte van de voeding op zich geen garanti is voor een voldoende inneming van KH en energie gedurende dagen met zware langdurige inspanning. De toevoeging van vloeibare energiekoncentraties aan het normale voedsel tijdens het verrichten van langdurige lichamelijke inspanning is in dit geval adekwaat gebleken met betrekking tot het handhaven van de energiebalans (hoofdstuk III).

De waarneming dat atleten die een KH-rijke voeding consumenten, samengesteld uit normale voedingsmiddelen, met daarbij een gemid-
delde eiwitinneming van 1.7 g.kg\(^{-1}\).dag\(^{-1}\), niet in staat waren om hun stikstofbalans te handhaven, roepen vragen op over de juistheid van een extrapolatie van de aanbeveling voor eiwit, verkregen uit onderzoeken bij de "normale" bevolking, naar atleten die lichamelijke arbeid verrichten tot aan de grenzen van het menselijke vermogen. Het gegeven dat KH suppletie enerzijds leidt tot het handhaven van de energie- en "KH"-balans, doch anderzijds niet tot het handhaven van de stikstofbalans, indien de dagelijkse eiwitinneming minder is dan 1.5 g.kg\(^{-1}\).dag\(^{-1}\), bewijst eveneens dat de dagelijkse eiwitbehoeftes van intensief trainende atleten hoger is dan de aanbevolen hoeveelheid voor normaal aktieve mensen (hoofdstuk III en IV).

De hier vermelde gegevens wijzen er echter niet op of er mogelijk een aanpassing aan een "relatieve lage" eiwit inneming optreedt indier herhaald uitputtende arbeid wordt verricht gedurende een langere periode. Een dergelijke adaptatie ten gevolge van training werd waargenomen tijdens het verrichten van submaximale, niet uitputtende, arbeid (Gontzea, 1975).

De toegenomen consumptie van mono- en di-sacchariden in de dagelijkse voeding is als risikofactor voor de ontwikkeling van hart- en vaatziekten en van diabetes gedurende de laatste 20 jaar onderwerp van diskussie geweest (Davidson, 1965). Het advies aan sporters om KH-oplossingen in te nemen tijdens de inspanning in plaats van de normale voeding of als toevoeging aan de normale maaltijden met het doel om de absolute KH inneming te vergroten, kan daarom strijdig lijken. Er is echter een aantal redenen aan te voeren waarom dit bepaalde omstandigheden niet zo is. Zo leidt de inneming van veel KH tijdens inspanning niet tot een sterke verhoging van de plasma-insulinespiegel. Naar aanleiding van deze waarneming en van de onderzoeken die aantonen dat de hoeveelheid KH die tijdens inspanning wordt ingenomen, wordt geoxideerd, kan de hypothese worden gesteld dat een grote KH-inneming tijdens het verrichten van zware lichamelijke inspanning geen nadelige gevolgen heeft te hebben.

De samenhang tussen KH-konsumptie enerzijds en de grootte van d KH-voorraad in het lichaam en het lichamelijk prestatievermogen anderzijds is in een reeks van onderzoeken aangetoond. In het voorliggende onderzoek wordt aangetoond dat het prestatievermogen significant verbeterde na de toevoeging van een KH-rijke vloeistof aan een reeds KH-rijke voeding bestaande uit normale voedingsmiddelen (hoofdstuk V). Tevens wordt aangetoond dat de additionele KH-inneming bij het verrichten van langdurige inspanning een eiwitparend effect heeft (hoofdstuk IV) en dat het herstel van d
glycogeenvoorraad gedurende 24 uur na de inspanning is versneld.

Fruktose is de laatste tijd, vanwege zijn "insuline onafhankelijk-
heid", aanbevolen als alternatieve KH bron voor sporters. Het
argument is dat fruktose de secretie van insuline niet stimuleert,
waardoor een maximale lipolytische activiteit mogelijk is, terwijl
tegelijkertijd koolhydraat wordt toegevoerd. In het voorliggende
werk wordt echter aangetoond dat suppletie van een KH-oplossing,
welke voor 50% uit vrij fruktose bestaat, resulteert in een sterk
toegenomen KH- en een afgenomen vetmetabolisme (hoofdstuk III).

De proefpersonen die deze voedingsmanipulatie ondergingen, blijken
tevens de grootste eiwitafbraak te hebben, gemeten aan de hand van
de totale stikstofuitscheiding. Deze resultaten, welke ondersteund
worden door het recente werk van Tappy et al (1986), spreken de
hypothese over een toegenomen vetverbranding na fruktose-, in
vergelijking tot na glucoseinneming, tegen.

In verschillende onderzoeken is een hoge plasma insuline spiegel
na de inneming van KH-oplossingen in nuchtere toestand tijdens rust
waargenomen, en daaraan gekoppeld een afname van het fysieke
prestatievermogen, indien de prestatie wordt begonnen op het
moment dat de insuline spiegel hoog is. De situatie en de voedings-
toestand waarin de atleten in deze onderzoeken getest werden
verschilt echter van die waarin de sporter zich in de praktijk bevindt
(hoofdstuk VI).

De in dit proefschrift gepubliceerde resultaten tonen aan dat de
inneming van KH-oplossingen na een ontbijt en tijdens een warming-
up geen nadelig effect heeft op de bloedglucosepiegel of op de
subjektieve vermoeidheid. Het blijkt daarentegen dat de bloedglu-
cosespiegel significant verhoogd blijft ten opzichte van de controle
waarden indien de inneming op de juiste wijze geschiedt. Met betrek-
king tot het voedingsadvies aan de sporter zou daarom benadrukt
moeten worden dat de inneming van KH-oplossingen alleen zou
moeten plaatsvinden aan het einde van de warming-up, indien deze
direct wordt gevolgd door de wedstrijd of ten hoogste 3-5 minuten
voor de start indien een warming-up niet mogelijk is.

In welke mate de inneming van voedsel en vocht tijdens de wedstrijd
leidt tot het ontstaan van buikklachten is niet geheel duidelijk
(hoofdstuk VII). Het is algemeen aanvaard dat het sporten met een
volle maag kan leiden tot buikklachten. Derhalve wordt het advies
gegeven om de laatste maaltijd tenminste 3 uur voor de inspanning te
nuttigen. Met betrekking tot de buikklachten welke tijdens de inspan-
ning op kunnen treden, kan dit echter te kort zijn omdat de maag-
lediging van een normale maaltijd, afhankelijk van de samen-
stelling, langer dan 5 uur kan duren. De konstatering dat homogenisering van een maaltijd leidt tot een duidelijke vermindering van de verblijftijd in de maag en dat vloeibare maaltijden met een laag gehalte aan voedingsvezel en vet een snelle maaglediging ten gevolge hebben, is een bijzonder belangrijk aspect met betrekking tot de voedselconsumptie tijdens langdurige arbeid. Sportbegeleiders hebben wel gesuggereerd dat het optreden van buikklachten en diarree bij hardlopers samenhangt met een slechte absorptie van het ingenomen voedsel. Er zijn echter observaties dat de maaglediging en voedingsstoffenabsorptie in de darm niet verandert bij arbeidsspaningen tot 70% VO\textsubscript{2} max. Bijna alle beschikbare onderzoekingen zijn uitgevoerd tijdens fietsbelasting en gegevens van loopexperimenten zijn niet voorhanden. Het gegeven dat de klachten vrijwel alleen optreden bij hardlopers en nauwelijks bij wielrenners, wijst erop dat de manier van voortbewegen een belangrijke ontstaansfactor kan zijn. De hypothese kan gesteld worden dat veranderingen in maag-darmmotilitiet en gevolge van een toegenomen reflexactiviteit van de darmwand, die wordt veroorzaakt door de schokkende bewegingen tijdens het hardlopen, het ontstaan van darmkramen veroorzaakt, terwijl ischemische necrose ten gevolge van ernstige dehydratie de oorzaak kan zijn van de bloedige diarree welke bij hardlopers is waargenomen.

Tot op heden zijn er echter geen gegevens van gekontroleerd onderzoek over dit probleem beschikbaar.

**konklusies**

Aan de hand van de hier verzamelde gegevens en literatuur kan gesteld worden dat op dagen van langdurige inspanning, voor wat de voedselconsumptie betreft, een drempelwaarde bestaat van ongeveer 20 MJ. Daarboven is voedselinname moeilijk dan wel onmogelijk. De verminderde eetlust ten gevolge van intensieve duurinspanning en de veelal door vast voedsel veroorzaakte overlast onder deze omstandigheden zijn daarbij waarschijnlijk de belangrijkste factoren.

Voor inspanningen van de beschreven intensiteit is de normaal aanbevolen inname van 60 en% KH in de vorm van gewone voedingsmiddelen niet voldoende om de "KH-" en energiebalans te handhaven. Indien KH oplossingen worden gesupplementeerd wordt de "KH-" en energiebalans gehandhaafd en neemt het prestatie vermogen toe.

Bij een dergelijke inspanning bedraagt de minimale eiwitbehoefte van de sporter vermoedelijk 1.5 tot 1.8 gram per kilogram
lichaamsgewicht per dag hetgeen hoger is dan de gebruikelijke aanbeveling van 0,8 g.kg⁻¹.dag⁻¹.

Bij langdurige uithoudingsvermogen treedt, ondanks de inneming van geconcentreerde KH oplossingen, een versterkte lipolyse op.

Door middel van supplementering met KH concentraten kan binnen 24 uur een supercompensatie van het glycogengehalte van spierweefsel worden bereikt.

Door de verkregen resultaten wordt de suggestie gewekt dat de inneming van fruktose bij lichamelijke inspanning leidt tot een stimulering van de KH stofwisseling en een remming van de vetstofwisseling.

In het algemeen kan worden gekonkludeerd dat toekomstig onderzoek op het gebied van de voeding van topsporters moet plaatsvinden onder gekontroleerde doch voor de sportpraktijk relevante omstandigheden.

Het verzamelen van informatie over de dagelijkse behoefte aan voedingsstoffen van topsporters is zeer gewenst. De bereidheid van deze zeer getalenteerde mannen en vrouwen om deel te nemen aan wetenschappelijk onderzoek zal echter een van de meest beperkende factoren zijn voor het verkrijgen van de gewenste gegevens.

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NAWOORD

Een alternatieve route
De weg die ik bewandeld heb om uiteindelijk dit werk te schrijven was geen gewone weg. Het was een toeristische route door het land van de sport, de lichamelijke opvoeding en uiteindelijk de wetenschap. Ik heb geboet dat ik onderweg zoveel gezien heb en dat ik zoveel vrienden ontmoette die wisten aan te geven welke route voor mij waarschijnlijk de meest boeiende zou zijn. Als actief wedstrijdschaatser geïnteresseerd geraakt in trainingsmethoden en het functioneren van het menselijk lichaam, volgde ik de opleiding voor bondstrainer, direct gevolgd door de cursus sportmassage. Op de T-splitsing waar ik toen arriveerde stond een bord richting fysiotherapie en een bord richting lichamelijke opvoeding; het was de laatste richting waar de heer Klaue mij bij het trainersexamen op wees: "voor jou een veel mooiere weg!"

Geprikkeld door nieuwsgierigheid belandde ik op de Academie voor Lichamelijke Opvoeding te Amsterdam waar mijn belangstelling voor de achtergronden van het menselijk bewegen definitief vorm kreeg. Ik ontdekte dat een snelle manier van leren en tevens studiefinanciering het schrijven van artikelen was en het was de "grote jachtakte" van Hein Ouwersloot die het mij mogelijk maakte om boeiende zaken te beschrijven wanneer de "rest in het water lag". De wind blies in mijn rug toen Niels Lommen een proefwerk als "Chateau neuf du Pape, grand cru" tussen een aantal tafelwijnen kwalificeerde en toen Piet Alkema mij stimuleerde om mijn scriptie over hardrijden op de schaats te presenteren, mijn eerste boekje. Na het behalen van het eindexamen in 1977 bleek al gauw dat het er in de wereld van de lichamelijke opvoeding helemaal niet zo rooskleurig voorstond. Een afnemend aantal leerlingen, grotere schoolklassen en een overschot aan leraren noopten mij tot het inslaan van een andere weg, leidend naar een functie bij de Koninklijke Nederlandse Atletiek Unie.

Trainingsleer en sportbegeleiding waren vanaf dat moment niet langer meer hobbylelementen. Na een periode van bijzondere belangstelling voor de training en belastbaarheid van jeugdigen en boeiende diskussies met Han Kemper en Jos Geisel ging mijn interesse in de richting van de energiehuishouding bij lichamelijke inspanning. De rol van de voeding kwam daarbij naar voren en het was in dat verband dat ik in St Etienne voor het eerst Wim Saris ontmoette, een groentje op het gebied van de sport dacht ik toen, niet wetend dat hij later hoogleraar en tevens een van mijn begeleiders zou worden. Gestuurd door de stimulerende invloed van Hans Keizer volgde de deelname aan een studiedag aan de Universiteit van Limburg met als onderwerp voeding en topsport. Mijn rol van discussieleider wekte de belangstelling van Wander Nederland waar ik korte tijd later een functie op de wetenschappelijke afdeling zou aanvaarden met als belangrijkste werkgebied voeding bij inspanning. Het was die functie die mij in de gelegenheid stelde om
kongressen te bezoeken waar mijn interesse in de rol van de voeding bij inspanning definitief gestalte kreeg. Het was de ontmoeting met Lars Hermansen, Dave Costill en Jack Wilmore die leidde tot de uitspraak dat de wetenschap wellicht een interessante toekomst voor mij verborg. De wens om een andere, meer wetenschappelijke weg te bewandelen werd definitief toen ik mij, gesteund door Rob Binkhorst, Foppe ten Hoor, Lars Hermansen en Wim Saris, aan de Vrije Universiteit van Brussel aanmelde voor een doktoraatus. De bemiddeling van Marcel Hebbelinck leidde tot een licentiaat equivalentie en de bemiddeling van Hartwig Kämpf en Hans Widmer resulterde in een research grant van Wander Bern. De keuze voor onderzoek viel op Maastricht. Ik verhuisde naar het land van het Bourgondische leven.

Dat ik nu in 1988 in Maastricht kan promoveren dank ik aan een nieuwe promotie regeling, een positief aspekt van het bewind Deetman.

Er is te weinig ruimte om alle andere vrienden die ik onderweg heb ontmoet en die mij hebben gesteund in het telkens weer kiezen van de weg die de grootste uitdaging leek te zijn, te vermelden. Het waren er vele en ik ben ze niet vergeten!

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Ed, hoe speel je het telkens weer klaar om “cool” te blijven terwijl alles om ons afbrant? Beseffend dat het in dit proefschrift beschreven materiaal nog niet de helft is van alles wat wij verzameld hebben, heb ik er vaak aan gedacht wat er allemal zou zijn mis gegaan zonder jouw laboratoriumexpertise en je grenze-

Dit proefschrift dankt zijn inhoud mede aan de stimulerende discussies die ik kon hebben met Lars Hermansen, Herman Adlercreutz, Jack Wilmore, Jacques Poortmans, Eric Newsholme, Per Henric Galbo, Pete Lemon, Peter Soeters en Rob Binkhorst. Zij waren allen betrokken bij de discussie over de pilotstudie en hadden tijd voor mij beschikbaar op het moment dat ik een boel moest leren.

"Scientific faults, if you survive them, make you love science more!"

Foppe en Wim, ik heb er heet wat gemaakt en ik ben steeds meer gaan houden van de wetenschap die mij zo boeit! Ik realiseer mij dat de ontspannen manier waarop jullie mij hebben begeleid en ook hebben geholpen op tijden dat ik het beslist niet gemakkelijk had, ertoe heeft geleid dat ik dit proefschrift in de geplande drie en een half jaar op tafel heb gekregen. Ik heb in deze periode van jullie altijd weer kritische vragen en stimulerende discussies ontzetend veel geleerd.

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CURRICULUM VITAE:


In de periode 1977-1980 was de auteur werkzaam als hoofd van de afdeling opleiding bij de Koninklijke Nederlandse Atletiek Unie te Utrecht en was lid van de projektgroep topsport van de Nederlandse Sport Federatie.

In 1978 werd deelgenomen aan de "International course on kinanthropometric techniques" aan de vrije universiteit van Brussel. Van 1980-1983 vervulde de auteur de functie van hoofd van de wetenschappelijke afdeling bij Wander Nederland bv. te Uden. Sinds november 1983 verrichtte de auteur wetenschappelijk onderzoek op het gebied van voeding en inspanning aan de Universiteit van Limburg te Maastricht onder leiding van Prof. Dr. Ir. W. H. M. Saris en Prof. Dr. F. ten Hoor.