

# Non-coding RNA species in heart failure

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## Chapter 7

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### Summary and general discussion

The aim of this thesis was to extend our knowledge on the role of different non-coding RNA species in the development of heart failure (HF). Over the last years it has been acknowledged that multiple cell types are critically involved in HF, including cardiomyocytes, cardiac fibroblasts, and resident or infiltrating immune cells. This thesis comprises research on the involvement of these three cell types in HF with a focus on microRNAs (miRNAs) and long non-coding RNAs (lncRNAs) as regulators of cell function. Only very recently, the cell type-specific effects of miRNAs in HF and their role in paracrine signaling have come to the center of attention<sup>1-5</sup>. Here we extend this knowledge by identifying miR-139 as regulator of cardiomyocyte function and the miR-221/222 family as repressor of fibroblast activation. In comparison, the role of lncRNAs in the different cardiac cell types is to date essentially unknown. We performed an in-depth analysis of the development of HF in mice lacking the lncRNA Malat-1 but surprisingly did not find a relevant function in any of the mentioned cell types. In contrast, we propose mascRNA, a processed product of Malat-1, to act as regulator of the immune system during VM. The overall conclusion is that ncRNAs play distinct roles in different cell types that could be exploited for targeted therapy strategies in cardiac disease.

## MIRNAS AS MODULATORS OF CARDIOMYOCYTE FUNCTION AND MYOCARDIAL FIBROSIS

In chapter 2 of this thesis we have shown that miR-139 is differentially expressed in human aortic valve stenosis and affects calcium handling in cardiomyocytes, probably by interfering with cAMP/PDE signaling. Stimulation of  $\beta$ -adrenergic receptors ( $\beta$ -AR) is a central physiological response to increased cardiac demands that evokes positive inotropic and lusitropic effects in cardiomyocytes<sup>6</sup>. The intracellular effect of  $\beta$ -AR activation is initiated by production of cAMP by adenylate cyclase, which leads to activation of cAMP effector proteins such as PKA and EPAC1, whereas phosphodiesterases (PDEs) terminate  $\beta$ -AR signaling by degrading cAMP. In HF, the adrenergic system is activated but the downstream effects are altered due to reduced number and function of the  $\beta$ -AR<sup>7</sup> and deregulation of the intracellular signaling cascade, including PDEs<sup>8-11</sup>. We validated the phosphodiesterases 3a, 4a and 4d as targets of miR-139 and found indications for increased phosphorylation of the L-type calcium channel in rat cardiomyocytes overexpressing miR-139. At the same time, miR-139 reduced spontaneous calcium release from the SR. These findings indicate that miR-139 is involved in calcium entry and intracellular cycling, although more detailed studies are needed to unravel the precise mechanism involved. *In vivo* we found that overexpression of miR-139 aggravated LV dilation after pressure overload. Defining the link between PDE repression and long-term effects after pressure overload *in vivo* was beyond the scope of this project because of the complexity of cAMP regulation and the multitude of possible miR-139 targets involved. Of note, it is conceivable that apart from affecting PKA signaling as assessed *in vitro*, miR-139 also influences transcriptional programs involved in cardiac hypertrophy, such as cAMP/EPAC1 signaling<sup>12</sup>. Therefore, further studies are warranted to delineate the effect of miR-139 on cAMP/PDE signaling in the diseased heart.

Besides cardiomyocyte dysfunction, myocardial fibrosis is a hallmark of HF and negatively affects cardiac contraction, electrical conduction, and oxygen supply<sup>13</sup>. In chapter 3 we have reported that the miRNA-221/222 family correlates negatively with the level of cardiac fibrosis in patients with aortic valve stenosis. In a mouse model of cardiac pressure overload we showed that inhibition of miR-221/222 aggravates the development of cardiac dysfunction and fibrosis. These data indicate that high myocardial levels of miR-221/222 may protect against excessive fibrosis in cardiac disease. To unravel the mechanism behind this, we modulated expression of miR-221/222 in rat cardiac fibroblasts and found a repressive effect on TGF $\beta$ -induced activation of fibroblasts into collagen-producing myofibroblasts. This effect is possibly mediated by the miR-221/222 target *Ets1*, a pro-fibrotic transcription factor that synergizes with TGF $\beta$ <sup>14,15</sup>. While we show a detrimental role of miR-221/222 in HF by acting on fibroblasts, other groups have investigated cardiomyocyte-specific effects of miR-221/222: Liu et al. found that miR-222 is necessary for physiological cardiac hypertrophy and that its overexpression in cardiomyocytes attenuates ischemic injury and reduces fibrosis<sup>16</sup>. In contrast, Su et al. reported that cardiomyocyte-restricted overexpression of miR-221 leads to spontaneous development of heart failure due to cardiomyocyte dysfunction and death<sup>17</sup>. Taken together, the outcome of systemic miRNA inhibition as performed in our study differs from that of cell type-restricted intervention. The prevailing phenotype of systemic inhibition probably depends on the cell type with highest expression of the targeted miRNA, which calls for the development of more targeted delivery strategies as discussed further below.

## LNCRNAS AS NOVEL PLAYERS IN HEART DISEASE

Several studies describe an important role for lncRNAs especially in cardiac development but also in heart disease as summarized in chapter 4. However, it remains challenging to identify lncRNAs with relevant functions in the heart and especially to define the precise pathway and binding partners mediating this function. Several lncRNAs have functions in the immune system<sup>18</sup> but their role in cardiac inflammation had not been investigated so far. In chapter 6 we have provided first evidence for a protective role of mascRNA, a lncRNA-derived small ncRNA, in the regulation of the immune response during VM. We found mascRNA to induce cellular defense mechanisms against viral entry and replication by upregulating IFITM and IFIT genes. Above that, systemic manipulation of mascRNA affected circulating immune cell populations. MascRNA is therefore a novel player in the pathophysiology of VM and an interesting study object in immunology and infection research in general.

Surprisingly, we found the host transcript of mascRNA, the lncRNA Malat-1, to be dispensable for the development of pressure overload-induced HF. Malat-1 is a highly abundant nuclear lncRNA with remarkable evolutionary conservation. It has been reported to affect vascularization<sup>19</sup>, to regulate the abundance of the muscle-specific miR-133<sup>20,21</sup>, and to activate the pro-hypertrophic ERK/MAPK pathway<sup>22</sup>. These reports notwithstanding, cardiac pressure overload in Malat-1 knockout mice had largely the same effect as in wild

type mice (chapter 5). We performed in-depth phenotyping including analysis of LV morphology, function, histology, and gene expression, all of which are known to be perturbed in HF. However, we only found an effect of Malat-1 ablation on splicing of *Ndr2*, which was apparently irrelevant for the clinical outcome. These unexpected observations suggest that the reported functions of Malat-1 as regulator of vascularization, scavenger of miR-133, and activator of ERK/MAPK signaling may be context-dependent and do not sum up to an important role of Malat-1 in cardiac hypertrophy and failure. Our results therefore stress that individually reported lncRNA functions need to be validated in complex disease models and highlight that sequence conservation and high expression level of a lncRNA do not necessarily indicate important (patho-)physiological functions.

### NOVEL TOOLS FOR NOVEL RNA

MiRNAs are known to function primarily via target gene repression (except for anecdotal reports about enhanced target gene translation<sup>23-25</sup>), and there are established tools to predict and validate miRNA targets and to modulate miRNA levels *in vitro* and *in vivo* by use of miRNA mimics, inhibitors, viral vectors or antagomirs<sup>26-29</sup>. LncRNAs, in strong contrast to miRNAs, are a very heterogeneous class of RNA molecules with variable length, post-transcriptional processing, three-dimensional structure, intracellular localization, and mode of action as outlined in chapter 4. A first bioinformatic method to predict the function of lncRNAs has very recently been developed but is limited to effects on gene regulation<sup>30</sup>. Therefore, research into lncRNA function so far relies on loss-of-function experiments by post-transcriptional knockdown or genomic deletion. Both approaches have their values but also some limitations, and novel approaches are needed to investigate this novel class of RNA.

LncRNAs may act in the cytoplasm or nucleus, and it is important to know the predominant location to design appropriate knockdown strategies. For example, siRNA-mediated knockdown is very efficient in the cytoplasm but its efficacy in the nucleus is controversial<sup>31-33</sup> and may relate to repression of transcription rather than post-transcriptional target degradation<sup>34,35</sup>. Therefore, we made use of gapmers to knock down nuclear Malat-1 (chapter 6). Gapmers are antisense oligonucleotides with an LNA-DNA-LNA backbone that potently induces cleavage of nuclear lncRNAs by RNaseH<sup>36,37</sup>. However, even with the correct approach, knockdown efficiency can be impaired by lncRNA secondary structure or binding partners that block the target site. For example, in chapter 6 we discussed difficulties to knock down the cytoplasmic mascRNA, probably due to its secondary structure. Nuclear lncRNAs can have functions in organizing nuclear structures (e.g. *Neat1* in paraspeckles<sup>38</sup>), affect mRNA processing (e.g. Malat-1 in splicing<sup>39,40</sup>), or regulate gene expression in *cis* or in *trans* (e.g. XIST and HOTAIR<sup>41</sup>). Importantly, gene regulatory effects may even be independent of the lncRNA transcript and emerge from the mere transcriptional activity at the lncRNA gene locus<sup>42-45</sup>. In that case, post-transcriptional knockdown of the lncRNA will either way fail to identify the function of the gene locus.

Next to antisense-mediated knockdown strategies, genomic deletion of lncRNAs is widely employed. However, interfering with genomic integrity at a lncRNA locus may remove or reorganize binding sites for regulatory factors, leading to off-target effects<sup>46</sup>. For example, it has been shown that blocking transcription of the lncRNA *Fendrr* has different consequences than replacing the gene while keeping the locus transcriptionally active<sup>47,48</sup>. Therefore, a careful study design including appropriate controls is mandatory in lncRNA research and elaborate considerations about the interpretation of *in vivo* studies have been summarized recently<sup>46</sup>. Of note, positional off-target effects may also occur after genomic manipulation of coding genes, which has been carried out for several years. The lessons learned from lncRNA research may thus also have important implications for established methods in research on coding RNA.

Another layer of complexity is added by lncRNA processing and Malat-1 is a good example for this: The primary Malat-1 transcript is processed into a long nuclear-retained fragment with reported functions in gene transcription and mRNA splicing<sup>39,49-51</sup>, but its 3' terminus also gives rise to mascRNA, a small cytoplasmic ncRNA for which no function has been published previously<sup>52</sup>. In chapter 6 we have described a role for mascRNA in VM, but disentangling the functions of Malat-1 and mascRNA remains challenging. Complete removal of the Malat-1 locus abolishes expression of mascRNA, whereas transcriptional blockade of Malat-1 may still allow for residual expression of downstream regions from an unknown promoter. Also post-transcriptional knockdown of a lncRNA may or may not lead to simultaneous depletion of its processed products, depending on the time frame between transcription and processing. Novel methods of genome engineering, such as the CRISPR/Cas-system, may be suitable for targeted removal of a lncRNA and/or its processed product from the genome.

In contrast to loss-of-function studies, overexpression of lncRNAs is rarely employed to date and is much more challenging than overexpression or mimicking miRNAs. The use of adeno-associated virus (AAV) vectors *in vivo* is well-established and different serotypes allow for some organ specificity in transgene delivery. In chapter 2 and 6 we employed AAV9 vectors for cardiomyocyte-specific overexpression, but these vectors are limited to transgenes smaller than 4.5 kb<sup>29</sup> and some lncRNAs including Malat-1 are simply too large to fit into viral vectors. Above that, transgene delivery is futile for lncRNAs with a *cis*-regulatory function that depends on correct chromosomal location. Interestingly, methods to enhance expression of endogenous genes by modified zinc finger proteins<sup>53,54</sup> and TALEs<sup>55,56</sup> have been developed in the last years. Especially the recently described CRIPR/Cas-mediated gene activation system may provide an easily applicable tool to overexpress lncRNAs from the endogenous locus<sup>57</sup>.

## THE BROAD FIELD OF NCRNA RESEARCH: WHICH WAY TO GO?

Our growing knowledge about miRNA function and the impressive potency of miRNA modulation in animal models has quickly elicited interest of pharmaceutical companies, and first miRNA-based therapeutics are already being developed and tested<sup>58-60</sup>. The transition from discovery to clinical testing has happened with remarkable speed: The first miRNA, lin-4, was discovered in 1993<sup>61</sup> and 5 years later the concept of RNAi was introduced<sup>62</sup>, although the term “microRNA” was only coined in 2001<sup>63-65</sup>. Already in 2008, the applicability of miRNA inhibitors in primates was tested<sup>66</sup> and soon followed by a clinical phase 2a trial in humans, which showed promising results for the treatment of hepatitis<sup>67</sup>. To date, we are clearly able to effectively inhibit miRNAs *in vivo*, and by now also the first miRNA mimic has entered a phase 1 clinical trial<sup>68</sup>. However, this haste in exploiting the therapeutic potential of miRNAs also calls for a sober view on the potential limitations and dangers. Possible adverse effects include activation of the immune system by exogenous RNA and off-target effects by unintentional modulation of unrelated miRNAs<sup>60</sup>. Above that, our results and the growing knowledge about cell type- and context-specific miRNA functions highlight the possibility of side effects by manipulating the intended miRNA in unintended cell types or organs. In chapter 3 we described anti-fibrotic effects of the miR-221/222 family in cardiac pressure overload, putting miRNA mimics forward as therapeutic option to dampen fibroblast activation. However, another group has shown that overexpression of miR-221 in cardiomyocytes causes cell death and leads to HF<sup>17</sup>. Above that, the miR-221/222 family can suppress or promote malignant diseases, depending on the type of tumor<sup>69</sup>. In view of the Janus-faced function of miRNAs, possible off-target effects need to be carefully assessed and advances in targeted delivery methods are much-needed before the full potential of miRNA-based medicine can safely be harnessed<sup>60,70</sup>.

While the first miRNAs enter the clinic, the involvement of lncRNAs in (patho-)physiological processes in the heart has only been investigated in the last few years. In chapter 4 we have summarized the progress that has been made in assigning functions to individual lncRNAs and their involvement in cardiac development, function, and disease. We showed that research on lncRNAs in cardiac disease is still in its infancy but offers a new opportunity to develop biomarkers and treatment options in heart failure. In the years while I was preparing this thesis, the number of publications on lncRNAs has steeply increased from about 100 publications indexed in PubMed in 2011 to more than 700 in 2014. lncRNAs are clearly a “hot topic”, and interest in this area was boosted in 2012 by multiple publications from the ENCODE consortium (Encyclopedia of DNA Elements), stating that the vast majority of the genome is not silent “junk” as previously considered<sup>71</sup> but transcribed and functional<sup>72,73</sup>. However, it did not take long until a debate about the interpretation of ENCODE arose<sup>74,75</sup>, although part of the controversy may relate to definitions and semantics. The current interest in lncRNAs exemplifies the problem of “hypes” in scientific research, leading to high expectations and possibly a one-sided and misleading interpretation of novel observations. The lncRNA Malat-1 is remarkably well conserved, highly expressed in most

tissues, and has a function in synaptogenesis<sup>40</sup>. Therefore, expectations were high that genomic deletion of Malat-1 would have severe consequences. Three independent groups made an effort to generate Malat-1 knockout mice, but surprisingly all three strains develop and breed normally<sup>51,76,77</sup>. Somewhat later, several reports indicated a role for Malat-1 in cardiac hypertrophy but we found Malat-1 to be dispensable for the development of pressure overload-induced cardiac hypertrophy and failure (chapter 5).

Obviously, several highly important lncRNAs have been identified to date, but it remains questionable if they constitute the “tip of the iceberg” of functional lncRNAs, or if most lncRNAs are in fact redundant or even non-functional. A publication from Sauvageau et al. exemplifies this conundrum and illustrates the problem of “misleading interpretation” as mentioned above. In their methodologically remarkable and insightful study “Multiple knockout mouse models reveal lincRNAs are required for life and brain development”, the authors generated 18 lncRNA knockout mice and found clear abnormalities in 5 of them, indeed suggesting that some lncRNAs are crucial for normal development<sup>48</sup>. However, in a detailed comment, Claudiu Bandea criticizes the misleading title and emphasizes that actually “most lncRNAs (i.e. 13 out of 18) do not appear to play critical roles *in vivo*”<sup>78</sup>. In conclusion, research of the last years has identified several highly interesting lncRNAs but the true potential of lncRNAs to revolutionize biomedical research is still difficult to assess.

## CONCLUDING REMARKS

This thesis provides a comprehensive picture of the cell types involved in the development of HF and highlights how ncRNAs can affect cell type-specific pathological mechanisms. It is evident from our data and from the large amount of work done by other groups that ncRNAs are crucial regulators of cell function and consequently important players in heart failure. The levels of knowledge, however, differ greatly when it comes to compare miRNAs and lncRNAs. Several miRNAs are known to play important roles in the development of HF, and we have contributed some more puzzle pieces to this knowledge. The utilization of miRNA-based therapeutics for HF appears to be a matter of time despite some hurdles that still need to be taken. In contrast, research on lncRNAs in heart failure is still in its infancy and is hampered by methodological challenges and the heterogeneity of lncRNA function. However, progress in science depends on sailing unknown seas and with the discovery of lncRNAs there is now a New World to explore.

## REFERENCES

1. Townley-Tilson WH, Callis TE and Wang D. MicroRNAs 1, 133, and 206: critical factors of skeletal and cardiac muscle development, function, and disease. *Int J Biochem Cell Biol.* 2010;42:1252-5.
2. Viereck J, Bang C, Foinquinos A and Thum T. Regulatory RNAs and paracrine networks in the heart. *Cardiovasc Res.* 2014;102:290-301.
3. Thum T. Noncoding RNAs and myocardial fibrosis. *Nat Rev Cardiol.* 2014;11:655-63.
4. Melman YF, Shah R and Das S. MicroRNAs in heart failure: is the picture becoming less miRky? *Circ Heart Fail.* 2014;7:203-14.
5. Tijssen AJ, Pinto YM and Creemers EE. Non-cardiomyocyte microRNAs in heart failure. *Cardiovasc Res.* 2012;93:573-82.
6. Bers DM. Cardiac excitation-contraction coupling. *Nature.* 2002;415:198-205.
7. Lohse MJ, Engelhardt S and Eschenhagen T. What is the role of beta-adrenergic signaling in heart failure? *Circ Res.* 2003;93:896-906.
8. Ding B, Abe J, Wei H, Huang Q, Walsh RA, Molina CA, Zhao A, Sadoshima J, Blaxall BC, Berk BC and Yan C. Functional role of phosphodiesterase 3 in cardiomyocyte apoptosis: implication in heart failure. *Circulation.* 2005;111:2469-76.
9. Pokreisz P, Vandewijngaert S, Bito V, Van den Bergh A, Lenaerts I, Busch C, Marsboom G, Gheysens O, Vermeersch P, Biesmans L, Liu X, Gillijns H, Pellens M, Van Lommel A, Buys E, Schoonjans L, Vanhaecke J, Verbeken E, Sipido K, Herijgers P, Bloch KD and Janssens SP. Ventricular phosphodiesterase-5 expression is increased in patients with advanced heart failure and contributes to adverse ventricular remodeling after myocardial infarction in mice. *Circulation.* 2009;119:408-16.
10. Abi-Gerges A, Richter W, Lefebvre F, Mateo P, Varin A, Heymes C, Samuel JL, Lugnier C, Conti M, Fischmeister R and Vandecasteele G. Decreased expression and activity of cAMP phosphodiesterases in cardiac hypertrophy and its impact on beta-adrenergic cAMP signals. *Circ Res.* 2009;105:784-92.
11. Mokni W, Keravis T, Etienne-Selloum N, Walter A, Kane MO, Schini-Kerth VB and Lugnier C. Concerted regulation of cGMP and cAMP phosphodiesterases in early cardiac hypertrophy induced by angiotensin II. *PLoS One.* 2010;5:e14227.
12. Metrich M, Lucas A, Gastineau M, Samuel JL, Heymes C, Morel E and Lezoualc'h F. Epac mediates beta-adrenergic receptor-induced cardiomyocyte hypertrophy. *Circ Res.* 2008;102:959-65.
13. Lopez B, Gonzalez A, Ravassa S, Beaumont J, Moreno MU, San Jose G, Querejeta R and Diez J. Circulating Biomarkers of Myocardial Fibrosis: The Need for a Reappraisal. *J Am Coll Cardiol.* 2015;65:2449-56.
14. Geisinger MT, Astaiza R, Butler T, Popoff SN, Planey SL and Arnott JA. Ets-1 is essential for connective tissue growth factor (CTGF/CCN2) induction by TGF-beta1 in osteoblasts. *PLoS One.* 2012;7:e35258.
15. Feng W, Chumley P, Hua P, Rezonzew G, Jaimes D, Duckworth MW, Xing D and Jaimes EA. Role of the transcription factor erythroblastosis virus E26 oncogen homolog-1 (ETS-1) as mediator of the renal proinflammatory and profibrotic effects of angiotensin II. *Hypertension.* 2012;60:1226-33.
16. Liu X, Xiao J, Zhu H, Wei X, Platt C, Damilano F, Xiao C, Bezzerides V, Bostrom P, Che L, Zhang C, Spiegelman BM and Rosenzweig A. miR-222 is necessary for exercise-induced cardiac growth and protects against pathological cardiac remodeling. *Cell Metab.* 2015;21:584-95.
17. Su M, Wang J, Wang C, Wang X, Dong W, Qiu W, Wang Y, Zhao X, Zou Y, Song L, Zhang L and Hui R. MicroRNA-221 inhibits autophagy and promotes heart failure by modulating the p27/CDK2/mTOR axis. *Cell Death Differ.* 2015;22:986-99.
18. Heward JA and Lindsay MA. Long non-coding RNAs in the regulation of the immune response. *Trends Immunol.* 2014;35:408-19.

19. Michalik KM, You X, Manavski Y, Doddaballapur A, Zornig M, Braun T, John D, Ponomareva Y, Chen W, Uchida S, Boon RA and Dimmeler S. Long noncoding RNA MALAT1 regulates endothelial cell function and vessel growth. *Circ Res*. 2014;114:1389-97.
20. Han X, Yang F, Cao H and Liang Z. Malat1 regulates serum response factor through miR-133 as a competing endogenous RNA in myogenesis. *FASEB J*. 2015;29:3054-64.
21. Care A, Catalucci D, Felicetti F, Bonci D, Addario A, Gallo P, Bang ML, Segnalini P, Gu Y, Dalton ND, Elia L, Latronico MV, Hoydal M, Autore C, Russo MA, Dorn GW, 2nd, Ellingsen O, Ruiz-Lozano P, Peterson KL, Croce CM, Peschle C and Condorelli G. MicroRNA-133 controls cardiac hypertrophy. *Nat Med*. 2007;13:613-8.
22. Wu XS, Wang XA, Wu WG, Hu YP, Li ML, Ding Q, Weng H, Shu YJ, Liu TY, Jiang L, Cao Y, Bao RF, Mu JS, Tan ZJ, Tao F and Liu YB. MALAT1 promotes the proliferation and metastasis of gallbladder cancer cells by activating the ERK/MAPK pathway. *Cancer Biol Ther*. 2014;15:806-14.
23. Vasudevan S, Tong Y and Steitz JA. Switching from repression to activation: microRNAs can up-regulate translation. *Science*. 2007;318:1931-4.
24. Orom UA, Nielsen FC and Lund AH. MicroRNA-10a binds the 5'UTR of ribosomal protein mRNAs and enhances their translation. *Mol Cell*. 2008;30:460-71.
25. Mortensen RD, Serra M, Steitz JA and Vasudevan S. Posttranscriptional activation of gene expression in *Xenopus laevis* oocytes by microRNA-protein complexes (microRNPs). *Proc Natl Acad Sci U S A*. 2011;108:8281-6.
26. Min H and Yoon S. Got target? Computational methods for microRNA target prediction and their extension. *Exp Mol Med*. 2010;42:233-44.
27. Svoboda P. A toolbox for miRNA analysis. *FEBS Lett*. 2015;589:1694-701.
28. Philippen LE, Dirix E, Wit JB, Burggraaf K, de Windt LJ and da Costa Martins PA. Antisense MicroRNA Therapeutics in Cardiovascular Disease: Quo Vadis? *Mol Ther*. 2015.
29. Zacchigna S, Zentilin L and Giacca M. Adeno-associated virus vectors as therapeutic and investigational tools in the cardiovascular system. *Circ Res*. 2014;114:1827-46.
30. Xiao Y, Lv Y, Zhao H, Gong Y, Hu J, Li F, Xu J, Bai J, Yu F and Li X. Predicting the functions of long noncoding RNAs using RNA-seq based on Bayesian network. *Biomed Res Int*. 2015;2015:839590.
31. Robb GB, Brown KM, Khurana J and Rana TM. Specific and potent RNAi in the nucleus of human cells. *Nat Struct Mol Biol*. 2005;12:133-7.
32. Zeng Y and Cullen BR. RNA interference in human cells is restricted to the cytoplasm. *RNA*. 2002;8:855-60.
33. Lennox K. *Knockdown of lncRNAs: exploring RNAi and antisense oligo methods*. In: Techvault/Video library/Gene silencing. Integrated DNA Technologies; 2015 Aug 25 [cited 2015 Nov 01]. Available from: <http://eu.idtdna.com/pages/support/technical-vault/video-library/gene-silencing/knockdown-of-lncrnas-exploring-rnai-and-antisense-oligo-methods>
34. Castel SE and Martienssen RA. RNA interference in the nucleus: roles for small RNAs in transcription, epigenetics and beyond. *Nat Rev Genet*. 2013;14:100-12.
35. Morris KV, Chan SW, Jacobsen SE and Looney DJ. Small interfering RNA-induced transcriptional gene silencing in human cells. *Science*. 2004;305:1289-92.
36. Kurreck J, Wyszko E, Gillen C and Erdmann VA. Design of antisense oligonucleotides stabilized by locked nucleic acids. *Nucleic Acids Res*. 2002;30:1911-8.
37. Kauppinen S, Vester B and Wengel J. Locked nucleic acid (LNA): High affinity targeting of RNA for diagnostics and therapeutics. *Drug Discov Today Technol*. 2005;2:287-90.
38. Chen LL and Carmichael GG. Altered nuclear retention of mRNAs containing inverted repeats in human embryonic stem cells: functional role of a nuclear noncoding RNA. *Mol Cell*. 2009;35:467-78.
39. Tripathi V, Ellis JD, Shen Z, Song DY, Pan Q, Watt AT, Freier SM, Bennett CF, Sharma A, Bubulya PA, Blencowe BJ, Prasanth SG and Prasanth KV. The nuclear-retained noncoding RNA MALAT1 regulates alternative splicing by modulating SR splicing factor phosphorylation. *Mol Cell*. 2010;39:925-38.

40. Bernard D, Prasanth KV, Tripathi V, Colasse S, Nakamura T, Xuan Z, Zhang MQ, Sedel F, Jourden L, Couplier F, Triller A, Spector DL and Bessis A. A long nuclear-retained non-coding RNA regulates synaptogenesis by modulating gene expression. *EMBO J.* 2010;29:3082-93.
41. Fatica A and Bozzoni I. Long non-coding RNAs: new players in cell differentiation and development. *Nat Rev Genet.* 2014;15:7-21.
42. Petruk S, Sedkov Y, Riley KM, Hodgson J, Schweisguth F, Hirose S, Jaynes JB, Brock HW and Mazo A. Transcription of bxd noncoding RNAs promoted by trithorax represses Ubx in cis by transcriptional interference. *Cell.* 2006;127:1209-21.
43. Martianov I, Ramadass A, Serra Barros A, Chow N and Akoulitchev A. Repression of the human dihydrofolate reductase gene by a non-coding interfering transcript. *Nature.* 2007;445:666-70.
44. Latos PA, Pauler FM, Koerner MV, Senergin HB, Hudson QJ, Stocsits RR, Allhoff W, Stricker SH, Klement RM, Warczok KE, Aumayr K, Pasierbek P and Barlow DP. Airn transcriptional overlap, but not its lncRNA products, induces imprinted Igf2r silencing. *Science.* 2012;338:1469-72.
45. Kornienko AE, Guenzl PM, Barlow DP and Pauler FM. Gene regulation by the act of long non-coding RNA transcription. *BMC Biol.* 2013;11:59.
46. Bassett AR, Akhtar A, Barlow DP, Bird AP, Brockdorff N, Duboule D, Ephrussi A, Ferguson-Smith AC, Gingeras TR, Haerty W, Higgs DR, Miska EA and Ponting CP. Considerations when investigating lncRNA function in vivo. *Elife.* 2014;3:e03058.
47. Grote P, Wittler L, Hendrix D, Koch F, Wahrisch S, Beisaw A, Macura K, Blass G, Kellis M, Werber M and Herrmann BG. The tissue-specific lncRNA Fendrr is an essential regulator of heart and body wall development in the mouse. *Dev Cell.* 2013;24:206-14.
48. Sauvageau M, Goff LA, Lodato S, Bonev B, Groff AF, Gerhardinger C, Sanchez-Gomez DB, Hacısuleyman E, Li E, Spence M, Liapis SC, Mallard W, Morse M, Swerdel MR, D'Ecclesiss MF, Moore JC, Lai V, Gong G, Yancopoulos GD, Friendewey D, Kellis M, Hart RP, Valenzuela DM, Arlotta P and Rinn JL. Multiple knockout mouse models reveal lincRNAs are required for life and brain development. *Elife.* 2013;2:e01749.
49. West JA, Davis CP, Sunwoo H, Simon MD, Sadreyev RI, Wang PI, Tolstorukov MY and Kingston RE. The long noncoding RNAs NEAT1 and MALAT1 bind active chromatin sites. *Mol Cell.* 2014;55:791-802.
50. Hutchinson JN, Ensminger AW, Clemson CM, Lynch CR, Lawrence JB and Chess A. A screen for nuclear transcripts identifies two linked noncoding RNAs associated with SC35 splicing domains. *BMC Genomics.* 2007;8:39.
51. Zhang B, Arun G, Mao YS, Lazar Z, Hung G, Bhattacharjee G, Xiao X, Booth CJ, Wu J, Zhang C and Spector DL. The lncRNA Malat1 is dispensable for mouse development but its transcription plays a cis-regulatory role in the adult. *Cell Rep.* 2012;2:111-23.
52. Wilusz JE, Freier SM and Spector DL. 3' end processing of a long nuclear-retained noncoding RNA yields a tRNA-like cytoplasmic RNA. *Cell.* 2008;135:919-32.
53. Beerli RR, Dreier B and Barbas CF, 3rd. Positive and negative regulation of endogenous genes by designed transcription factors. *Proc Natl Acad Sci U S A.* 2000;97:1495-500.
54. Beerli RR and Barbas CF, 3rd. Engineering polydactyl zinc-finger transcription factors. *Nat Biotechnol.* 2002;20:135-41.
55. Zhang F, Cong L, Lodato S, Kosuri S, Church GM and Arlotta P. Efficient construction of sequence-specific TAL effectors for modulating mammalian transcription. *Nat Biotechnol.* 2011;29:149-53.
56. Miller JC, Tan S, Qiao G, Barlow KA, Wang J, Xia DF, Meng X, Paschon DE, Leung E, Hinkley SJ, Dulay GP, Hua KL, Ankoudinova I, Cost GJ, Urnov FD, Zhang HS, Holmes MC, Zhang L, Gregory PD and Rebar EJ. A TALE nuclease architecture for efficient genome editing. *Nat Biotechnol.* 2011;29:143-8.
57. Perez-Pinera P, Kocak DD, Vockley CM, Adler AF, Kabadi AM, Polstein LR, Thakore PI, Glass KA, Ousterout DG, Leong KW, Guilak F, Crawford GE, Reddy TE and Gersbach CA. RNA-guided gene activation by CRISPR-Cas9-based transcription factors. *Nat Methods.* 2013;10:973-6.
58. van Rooij E, Marshall WS and Olson EN. Toward microRNA-based therapeutics for heart disease: the sense in antisense. *Circ Res.* 2008;103:919-28.

59. Broderick JA and Zamore PD. MicroRNA therapeutics. *Gene Ther.* 2011;18:1104-10.
60. van Rooij E, Purcell AL and Levin AA. Developing microRNA therapeutics. *Circ Res.* 2012;110:496-507.
61. Lee RC, Feinbaum RL and Ambros V. The *C. elegans* heterochronic gene *lin-4* encodes small RNAs with antisense complementarity to *lin-14*. *Cell.* 1993;75:843-54.
62. Fire A, Xu S, Montgomery MK, Kostas SA, Driver SE and Mello CC. Potent and specific genetic interference by double-stranded RNA in *Caenorhabditis elegans*. *Nature.* 1998;391:806-11.
63. Lee RC and Ambros V. An extensive class of small RNAs in *Caenorhabditis elegans*. *Science.* 2001;294:862-4.
64. Lau NC, Lim LP, Weinstein EG and Bartel DP. An abundant class of tiny RNAs with probable regulatory roles in *Caenorhabditis elegans*. *Science.* 2001;294:858-62.
65. Lagos-Quintana M, Rauhut R, Lendeckel W and Tuschl T. Identification of novel genes coding for small expressed RNAs. *Science.* 2001;294:853-8.
66. Elmen J, Lindow M, Schutz S, Lawrence M, Petri A, Obad S, Lindholm M, Hedtjarn M, Hansen HF, Berger U, Gullans S, Kearney P, Sarnow P, Straarup EM and Kauppinen S. LNA-mediated microRNA silencing in non-human primates. *Nature.* 2008;452:896-9.
67. Janssen HL, Reesink HW, Lawitz EJ, Zeuzem S, Rodriguez-Torres M, Patel K, van der Meer AJ, Patick AK, Chen A, Zhou Y, Persson R, King BD, Kauppinen S, Levin AA and Hodges MR. Treatment of HCV infection by targeting microRNA. *N Engl J Med.* 2013;368:1685-94.
68. Bouchie A. First microRNA mimic enters clinic. *Nat Biotechnol.* 2013;31:577.
69. Garofalo M, Quintavalle C, Romano G, Croce CM and Condorelli G. miR221/222 in cancer: their role in tumor progression and response to therapy. *Curr Mol Med.* 2012;12:27-33.
70. van Rooij E and Kauppinen S. Development of microRNA therapeutics is coming of age. *EMBO Mol Med.* 2014;6:851-64.
71. Little WC, Ohno M, Kitzman DW, Thomas JD and Cheng CP. Determination of left ventricular chamber stiffness from the time for deceleration of early left ventricular filling. *Circulation.* 1995;92:1933-9.
72. Djebali S, Davis CA, Merkel A, Dobin A, Lassmann T, Mortazavi A, Tanzer A, Lagarde J, Lin W, Schlesinger F, Xue C, Marinov GK, Khatun J, Williams BA, Zaleski C, Rozowsky J, Roder M, Kokocinski F, Abdelhamid RF, Alioto T, Antoshechkin I, Baer MT, Bar NS, Batut P, Bell K, et al. Landscape of transcription in human cells. *Nature.* 2012;489:101-8.
73. Consortium EP. An integrated encyclopedia of DNA elements in the human genome. *Nature.* 2012;489:57-74.
74. Doolittle WF. Is junk DNA bunk? A critique of ENCODE. *Proc Natl Acad Sci U S A.* 2013;110:5294-300.
75. Graur D, Zheng Y, Price N, Azevedo RB, Zufall RA and Elhaik E. On the immortality of television sets: "function" in the human genome according to the evolution-free gospel of ENCODE. *Genome Biol Evol.* 2013;5:578-90.
76. Eissmann M, Gutschner T, Hammerle M, Gunther S, Caudron-Herger M, Gross M, Schirmacher P, Rippe K, Braun T, Zornig M and Diederichs S. Loss of the abundant nuclear non-coding RNA MALAT1 is compatible with life and development. *RNA Biol.* 2012;9:1076-87.
77. Nakagawa S, Ip JY, Shioi G, Tripathi V, Zong X, Hirose T and Prasanth KV. Malat1 is not an essential component of nuclear speckles in mice. *RNA.* 2012;18:1487-99.
78. Bandea C. *Comment on PMID 24381249: Multiple knockout mouse models reveal lincRNAs are required for life and brain development.* In: PubMed Commons [Internet]. Bethesda (MD): National Library of Medicine; 2014 Aug 08 [cited 2015 Oct 28]. Available from: [http://www.ncbi.nlm.nih.gov/pubmed/24381249#cm24381249\\_5641](http://www.ncbi.nlm.nih.gov/pubmed/24381249#cm24381249_5641)