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Modulation of Cell Death in the Tumor Microenvironment

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The microenvironment of solid human tumors is characterized by heterogeneity in oxygenation. Hypoxia arises early in the process of tumor development because rapidly proliferating tumor cells outgrow the capacity of the host vasculature. Formation of solid tumors thus requires coordination of angiogenesis with continued tumor cell proliferation. However, despite such neovascularization, hypoxia is persistent and frequently found in tumors at the time of diagnosis. Tumors with low oxygenation have a poor prognosis, and strong evidence suggests this is because of the effects of hypoxia on malignant progression, angiogenesis, metastasis, and therapy resistance. The presence of viable hypoxic cells is likely a reflection of the development of hypoxia tolerance resulting from modulation of cell death in the microenvironment. This acquired feature has been explained on the basis of clonal selection—the hypoxic microenvironment selects cells capable of surviving in the absence of normal oxygen availability. However, the persistence and frequency of

hypoxia in solid tumors raises a second potential explanation. We suggest that stable microregions of hypoxia may play a positive role in tumor growth. Although hypoxia inhibits cell proliferation and in tumor cells will eventually induce cell death, hypoxia also provides angiogenic and metastatic signals. The development of hypoxia tolerance will thus allow prolonged survival in the absence of oxygen and generation of a persistent angiogenic signal. We will discuss the concept of hypoxia tolerance and review mechanisms used by cancer cells to acquire this phenotype. The concept of hypoxia tolerance has important implications for current and future therapeutic approaches. Most therapeutic efforts to combat hypoxia have focused on targeting the presence of hypoxia itself. Our hypothesis predicts that targeting the biological responses to hypoxia and the pathways leading to hypoxia tolerance may also be attractive therapeutic strategies.

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The Concept of Hypoxia Tolerance

Evidence implicating hypoxia in the pathogenesis of solid human tumors continues to accumulate. Tumor hypoxia was hypothesized approximately 50 years ago to be important in the radiotherapeutic management of cancer because hypoxic cells are intrinsically more resistant to radiation than aerobic cells.¹ This initial interest in the radiobiological consequences of tumor hypoxia formed the basis of decades of research that, in recent years, has led to a close examination of the biological phenotypes of hypoxic cells. A number of seminal discoveries has led to the realization that hypoxia may be even more important than originally believed, contributing not only to therapy resistance but also to tumor malignancy. A wealth of data has shown that hypoxia can contribute to the malignant phenotype of tumors.

Hypoxia has been implicated in promoting metastasis, angiogenesis, and the selection of cells with a more malignant phenotype.²⁻⁸ The importance of hypoxia has been shown clinically when it predicts for worse outcome in the treatment of cancer of the head and neck, uterine cervix, and soft-tissue sarcomas.⁹⁻¹¹

In nonpathological tissue, a structurally and functionally normal vasculature provides cells with an adequate oxygen and nutrient supply. However, the situation for aggressively growing tumors is much different. Although deregulated cell growth may be sustained by the host vasculature for a short period of time, rapid cell proliferation will eventually lead to excessive demand for oxygen. The establishment of hypoxia is thus believed to occur very early in the development of a tumor, producing a microenvironment that is hypoxic, acidic, and low in nutrients. The response of cells to this environment is critical for the continued growth of the tumor.

If the tumor mass is to maintain its growth, a continuous supply of oxygen and nutrients is essential. Hypoxia can stimulate the formation of new blood vessels through various mechanisms such as increased secretion of vascular endothelial cell growth factor (VEGF).¹² Although angio-

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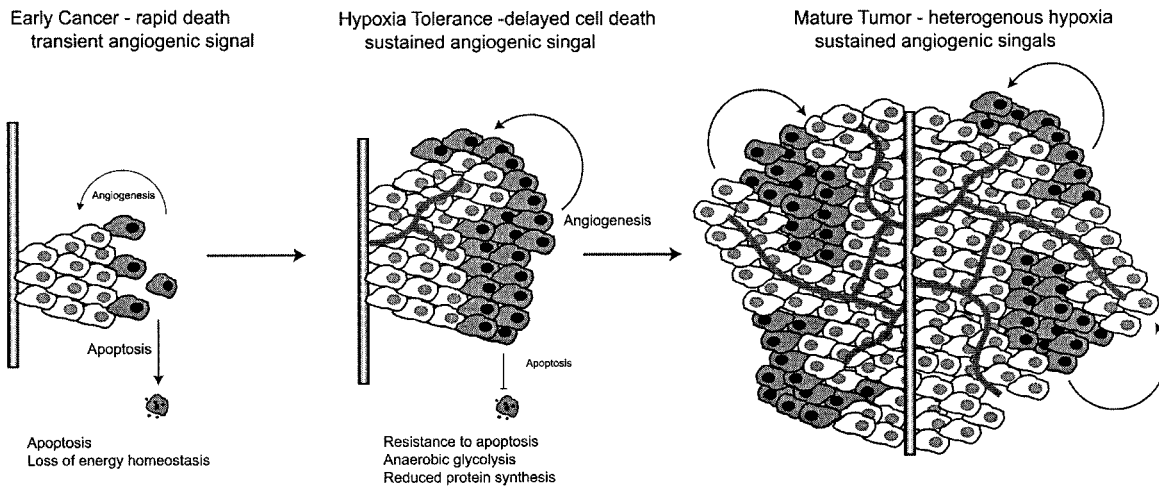


Figure 1. Hypoxia tolerance in the development of cancer. Hypoxia is hypothesized to arise early in the process of tumor development. Deregulation of cell growth results in an excess demand for oxygen and leads to cellular hypoxia. In normal and minimally transformed cells, hypoxia leads to cell death through activation of apoptosis or through loss of ATP. To continue to proliferate, tumors need to induce angiogenesis. During tumor evolution, cells become resistant to hypoxia-induced cell death. We hypothesize that this hypoxia tolerance arises such that a persistent hypoxia-induced angiogenic signal can be produced. The continued presence of viable hypoxic cells allows the coordination between angiogenesis and tumor expansion. This hypothesis implies that heterogeneity in oxygenation is beneficial to overall tumor growth.

genesis clearly occurs in solid tumors, clinical data have shown that hypoxia remains a common feature of tumors.^{9,10,13-15} Even in the case of tumors that have developed throughout a period of 30 years or more, hypoxia is persistent. This implies that the vasculature that develops in tumors is inadequate to provide normal levels of oxygenation. Consistent with this idea is the observation that tumor vasculature is often abnormal, characterized by sluggish or intermittent blood flow, leakiness, and structural abnormalities that further contribute to tumor hypoxia.^{16,17} We can thus conclude that hypoxia occurs early and remains a common feature of tumors throughout their development.

Cells can survive for only limited periods of time at low oxygen. In normal cells, hypoxia leads to the inhibition of cell growth and eventually to cell death. These same effects are also observed frequently in tumor cells but are generally less severe and/or develop with slower kinetics. This hypoxia tolerance is often explained as a result of a selective pressure in tumors that are forced to develop in an environment characterized by low oxygen availability. However, one could also hypothesize that the persistence of hypoxia in human tumors is a reflection of the fact that hypoxia

can act as a net positive factor in tumor growth. Because hypoxia can stimulate angiogenesis, the presence of heterogeneous areas of hypoxia may be beneficial to the overall growth of the tumor. This hypothesis predicts that tumors containing hypoxia tolerant cells will maintain a growth advantage—a prediction supported by many examples from both the laboratory and the clinic.^{2-8,18-20} Not only will such cells be able to survive limited exposures to hypoxia and perhaps proliferate again when oxygen becomes available, these cells can deliver a more prolonged angiogenic signal ensuring coordinated angiogenesis and cell growth. Thus, tumor cell growth, hypoxia, and angiogenesis become intrinsically linked. As tumor cells learn to tolerate hypoxia, both tumor cell growth and angiogenesis will be positively affected.

This leads us to an intriguing question. Is tolerance to hypoxia in cancer a common, or even necessary event in tumorigenesis? And if so, what are the implications for current therapies? If we can better understand the molecular mechanisms that control the adaptive responses to hypoxia in tumors, improved therapeutic approaches to the treatment of malignancies may be developed (Fig 1).²⁷

Mechanisms of Hypoxia Tolerance

The formation and continued proliferation of solid tumors requires persistent angiogenesis. Reminiscent of many biological processes, angiogenesis is the result of subtle and often complex interactions balancing pro- and antiangiogenic molecules. This equilibrium is upset in various diseases, including cancer. Angiogenesis is virtually nonexistent in healthy adult tissue with the exception of a few physiological processes such as wound healing and the female menstrual cycle.^{21,22} Stable regions of tumor hypoxia may upset this balance and provide the requisite proangiogenic signal. Viable hypoxic cells initiate angiogenesis principally through HIF-1 dependent upregulation of VEGF, although it also facilitates this process through a variety of other mechanisms.²³ We propose that development of hypoxia tolerance through modulation of cell death in the tumor microenvironment may be a common pathway that allows the generation of a persistent angiogenic signal. Whereas in normal cells hypoxia will lead to cell death and thus removal of the proangiogenic signal, hypoxia-tolerant cells maintain their survival and delivery of the signal. Two principle mechanisms have emerged that can explain how cell death in the tumor microenvironment is altered. The first is through suppression of intrinsic cell death pathways normally initiated by hypoxia and the second is through adaptation to the hypoxic environment through decreased energy use and increased energy production.

Hypoxia Tolerance

Suppression of cell death pathways. Hypoxia imposes a stress response that can lead to cell death. In many cell types, hypoxia promotes apoptosis through the induction of genes such as *p53*,⁴ *bik*,²⁴ *bnip3*,^{25,26} and others. However, the cellular decision of life or death is the result of the net balance between proapoptotic and antiapoptotic (survival) signals. It is thought that the very existence of cancer implies suppressed apoptosis and deregulated dependence on survival signals.²⁷ Suppression of proapoptotic signals often occurs through mutations in apoptosis-triggering tumor suppressor genes such as *p53*. Similarly, antiapoptotic (survival)-signaling pathways are often constitutively upregulated through activation of oncogenes such as *ras* or

loss of tumor suppressors such as *PTEN*. Deregulated susceptibility to apoptosis may in itself lead to increased resistance to death induced by environmental stress such as hypoxia. The suppression of apoptosis in cancer cells also may contribute to genomic instability by failure to eliminate damaged cells. This provides cancer cells with an inherent adaptability to stress conditions such as hypoxia and hence substantial responsiveness to the selection pressure that it evokes. Thus, the presence of hypoxic areas in tumors may contribute to malignancy by promoting the clonal expansion of cell variants with a survival advantage in this microenvironment.

The relevance of hypoxia-induced selection pressure has been shown experimentally in several models. Graeber and colleagues⁴ showed that a small number of transformed cells lacking the apoptosis stimulating tumor suppressor *p53* would overtake similar cells expressing *p53* when treated with hypoxia in vitro. Likewise, Kim et al⁵ showed that the exposure of cell cultures to hypoxia greatly accelerated the selection for subpopulations of cells with diminished apoptotic potential. In vivo, hypoxic and apoptotic areas coincided in transplanted tumors expressing wild-type *p53* but not in *p53*-deficient tumors.⁴ Furthermore, the conversion of well-vascularized solid tumors to hypoxic ascites tumors favors the selection of cell variants with mutant *p53*.²⁸ The selection of apoptosis-resistant cells by hypoxia can also occur through *p53*-independent pathways. An example of this was shown in human colorectal cancer cells that acquired sustained resistance to apoptosis after hypoxia exposure in spite of active *p53*.²⁹

Increased Energy Production. As described earlier, one of the means by which cells can become hypoxia tolerant is through selection of cells with mutations in genes that predispose them to hypoxia-induced cell death. However, tumor cells may also become hypoxia tolerant through adaptation of normal physiologic responses. The lack of oxygen necessitates a switch from oxidative phosphorylation to anaerobic glycolysis for adenosine triphosphate (ATP) production, and this switch has been shown to coincide with the oxygen gradient around blood vessels.³¹ Increased glycolysis during hypoxia is facilitated by increasing the activity and expression of proteins in the glycolytic pathway such as glucose transporters (GLUTs),³² phosphoglycerate kinase-1 (PGK-1),³³ and pyruvate kinase M (PK-

M).³⁴ In fact, the hypoxia-induced transcription factor HIF-1 mediates transcriptional activation of the entire glycolytic pathway from glucose uptake to lactate production.⁸ Transcription of HIF-1-responsive genes is stimulated through binding of HIF-1 to a hypoxia response element (HRE) in the gene promoter. The HIF-1 transcription factor itself is regulated by a post-translational mechanism. HIF-1 is a heterodimer consisting of the 2 subunits, HIF-1 α and HIF-1 β , which are both ubiquitously expressed. HIF-1 β protein is stable, whereas HIF-1 α is targeted for ubiquitination by the von Hippel-Lindau tumor suppressor protein (VHL) and rapidly degraded by the proteasome under well-oxygenated conditions.³⁵ VHL recognizes hydroxylized prolyl residues in the HIF-1 α protein, which remain unhydroxylated during hypoxic conditions.^{36,37} Thus, HIF-1 α is stabilized during hypoxia and can dimerize with its partner HIF-1 β to induce transcription of HRE-responsive genes.

Evidence showing the significance of the HIF-1 pathway for tumor cell viability has accumulated over the last few years. HIF-1 α - or β -deficient Chinese hamster ovary cells have been shown to be sensitive to hypoglycaemia and altered redox status by inhibition of cytochrome oxidase,³⁸ and mouse embryo fibroblasts lacking HIF-1 α grow more slowly under hypoxia than wild-type cells.³⁹ Using genetically matched cell lines derived from wild-type and HIF-1 α knockout mice, a proteomic analysis showed that HIF-1 was strictly required for the upregulation of key enzymes in the glycolytic pathway.³⁹ The decreased glycolytic capacity of HIF-deficient cells resulted in dramatically lowered free ATP levels. These findings are in support of a significant role for HIF-1 in the protective adaptation to the tumor microenvironment. This notion has been confirmed *in vivo* because HIF-1-deficient transformed cells are less tumorigenic than wild-type cells and that the resulting xenografts show slower growth. The slower tumor growth in these studies could not be attributed to differences in tumor vascularization, and the authors therefore concluded that it was because of impaired upregulation of glycolysis as observed *in vitro*.^{39,40}

Constitutively upregulated HIF and enhanced glycolysis even under aerobic conditions is a common characteristic of many tumors, suggesting that this pathway may become deregulated as a consequence of tumor progression.⁴¹ The observed reduced growth rates of HIF-deficient tu-

mors provide an explanation for why this may be beneficial for tumor development. Presumably, a constitutively activated glycolytic pathway can contribute to hypoxia tolerance by allowing tumor cells to maintain their energy homeostasis during periods of low-oxygen availability.

Decreased Energy Consumption

Another strategy for increasing survival under conditions of limiting oxygen and energy is to decrease ATP consumption. A well-characterized consequence of hypoxic stress is a pronounced repression in the rate of oxygen consumption and energy turnover.^{42,43} The main ATP-demand pathways in hypoxic cells are the Na⁺/K⁺ ATPase pump, protein synthesis, and degradation and gluconeogenesis.^{44,45} It has been estimated that under severe hypoxia, the ATP demand for protein synthesis drops to about 7% of that of normoxic cells. This drop correlates with a substantial and rapid drop in the rate of protein synthesis.⁴⁶ The rapid kinetics of this response and the fact that it precedes ATP depletion argues for a tightly regulated mechanism that is activated by low oxygen availability.⁴⁷ The molecular pathways responsible for the downregulation of protein synthesis during hypoxia are not yet completely understood. However, recent data suggest the involvement of eukaryotic initiation factors (eIFs) that modulate translation initiation.⁴⁸ The eIFs facilitate the correct assembly of the messenger RNA (mRNA) template, the ribosomal 40S subunit and the aminoacylated initiator transfer RNA (Met-tRNA). A rate-limiting factor in this process is eIF2, which binds Met-tRNA in its energized eIF2-guanosine triphosphate form, and thereby recruits Met-tRNA to the ribosome. The activity of eIF2 is tightly regulated through phosphorylation of the eIF2 α subunit, which inhibits the exchange of guanosine triphosphate for guanosine diphosphate bound to eIF2, and thereby the binding and recruitment of Met-tRNA. Translation has previously been shown to be downregulated through phosphorylation of eIF2 α in response to stress such as virus infection, unfolded proteins, serum starvation, or amino acid deprivation. Recently it was shown in several transformed cell lines that hypoxia induces a rapid phosphorylation of the eIF2 α subunit, which results in reduced protein synthesis.⁴⁹ The phosphorylation of eIF2 α appears to be mediated by the endoplasmic reticulum kinase

PERK, which previously has been known to be activated in response to unfolded proteins in the endoplasmic reticulum. This rapid inhibition in protein translation in response to hypoxia strongly suggests that this pathway is critical for cells to survive during hypoxic exposure. It will be interesting to test if disruption of this response will sensitize cells to hypoxia-induced cell death.

Another rate-limiting eukaryotic initiation factor is eIF4E, which recognizes and binds the m⁷G cap-structure of the 5' mRNA. eIF4E facilitates the bridging of the mRNA to the 40S subunit through participation in the scaffolding eIF4F protein complex. eIF4E is mainly regulated through a set of binding proteins (4E-BPs) that bind eIF4E in their hypophosphorylated form and thereby inhibit its participation in the eIF4F complex. The 4E-BPs have previously been shown to be phosphorylated in response to stimuli such as hormones or growth factors, and dephosphorylated in response to stress such as heat shock, virus infection, or serum starvation, leading to stimulation and repression of translation, respectively. We have recently found that hypoxia disrupts the eIF4F complex through multiple mechanisms including dephosphorylation of the 4E-BPs.⁵⁰ Furthermore, we have also shown that the eIF4E translocates to the cell nucleus and thus becomes unavailable for translation during hypoxic conditions.⁵⁰

The regulation of eIF2 α and eIF4E in response to hypoxia appears to be significantly weaker in human normal fibroblasts than in human tumor cell lines (Wouters, unpublished data, 2002). This finding supports the idea that regulatory responsiveness to hypoxia is an adapted feature of tumor cells that is beneficial for their survival.

Therapy Implications

The fact that hypoxia negatively impacts on therapy is well established. Hypoxic cells are radiation and chemoresistant for a variety of reasons,⁵¹ and thus effective therapy requires strategies to overcome this resistance. Many attempts have been made, most of these focused on trying to restore normal oxygenation (or mimic it) to the tumor. The concept that tumor cells become hypoxic tolerant, and furthermore that the presence of microregions of hypoxia may be advantageous to overall tumor growth, has its own therapeutic implications. New approaches aimed

specifically at exploiting or altering the mechanisms that lead to hypoxia tolerance may provide better efficacy in future therapies

Restore Oxygenation

In the past 40 years, numerous attempts have been made to improve radiotherapy by restoring (or mimicking with hypoxic radiosensitisers) the oxygen supply to tumor cells. This includes such treatments as hyperbaric oxygen breathing, transfusions to improve the haemoglobin level, and electron affinic radiosensitisers. Although early studies showed mixed results, a large meta-analysis of all oxygen-modifying head and neck cancer trials did show a significant improvement in local control and disease-specific survival.^{52,53} This is likely a reflection of the fact that hypoxic cells are radiation resistant, and even modest improvements in oxygenation should improve outcome. More recently, the use of recombinant erythropoietin (EPO) has become subject of study. EPO is a HIF-1-regulated haematopoietic growth factor produced by the kidneys in response to hypoxia. It stimulates the erythrocyte production in the bone marrow. Several trials have been conducted to investigate the effect of recombinant human EPO in patients with low hemoglobin (Hb) levels. A relationship between Hb levels and the response to radiation therapy has been shown for carcinomas of the uterine cervix,⁵⁴⁻⁵⁶ head and neck cancers,⁵⁷⁻⁶¹ lung cancer,⁶²⁻⁶⁴ bladder cancer,⁶⁵⁻⁶⁸ and prostate carcinoma.⁶⁹ In these studies, patients with low Hb levels achieved lower local control rates and survival.

The combination of accelerated radiotherapy with carbogen and nicotinamide, known as the ARCON protocol, is also being evaluated in the clinic. Carbogen (95% O₂ + 5% CO₂) reduces diffusion limited hypoxia and nicotinamide can antagonize vasculature shutdown.^{14,70-74} A significant effect on both locoregional control and disease specific survival for stage T₃-T₄ SCC laryngeal tumors has been reported.^{75,76} In a recent publication an overall local control rate of 80% was reported for T₃ and T₄ larynx carcinomas,⁷⁷ offering possibilities for organ preservation. A phase II trial for bladder cancer also showed better results using the ARCON protocol, compared with historical data.⁷⁸

Despite some success of these approaches, the concept of improving tumor oxygenation ignores

Table 1. Targeting Treatment at Hypoxia

| | |
|----------------------|-------------------------------------------------------------------------------------------------|
| Hypoxia tolerance | Restore cell death pathways Target survival pathways Block adaptation to ATP preservation |
| Viable hypoxic cells | Restore oxygenation Bioreductive drugs Gene therapy with anaerobic bacteria |
| Biological response | Block angiogenesis Block metastasis |

in part the biological effects of hypoxia that may be important in malignancy and treatment response outside of therapy resistance. In other words intrinsic resistance to therapy is only one of the mechanisms by which hypoxia impacts on prognosis. Our increased understanding of the biology of hypoxic cells has led to new ideas for treating hypoxic tumors (Table 1).

Exploit Hypoxia

The concept that hypoxia tolerance in tumors is a selected phenotype that supports angiogenesis provides us with new problems and possibilities for therapy. Instead of attempting to rid tumors of hypoxia, which may be impossible because of hypoxia tolerance and the nature of their vasculature, we can instead attempt to exploit hypoxia for therapeutic advantage. Stable regions of hypoxia in human tumors provide the possibility of directing therapy specifically against this unique feature. Current attempts to exploit this feature of tumors include both pharmaceutical and gene-therapy approaches.

Bioreductive Drugs

Bioreductive drugs are compounds that are reduced by enzymes to their toxic, active metabolites. They are designed such that this metabolism occurs only or preferentially in the absence of oxygen. The use of these drugs in combination with traditional therapies has the potential to greatly improve treatment outcome by increasing cytotoxicity to the hypoxic tumor fraction. In theory, such an approach can be superior to an alternative therapy that would fully reoxygenate the tumor.⁸⁰ Tirapazamine is the leading compound in this class of agents and has shown promising results in a number of clinical trials when used in combination with cisplatin and/or radio-

therapy. A wide variety of cell lines are sensitive to tirapazamine, regardless of their p53 status, and require 50 to 150 times higher dose for the same toxicity under aerobic conditions.^{81,82} For a more detailed discussion of bioreductive agent therapies, see the accompanying article by Stratford et al in this issue.

Gene Therapy With Bacterial Vectors

The aim of gene therapy is to transfer genetic material to the tumor cell or its microenvironment in quantities sufficient to obtain a therapeutic level of expression. However, strategies devised to date have limited efficiency, most notably because of deficiencies in the delivery systems used. A recent approach to this problem uses the concept of targeting anaerobic bacteria to the hypoxic/necrotic areas of solid tumors. Currently, both *Clostridium spp* and attenuated *Salmonella typhimurium* auxotrophs are being investigated as systems to deliver antitumor compounds specifically to the tumor site.^{83,84} The latter strain grows under aerobic and anaerobic conditions, with selectivity for tumors reported as a consequence of its auxotrophic nature. The specificity of clostridia for tumors resides in its obligate requirement for anaerobic conditions. Intravenously injected spores of a nonpathogenic clostridial species have been shown to localize to, and germinate in, the hypoxic/necrotic regions of solid tumors. Although growth alone in the tumor is not sufficient for therapeutic efficacy, the possibility now exists to engineer *Clostridium spp* to produce a variety of therapeutic proteins with anticancer properties. Clostridia can thus be used as highly selective in situ cell factories able to produce and secrete antitumor therapeutics specifically at the tumor site. Moreover, it has been shown that the immune response does not hinder repeated administration of clostridial spores, that colonization can be improved using vascular targeting, and that gene expression can be stopped at any time using suitable antibiotics. We and others showed that it is possible to express therapeutic proteins, not only in vitro but also in vivo after administration of the recombinant clostridia to tumor-bearing animals.^{85,86}

Target Biological Responses to Hypoxia

We have proposed that one of the principle reasons that tumors contain hypoxic cells is the fact that these cells can provide a prolonged angio-

genic signal. One can thus envisage a situation in which non-proliferating or even non-clonogenic hypoxic cells contribute in a positive way to tumor growth. The concept that hypoxia tolerance is critically important for tumor growth suggests that interfering with either the mechanisms that lead to hypoxia tolerance or the biological consequences of the resulting hypoxic areas in tumors (angiogenesis) may become new effective means of treatment. Therapeutic interventions to counteract these biological responses are possible at different levels.

Block Angiogenesis

Perhaps the most important consequence of tumor hypoxia is the induction of angiogenesis. Progressive growth of solid tumors is largely dependent on this process⁸⁷ and thus antiangiogenic therapy may relieve much of the impact of hypoxia on prognosis. The complex process of angiogenesis offers potential therapeutic targets at different levels.⁸⁸

VEGF is perhaps the most important factor in tumors for promoting compensatory angiogenesis in circumstances of oxygen shortage.⁸⁹ It promotes endothelial cell migration, modulates proteases needed for basement degradation, and stimulates plasminogen expression. Hypoxic upregulation of VEGF occurs as a result of both increased transcription mediated by HIF-1 and increased mRNA stability dependent on the 3' untranslated region.^{90,91} VEGF receptors have been shown to be upregulated in surrounding endothelial cells.⁹² Therefore, VEGF, its receptors (flt-1 and flk-1) and the signal transduction pathway present realistic therapeutic targets. Several therapeutic strategies aimed at targeting the process of angiogenesis, including those aimed at interfering with VEGF signaling pathways, are currently under active investigation both in the laboratory and the clinic. This topic is the subject of another article in the current issue (Siemann and Shi, in this issue).

One interesting aspect of antiangiogenic therapies possibly related to the concept of hypoxia tolerance is that there are examples of gradual loss of response, especially when drugs are administered as monotherapies. Several recent reports^{107,108} support this notion and provide a possible explanation. Kerbel and colleagues¹⁰⁸ showed that the genetic background of a tumor cell (in particular the presence/absence of p53)

may be an important determinant of response to antiangiogenic therapy. They concluded that loss of p53 may allow tumor cells to survive the temporary inhibition of angiogenesis as a consequence of their reduced apoptotic potential during hypoxia. Antiangiogenic therapy is expected to result in increased tumor hypoxia, and thus tumors that are better able to survive hypoxia would be expected to maintain a growth advantage. As discussed earlier, the reduced reliance on vascular supply through modulation of cell death during hypoxia can occur through many mechanisms including changes in the HIF-1 pathway.¹⁰⁹

Block Tolerance

The recent suggestion that even antiangiogenic strategies have reduced efficacy against tumors that have developed mechanisms of increased hypoxia tolerance suggests it may be necessary to target an even earlier step in this system. One obvious strategy is to interfere with the mechanisms that tumor cells have used to modulate their sensitivity to cell death in response to hypoxia. Some approaches are based on the knowledge that several genes, when mutated, contribute not only to tumor progression but also to survival under hypoxic conditions. Gain-of-function mutations in key oncogenes and/or loss-of-function mutations in tumor suppressor genes can prevent commitment of cells to apoptosis. Perhaps the best-characterized survival signaling pathway is mediated by PI3-kinase (PI3K-PTEN-AKT-FRAP/MTOR pathway). This pathway has been implicated in both the response of cells to hypoxia and to angiogenesis.¹¹⁰ Pharmacologic agents that inhibit PI3K (Wortmannin or LY294002) or its downstream effectors FRAP/MTOR (rapamycin) have been shown to have some therapeutic efficacy.¹¹¹ Altering the malignant phenotype by blocking dominant negative oncogenes that are implicated in the hypoxic response (such as *myc* and *ras*) at transcriptional or translational levels is also an attractive target being evaluated in several clinical trials. Methods involve the use of antisense oligonucleotides,¹¹² ribozymes,¹¹³ and intracellular single-chain antibodies.¹¹⁴ Antibodies to HER2/neu or the epidermal growth factor receptor, both of which can provide survival and angiogenic signals, are also attractive candidates as contributors to hypoxic cell survival. These antibodies are being tested

and have shown some significant efficacy against angiogenesis *in vivo*.^{115,116} Based on the fact that p53 induces apoptosis in response to hypoxia, perhaps the most attractive method to restore cell death is to restore p53 function. Many investigations using gene therapy with p53 are already underway.^{117,118} Other attractive approaches involve the conversion of anti-apoptotic or proliferative signals into signals that trigger apoptosis.^{119,120}

Another important mechanism of hypoxia tolerance arises through the increased ability to regulate ATP supply during hypoxic exposure. As discussed earlier, hypoxic tumor cells are able to decrease their requirements for ATP by shutting down overall protein synthesis while at the same time inducing a transition from oxidative phosphorylation to anaerobic glycolysis for the generation of ATP. Interfering with either of these 2 processes would be expected to selectively target hypoxia tolerant cells. The process of translation initiation is facilitated by the eukaryotic initiation factors, and several of these factors are controlled through upstream signaling pathways. Thus, it may be possible to interfere with this response indirectly with agents that influence these upstream pathways. For example, translation initiation is increased in response to PI3 kinase signaling through FRAP/mTOR. Deregulation of inhibited translation initiation during hypoxia would be expected to result in increased ATP consumption and thus make tumor cells less able to survive prolonged periods of hypoxia.

Similarly, it may be possible to interfere with ATP generation in hypoxic tumor cells. Tumors in general have increased rates of glycolysis, a fact that may be related to hypoxia tolerance. Targeting of the enzymes or substrates involved in this well understood pathway would thus also be expected to specifically target hypoxic cells or at least reduce the tolerance of tumor cells to hypoxia. Recent results showing reduced tumor growth in transformed cells that are unable to induce anaerobic glycolysis as a result of loss of HIF-1 support this idea.³⁹

References

1. Gray LH, Conger AD, Ebert M, et al: Concentration of oxygen dissolved in tissues at the time of irradiation as a factor in radiotherapy. *Br J Radiol* 26:638-648, 1953
2. Young SD, Marshall RS, Hill RP: Hypoxia induces DNA overreplication and enhances metastatic potential of murine tumor cells. *Proc Natl Acad Sci U S A* 85:9533-9537, 1988
3. De Jaeger K, Kavanagh MC, Hill RP: Relationship of hypoxia to metastatic ability in rodent tumours. *Br J Cancer* 84:1280-1285, 2001
4. Graeber TG, Osmanian C, Jacks T, et al: Hypoxia-mediated selection of cells with diminished apoptotic potential in solid tumours. *Nature* 379:88-91, 1996
5. Kim CY, Tsai MH, Osmanian C, et al: Selection of human cervical epithelial cells that possess reduced apoptotic potential to low-oxygen conditions. *Cancer Res* 57:4200-4204, 1997
6. Rofstad EK, Danielsen T: Hypoxia-induced metastasis of human melanoma cells: Involvement of vascular endothelial growth factor-mediated angiogenesis. *Br J Cancer* 80:1697-1707, 1999
7. Royds JA, Dower SK, Qwarnstrom EE, et al: Response of tumour cells to hypoxia: Role of p53 and NFkB. *Mol Pathol* 51:55-61, 1998
8. Semenza GL: Hypoxia, clonal selection, and the role of HIF-1 in tumor progression. *Crit Rev Biochem Mol Biol* 35:71-103, 2000
9. Hockel M, Schlenger K, Aral B, et al: Association between tumor hypoxia and malignant progression in advanced cancer of the uterine cervix. *Cancer Res* 56:4509-15, 1996
10. Brizel DM, Sibley GS, Prosnitz LR, et al: Tumor hypoxia adversely affects the prognosis of carcinoma of the head and neck. *Int J Radiat Oncol Biol Phys* 38:285-289, 1997
11. Nordsmark M, Alsner J, Keller J, et al: Hypoxia in human soft tissue sarcomas: Adverse impact on survival and no association with p53 mutations. *Br J Cancer* 84:1070-1075, 2001
12. Semenza GL: Regulation of hypoxia-induced angiogenesis: A chaperone escorts VEGF to the dance. *J Clin Invest* 108:39-40, 2001
13. Lartigau E, Le Ridant AM, Lambin P, et al: Oxygenation of head and neck tumors. *Cancer* 71:2319-2325, 1993
14. Martin L, Lartigau E, Weeger P, et al: Changes in the oxygenation of head and neck tumors during carbogen breathing. *Radiother Oncol* 27:123-130, 1993
15. Nordsmark M, Overgaard M, Overgaard J: Pretreatment oxygenation predicts radiation response in advanced squamous cell carcinoma of the head and neck. *Radiother Oncol* 41:31-39, 1996
16. Kimura H, Braun RD, Ong ET, et al: Fluctuations in red cell flux in tumor microvessels can lead to transient hypoxia and reoxygenation in tumor parenchyma. *Cancer Res* 56:5522-5528, 1996
17. Brown EB, Campbell RB, Tsuzuki Y, et al: In vivo measurement of gene expression, angiogenesis and physiological function in tumors using multiphoton laser scanning microscopy. *Nat Med* 7:864-868, 2001
18. Rofstad EK: Microenvironment-induced cancer metastasis. *Int J Radiat Biol* 76:589-605, 2000
19. Yuan J, Glazer PM: Mutagenesis induced by the tumor microenvironment. *Mutat Res* 400:439-446, 1998
20. Hockel M, Vaupel P: Biological consequences of tumor hypoxia. *Semin Oncol* 28:36-41, 2001
21. Ferrara N, Chen H, Davis-Smyth T, et al: Vascular endothelial growth factor is essential for corpus luteum angiogenesis. *Nat Med* 4:336-340, 1998

22. Hobson B, Denekamp J: Endothelial proliferation in tumours and normal tissues: continuous labelling studies. *Br J Cancer* 49:405-413, 1984
23. Semenza GL: Regulation of mammalian O₂ homeostasis by hypoxia-inducible factor 1. *Annu Rev Cell Dev Biol* 15:551-578, 1999
24. Koong AC, Denko NC, Hudson KM, et al: Candidate genes for the hypoxic tumor phenotype. *Cancer Res* 60:883-887, 2000
25. Sowter HM, Ratcliffe PJ, Watson P, et al: HIF-1-dependent regulation of hypoxic induction of the cell death factors BNIP3 and NIX in human tumors. *Cancer Res* 61:6669-6673, 2001
26. Bruick RK: Expression of the gene encoding the proapoptotic Nip3 protein is induced by hypoxia. *Proc Natl Acad Sci U S A* 97:9082-9087, 2000
27. Green DR, Evan GI: A matter of life and death. *Cancer Cell* 1:19-30, 2002
28. Magnusson KP, Satalino R, Qian W, et al: Is conversion of solid into more anoxic ascites tumors associated with p53 inactivation? *Oncogene* 17:2333-2337, 1998
29. Kinoshita M, Johnson DL, Shatney CH, et al: Cancer cells surviving hypoxia obtain hypoxia resistance and maintain anti-apoptotic potential under reoxygenation. *Int J Cancer* 91:322-326, 2001
30. Liang BC: Effects of hypoxia on drug resistance phenotype and genotype in human glioma cell lines. *J Neurooncol* 29:149-155, 1996
31. Walenta S, Snyder S, Haroon ZA, et al: Tissue gradients of energy metabolites mirror oxygen tension gradients in a rat mammary carcinoma model. *Int J Radiat Oncol Biol Phys* 51:840-848, 2001
32. Sivitz WI, Lund DD, Yorek B, et al: Pretranslational regulation of two cardiac glucose transporters in rats exposed to hypobaric hypoxia. *Am J Physiol* 263:E562-569, 1992
33. Firth JD, Ebert BL, Pugh CW, et al: Oxygen-regulated control elements in the phosphoglycerate kinase 1 and lactate dehydrogenase A genes: Similarities with the erythropoietin 3' enhancer. *Proc Natl Acad Sci U S A* 91:6496-500, 1994
34. Semenza GL, Roth PH, Fang HM, et al: Transcriptional regulation of genes encoding glycolytic enzymes by hypoxia-inducible factor 1. *J Biol Chem* 269:23757-23763, 1994
35. Maxwell PH, Wiesener MS, Chang GW, et al: The tumour suppressor protein VHL targets hypoxia-inducible factors for oxygen-dependent proteolysis. *Nature* 399:271-275, 1999
36. Jaakkola P, Mole DR, Tian YM, et al: Targeting of HIF- α to the von Hippel-Lindau ubiquitylation complex by O₂-regulated prolyl hydroxylation. *Science* 292:468-472, 2001
37. Ivan M, Kondo K, Yang H, et al: HIF α targeted for VHL-mediated destruction by proline hydroxylation: Implications for O₂ sensing. *Science* 292:464-468, 2001
38. Williams KJ, Telfer BA, Airley RE, et al: A protective role for HIF-1 in response to redox manipulation and glucose deprivation: Implications for tumorigenesis. *Oncogene* 21:282-290, 2002
39. Seagroves TN, Ryan HE, Lu H, et al: Transcription factor HIF-1 is a necessary mediator of the pasteur effect in mammalian cells. *Mol Cell Biol* 21:3436-3444, 2001
40. Williams KJ, Telfer BA, Airley RE, et al: A protective role for HIF-1 in response to redox manipulation and glucose deprivation: Implications for tumorigenesis. *Oncogene* 21:282-290, 2002
41. Dang CV, Semenza GL: Oncogenic alterations of metabolism. *Trends Biochem Sci* 24:68-72, 1999
42. Sutherland R, Freyer J, Mueller-Klieser W, et al: Cellular growth and metabolic adaptations to nutrient stress environments in tumor microregions. *Int J Radiat Oncol Biol Phys* 12:611-615, 1986
43. Koch CJ: Oxygen effects in radiobiology. *Adv Exp Med Biol* 157:123-144, 1982
44. Sutherland R, Freyer J, Mueller-Klieser W, et al: Cellular growth and metabolic adaptations to nutrient stress environments in tumor microregions. *Int J Radiat Oncol Biol Phys* 12:611-615, 1986
45. Hochachka PW, Buck LT, Doll CJ, et al: Unifying theory of hypoxia tolerance: Molecular/metabolic defense and rescue mechanisms for surviving oxygen lack. *Proc Natl Acad Sci U S A* 93:9493-9498, 1996
46. Pettersen EO, Juul NO, Ronning OW: Regulation of protein metabolism of human cells during and after acute hypoxia. *Cancer Res* 46:4346-4351, 1986
47. Lefebvre VH, Van Steenbrugge M, Beckers V, et al: Adenine nucleotides and inhibition of protein synthesis in isolated hepatocytes incubated under different pO₂ levels. *Arch Biochem Biophys* 304:322-331, 1993
48. Hershey JW, Merrick WC: Pathway and mechanism of initiation of protein synthesis, in Sonenberg N, Hershey JW, Mathews MB (eds): *Translational control of gene expression*. Cold Spring Harbor, Cold Spring Harbor Laboratory Press, 2000, pp 33-88
49. Koumenis C, Naczki C, Koritzinsky M, et al: Regulation of protein synthesis by hypoxia via activation of the protein kinase PERK and phosphorylation of eIF2 α . *Mol Cell Biol* 22:7405-7416, 2002
50. Koritzinsky M, Koumenis C, Sonenberg N, et al: Regulation of protein translation during hypoxic conditions. Paper presented at the 93rd Annual Meeting of the American Association for Cancer Research, San Francisco, CA, April 6-10, 2002
51. Wouters BG, Wepler SA, Koritzinsky M, et al: Hypoxia as a target for combined modality treatments. *Eur J Cancer* 38:240-257, 2002
52. Overgaard J, Horsman MR: Modification of hypoxia-induced radioresistance in tumors by the use of oxygen and sensitizers. *Semin Radiat Oncol* 6:10-21, 1996
53. Saunders M, Dische S: Clinical results of hypoxic cell radiosensitisation from hyperbaric oxygen to accelerated radiotherapy, carbogen and nicotinamide. *Br J Cancer* 27:S271-S278, 1996 (Suppl)
54. Overgaard J: Sensitization of hypoxic tumour cells—Clinical experience. *Int J Radiat Biol* 56:801-811, 1989
55. Pedersen D, Sogaard H, Overgaard J, et al: Prognostic value of pretreatment factors in patients with locally advanced carcinoma of the uterine cervix treated by radiotherapy alone. *Acta Oncol* 34:787-795, 1995
56. Girinski T, Pejovic-Lenfant MH, Bourhis J, et al: Prognostic value of hemoglobin concentrations and blood transfusions in advanced carcinoma of the cervix treated

- by radiation therapy: Results of a retrospective study of 386 patients. *Int J Radiat Oncol Biol Phys* 16:37-42, 1989
57. Fein DA, Lee WR, Hanlon AL, et al: Pretreatment hemoglobin level influences local control and survival of T1-T2 squamous cell carcinomas of the glottic larynx. *J Clin Oncol* 13:2077-2083, 1995
 58. Lee WR, Berkey B, Marcial V, et al: Anemia is associated with decreased survival and increased locoregional failure in patients with locally advanced head and neck carcinoma: A secondary analysis of RTOG 85-27. *Int J Radiat Oncol Biol Phys* 42:1069-1075, 1998
 59. van Acht MJ, Hermans J, Boks DE, et al: The prognostic value of hemoglobin and a decrease in hemoglobin during radiotherapy in laryngeal carcinoma. *Radiother Oncol* 23:229-235, 1992
 60. Warde P, O'Sullivan B, Bristow RG, et al: T1/T2 glottic cancer managed by external beam radiotherapy: The influence of pretreatment hemoglobin on local control. *Int J Radiat Oncol Biol Phys* 41:347-353, 1998
 61. Dubray B, Mosseri V, Brunin F, et al: Anemia is associated with lower local-regional control and survival after radiation therapy for head and neck cancer: a prospective study. *Radiology* 201:553-558, 1996
 62. Dische S, Warburton MF, Saunders MI: Radiation myelitis and survival in the radiotherapy of lung cancer. *Int J Radiat Oncol Biol Phys* 15:75-81, 1988
 63. Macchiarini P, Silvano G, Janni A, et al: Results of treatment and lessons learned from pathologically staged T4 non-small cell lung cancer. *J Surg Oncol* 47:209-214, 1991
 64. Sasai K, Ono K, Hiraoka M, et al: The effect of arterial oxygen content on the results of radiation therapy for epidermoid bronchogenic carcinoma. *Int J Radiat Oncol Biol Phys* 16:1477-1481, 1989
 65. Cole CJ, Pollack A, Zagars GK, et al: Local control of muscle-invasive bladder cancer: Preoperative radiotherapy and cystectomy versus cystectomy alone. *Int J Radiat Oncol Biol Phys* 32:331-340, 1995
 66. Hannisdal E, Fossa SD, Host H: Blood tests and prognosis in bladder carcinomas treated with definitive radiotherapy. *Radiother Oncol* 27:117-122, 1993
 67. Wijkstrom H, Nilsson B, Tribukait B: DNA analysis in predicting survival of irradiated patients with transitional cell carcinoma of bladder. *Br J Urol* 69:49-55, 1992
 68. Greven KM, Solin IJ, Hanks GE: Prognostic factors in patients with bladder carcinoma treated with definitive irradiation. *Cancer* 65:908-912, 1990
 69. Dunphy EP, Petersen IA, Cox RS, et al: The influence of initial hemoglobin and blood pressure levels on results of radiation therapy for carcinoma of the prostate. *Int J Radiat Oncol Biol Phys* 16:1173-1178, 1989
 70. Rojas A, Joiner MC, Denekamp J: Extrapolations from laboratory and preclinical studies for the use of carbogen and nicotinamide in radiotherapy. *Radiother Oncol* 24:123-124, 1992
 71. Chaplin DJ, Horsman MR, Trotter MJ: Effect of nicotinamide on the microregional heterogeneity of oxygen delivery within a murine tumor. *J Natl Cancer Inst* 82:672-676, 1990
 72. Rojas A: Radiosensitization with normobaric oxygen and carbogen. *Radiother Oncol* 1:65-70, 1991 (Suppl)
 73. Horsman MR, Nordmark M, Khalil AA, et al: Reducing acute and chronic hypoxia in tumours by combining nicotinamide with carbogen breathing. *Acta Oncol* 33:371-376, 1994
 74. Laurence VM, Ward R, Dennis IF, et al: Carbogen breathing with nicotinamide improves the oxygen status of tumours in patients. *Br J Cancer* 72:198-205, 1995
 75. Kaanders JH, Pop LA, Marres HA, et al: Accelerated radiotherapy with carbogen and nicotinamide (ARCON) for laryngeal cancer. *Radiother Oncol* 48:115-122, 1998
 76. Bussink J, Kaanders JH, Van der Kogel AJ: Clinical outcome and tumour microenvironmental effects of accelerated radiotherapy with carbogen and nicotinamide. *Acta Oncol* 38:875-882, 1999
 77. Kaanders JH, Pop LA, Marres HA, et al: ARCON: Experience in 215 patients with advanced head-and-neck cancer. *Int J Radiat Oncol Biol Phys* 52:769-778, 2002
 78. Hoskin PJ, Saunders MI, Dische S: Hypoxic radiosensitizers in radical radiotherapy for patients with bladder carcinoma: Hyperbaric oxygen, misonidazole, and accelerated radiotherapy, carbogen, and nicotinamide. *Cancer* 86:1322-1328, 1999
 79. Bernier J, Denekamp J, Rojas A, et al: ARCON: Accelerated radiotherapy with carbogen and nicotinamide in head and neck squamous cell carcinomas. The experience of the Co-operative group of radiotherapy of the european organization for research and treatment of cancer (EORTC). *Radiother Oncol* 55:111-1199, 2000
 80. Brown JM, Koong A: Therapeutic advantage of hypoxic cells in tumors: A theoretical study. *J Natl Cancer Inst* 83:178-185, 1991
 81. Wouters BG, Wang LH, Brown JM: Tirapazamine: a new drug producing tumor specific enhancement of platinum-based chemotherapy in non-small-cell lung cancer. *Ann Oncol* 10:S29-33, 1999
 82. Brown JM: SR 4233 (tirapazamine): A new anticancer drug exploiting hypoxia in solid tumours. *Br J Cancer* 67:1163-1170, 1993
 83. Lambin P, Theys J, Landuyt W, et al: Colonisation of Clostridium in the body is restricted to hypoxic and necrotic areas of tumors. *Anaerobe* 4:183-188, 1998
 84. Pawelek JM, Low KB, Bermudes D: Tumor-targeted Salmonella as a novel anticancer vector. *Cancer Res* 57:4537-4544, 1997
 85. Lemmon MJ, van Zijl P, Fox ME, et al: Anaerobic bacteria as a gene delivery system that is controlled by the tumor microenvironment. *Gene Ther* 4:791-796, 1997
 86. Theys J, Landuyt W, Nuyts S, et al: Specific targeting of cytosine deaminase to solid tumors by engineered Clostridium acetobutylicum. *Cancer Gene Ther* 8:294-297, 2001
 87. Folkman J: Tumor angiogenesis: therapeutic implications. *N Engl J Med* 285:1182-1186, 1971
 88. McNamara DA, Harmey JH, Walsh TN, et al: Significance of angiogenesis in cancer therapy. *Br J Surg* 85:1044-1055, 1998
 89. Shweiki D, Itin A, Soffer D, Keshet E: Vascular endothelial growth factor induced by hypoxia may mediate hypoxia-initiated angiogenesis. *Nature* 359:843-845, 1992
 90. Stein I, Neeman M, Shweiki D, Itin A, et al: Stabilization of vascular endothelial growth factor mRNA by hypoxia

- and hypoglycemia and coregulation with other ischemia-induced genes. *Mol Cell Biol* 15:5363-5368, 1995
91. Forsythe JA, Jiang BH, Iyer NV, et al: Activation of vascular endothelial growth factor gene transcription by hypoxia-inducible factor 1. *Mol Cell Biol* 16:4604-4613, 1996
 92. Plate KH, Breier G, Millauer B, Ullrich A, Risau W: Up-regulation of vascular endothelial growth factor and its cognate receptors in a rat glioma model of tumor angiogenesis. *Cancer Res* 53:5822-5827, 1993
 93. Fong TA, Shawver LK, Sun L, et al: SU5416 is a potent and selective inhibitor of the vascular endothelial growth factor receptor (Flk-1/KDR) that inhibits tyrosine kinase catalysis, tumor vascularization, and growth of multiple tumor types. *Cancer Res* 59:99-106, 1999
 94. Laird AD, Vajkoczy P, Shawver LK, et al: SU6668 is a potent antiangiogenic and antitumor agent that induces regression of established tumors. *Cancer Res* 60:4152-4160, 2000
 95. Wood JM, Bold G, Buchdunger E, et al: PTK787/ZK 222584, a novel and potent inhibitor of vascular endothelial growth factor receptor tyrosine kinases, impairs vascular endothelial growth factor-induced responses and tumor growth after oral administration. *Cancer Res* 60:2178-2189, 2000
 96. Usman N, Blatt LM: Nuclease-resistant synthetic ribozymes: developing a new class of therapeutics. *J Clin Invest* 106:1197-1202, 2000
 97. Rubenstein JL, Kim J, Ozawa T, et al: Anti-VEGF antibody treatment of glioblastoma prolongs survival but results in increased vascular cooption. *Neoplasia* 2:306-314, 2000
 98. O'Reilly MS: Angiostatin: An endogenous inhibitor of angiogenesis and of tumor growth. *Exs* 79:273-294, 1997
 99. O'Reilly MS, Boehm T, Shing Y, et al: Endostatin: An endogenous inhibitor of angiogenesis and tumor growth. *Cell* 88:277-285, 1997
 100. Figg WD, Dahut W, Duray P, et al: A randomized phase II trial of thalidomide, an angiogenesis inhibitor, in patients with androgen-independent prostate cancer. *Clin Cancer Res* 7:1888-1893, 2001
 101. Zetter BR: Angiogenesis and tumor metastasis. *Annu Rev Med* 49:407-424, 1998
 102. Min HY, Doyle LV, Vitt CR, et al: Urokinase receptor antagonists inhibit angiogenesis and primary tumor growth in syngeneic mice. *Cancer Res* 56:2428-2433, 1996
 103. Li H, Lu H, Griscelli F, et al: Adenovirus-mediated delivery of a uPA/uPAR antagonist suppresses angiogenesis-dependent tumor growth and dissemination in mice. *Gene Ther* 5:1105-1113, 1998
 104. Eliceiri BP, Cheresh DA: The role of alphaV integrins during angiogenesis: Insights into potential mechanisms of action and clinical development. *J Clin Invest* 103:1227-1230, 1999
 105. Reynolds LE, Wyder L, Lively JC, et al: Enhanced pathological angiogenesis in mice lacking beta3 integrin or beta3 and beta5 integrins. *Nat Med* 8:27-34, 2002
 106. Carmeliet P: Integrin indecision. *Nat Med* 8:14-6, 2002
 107. Yu JL, Rak JW, Coomber BL, et al: Effect of p53 status on tumor response to antiangiogenic therapy. *Science* 295:1526-1528, 2002
 108. Kerbel RS, Yu J, Tran J, et al: Possible mechanisms of acquired resistance to anti-angiogenic drugs: Implications for the use of combination therapy approaches. *Cancer Metastasis Rev* 20:79-86, 2001
 109. Yu JL, Rak JW, Carmeliet P, et al: Heterogeneous vascular dependence of tumor cell populations. *Am J Pathol* 158:1325-1334, 2001
 110. Zundel W, Schindler C, Haas-Kogan D, et al: Loss of PTEN facilitates HIF-1-mediated gene expression. *Genes Dev* 14:391-396, 2000
 111. Zhong H, Chiles K, Feldser D, et al: Modulation of hypoxia-inducible factor 1alpha expression by the epidermal growth factor/phosphatidylinositol 3-kinase/PTEN/AKT/FRAP pathway in human prostate cancer cells: Implications for tumor angiogenesis and therapeutics. *Cancer Res* 60:1541-1545, 2000
 112. Chang EH, Miller PS, Cushman C, et al: Antisense inhibition of ras p21 expression that is sensitive to a point mutation. *Biochemistry* 30:8283-8286, 1991
 113. Feng M, Cabrera G, Deshane J, et al: Neoplastic reversion accomplished by high efficiency adenoviral-mediated delivery of an anti-ras ribozyme. *Cancer Res* 55:2024-2028, 1995
 114. Richardson JH, Marasco WA: Intracellular antibodies: Development and therapeutic potential. *Trends Biotechnol* 13:306-310, 1995
 115. Izumi Y, Xu L, di Tomaso E, et al: Tumour biology: Herceptin acts as an anti-angiogenic cocktail. *Nature* 416:279-280, 2002
 116. Herbst RS, Langer CJ: Epidermal growth factor receptors as a target for cancer treatment: The emerging role of IMC-C225 in the treatment of lung and head and neck cancers. *Semin Oncol* 29:27-36, 2002
 117. Eastham JA, Hall SJ, Schgal I, et al: In vivo gene therapy with p53 or p21 adenovirus for prostate cancer. *Cancer Res* 55:5151-5155, 1995
 118. Roth JA, Nguyen D, Lawrence DD, et al: Retrovirus-mediated wild-type p53 gene transfer to tumors of patients with lung cancer. *Nat Med* 2:985-991, 1996
 119. Vigneri P, Wang JY: Induction of apoptosis in chronic myelogenous leukemia cells through nuclear entrapment of BCR-ABL tyrosine kinase. *Nat Med* 7:228-234, 2001
 120. Chen YN, Sharma SK, Ramsey TM, et al: Selective killing of transformed cells by cyclin/cyclin-dependent kinase 2 antagonists. *Proc Natl Acad Sci U S A* 96:4325-4329, 1999