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From novel discovery tools and biomarkers to precision medicine—basic cardiovascular science highlights of 2021/22

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Abstract

Here, we review the highlights of cardiovascular basic science published in 2021 and early 2022 on behalf of the European Society of Cardiology Council for Basic Cardiovascular Science. We begin with non-coding RNAs which have emerged as central regulators cardiovascular biology, and then discuss how technological developments in single-cell 'omics are providing new insights into cardiovascular development, inflammation, and disease. We also review recent discoveries on the biology of extracellular vesicles in driving either protective or pathogenic responses. The Nobel Prize in Physiology or Medicine 2021 recognized the importance of the molecular basis of mechanosensing and here we review breakthroughs in cardiovascular sensing of mechanical force. We also summarize discoveries in the field of atherosclerosis including the role of clonal haematopoiesis of indeterminate potential, and new mechanisms of crosstalk between hyperglycaemia, lipid mediators, and inflammation. The past 12 months also witnessed major advances in the field of cardiac arrhythmia including new mechanisms of fibrillation. We also focus on inducible pluripotent stem cell technology which has demonstrated disease causality for several genetic polymorphisms in long-QT syndrome and aortic valve disease, paving the way for

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personalized medicine approaches. Finally, the cardiovascular community has continued to better understand COVID-19 with significant advancement in our knowledge of cardiovascular tropism, molecular markers, the mechanism of vaccine-induced thrombotic complications and new anti-viral therapies that protect the cardiovascular system.

Keywords

Cardiology • Vascular • Biomarkers • Precision medicine

1. Introduction

The aim of this review from the European Society of Cardiology (ESC) Council for Basic Cardiovascular Science is to highlight the most noteworthy developments over the past year, in the field of cardiovascular basic science. The cited reports were selected as representative examples of studies which provided robust evidence for particularly novel insights. *Cardiovascular Research* previously reviewed the highlights of 2020 divided into vascular and cardiac topics,^{1,2} but here we integrate both areas to generate the Basic Cardiovascular Science Highlights of 2021/22.

2. Cardiovascular RNA universe

2.1 Non-coding RNAs

In addition to the role of messenger RNA (mRNAs) in the 'central dogma' of molecular biology as a template for protein synthesis, the RNA universe also contains multiple constellations of microRNAs (miRNAs; miRs), long non-coding RNAs (lncRNAs) and circular RNAs (circRNAs) that control fundamental processes of life. These RNA species adopt complex structures and interact with nucleotides, proteins and lipids to control multiple functions including chromatin structure, transcription, RNA splicing and stability, intracellular signalling, and organelle dynamics. Research reported in 2021 has provided further insight into the role of miRNAs, lncRNAs, and circRNAs in the regulation of vascular remodelling and cardiac disease. Using both single-cell (sc) and bulk RNA-sequencing to investigate transcriptional changes associated with endothelial-to-mesenchymal transition (EndMT), Monteiro et al. identified for the first time the genomic locus hosting the lncRNA MIR503HG as necessary to maintain endothelial cell (EC) identity and function.³ In a series of our loss- and gain-of-function experiments the group demonstrated that loss of lncRNA is a causal event in EndMT observed in pulmonary arterial hypertension (PAH) in association with vascular remodelling (Figure 1). Furthermore, located upstream from the vascular smooth muscle cell (vSMC)-associated miR-143 and -145 cluster, the lncRNA CARMN (Cardiac Mesoderm Enhancer-associated Noncoding RNA) was recently identified as key regulator of vSMC function and the pathophysiology of atherosclerotic disease.⁴ Crucially, while crosstalk between lncRNA host genes and coupled miRNAs is often seen, CARMN was found to function independently from miR-143/-145 in regulating vSMC and activating a pro-atherogenic proliferative state (Figure 1).

Gong et al.⁵ identified in atherosclerotic mouse models a novel circRNA, circEys2, involved in vascular remodelling through the targeted inhibition of alternative mRNA splicing. By performing loss- and gain-of-function mutation analyses in vascular smooth muscle cells, circEys2 was shown to enhance cell proliferation and migration and blunt apoptosis and differentiation. Furthermore, silencing of circEys2 prevented neointima formation while circEys2 overexpression enhanced neointimal hyperplasia in an *in vivo* model of carotid artery injury. The role of miRNAs in atherosclerosis progression was examined by

Liu et al. by describing the role of the Nuclear Factor of Activated T-cell isoform c3 (NFATc3)/miR-204 axis in the regulation of foam cell formation in atherosclerosis. Using genetically modified mice, they showed that NFATc3 prevents macrophage foam cell formation and limits the expression of scavenger receptors SR-A and CD36 by inducing expression of the microRNA miR-204,⁶ suggesting the NFATc3/miR-204 axis as a potential therapeutic target to reduce plaque formation. In a separate study involving macrophages, Schober et al. illuminated the circadian patterns of myocardial infarction (MI) by evaluating macrophage-related miRNAs. They evidence, in a murine model of atherosclerosis, that macrophage miR-21 drives circadian regulation of macrophage apoptosis by targeting proapoptotic Xaf1 (XIAP-associated factor 1), thereby regulating plaque composition and susceptibility to rupture.⁷ Further studies in a murine model of pressure-overload heart failure have also found a key role for macrophage miR-21 in modulating cardiac fibrosis by regulating macrophage polarization towards a pro-inflammatory (M1) phenotype.⁸ In addition, Hinkel et al. identified a pivotal role of miR-132 in the mediation of pathologic cardiac hypertrophy in a novel porcine model of percutaneous aortic constriction by stent implantation.⁹

ncRNAs have also continued to attract attention as biomarkers with prognostic and diagnostic potential. A landmark study from Blanco-Dominguez et al. identified a novel miRNA with potential diagnostic value in acute myocarditis. The authors performed miRNA microarray analyses in sorted CD4+ T cells and Type 17 helper T (Th17) cells after inducing experimental autoimmune myocarditis or MI in mice and identified mmu-miR-72 as a differentially expressed miRNA. They further identified the human homologue hsa-miR-Chr8:96 and demonstrated its potential to distinguish patients with myocarditis from those with MI and healthy controls.¹⁰ Thus, miR-Chr8:96 has translational potential as a novel biomarker to diagnose myocarditis. miR-133a is a well-established, diagnostic circulating biomarker in patients with heart failure.¹¹ Escate et al. expanded on the diagnostic potential of this miRNA by demonstrating that elevated plasma levels of miR-133a predict the future occurrence of major adverse cardiovascular events (MACE) in patients with familial hypercholesterolaemia (FH).¹² This observation supports the potential utility of miR-133a in improving risk stratification and prognosis in high-risk patients. More broadly, an international consortium supporting collaboration and research on ncRNAs in cardiovascular disease (CardioRNA Cost Action CA17129) published a Position Paper on the pathophysiologic role of ncRNAs, and to provide recommendations to translate this into clinical practice.¹³

Other studies have progressed ncRNA candidates with therapeutic potential towards clinical translation.^{9,14,15} Kay et al. examined the potential of targeting ncRNAs to promote cell-based regenerative strategies for heart disease. Using an integrated approach, they identified CARMA (CARDiomyocyte Maturation-Associated lncRNA), a conserved lncRNA controlling cardiomyocyte differentiation and maturation in human embryonic stem cells. CARMA knockdown promoted cardiogenic commitment and cardiomyocyte differentiation in embryonic stem cells, and is therefore a novel target for improving human

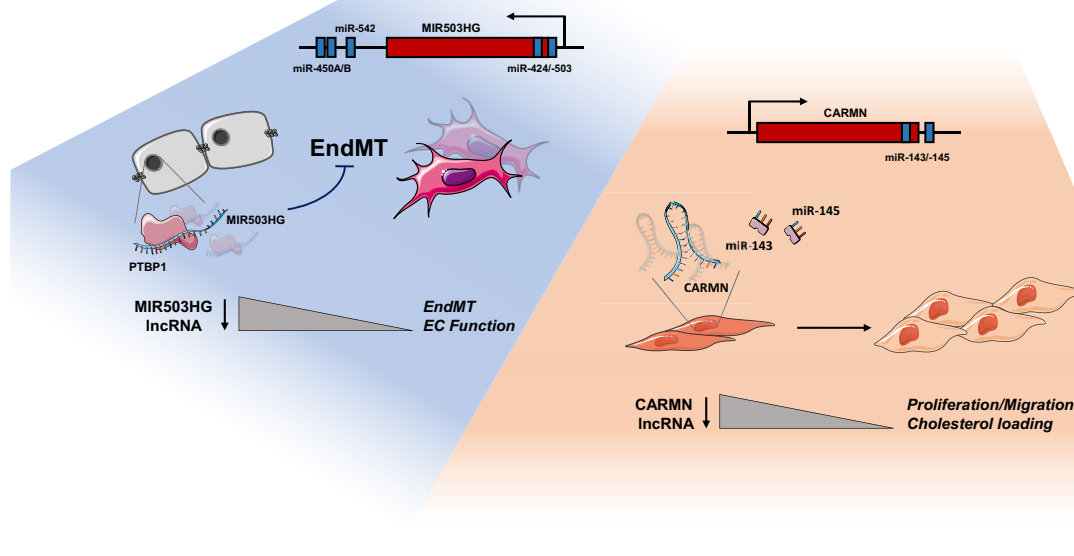


Figure 1 Novel insights into the role of ncRNAs. Several complex loci composed of lncRNA and miRNA clusters have been identified throughout the genome. Nonetheless, despite their genomic and often transcriptional overlap, they have been found to have distinct functional and regulatory targets. The X-linked lncRNA MIR503HG maintains EC identity by interacting with the RNA splicing regulatory protein PTBP1, with decreased expression leading to broad changes associated with EndMT. Importantly, these phenotypic changes seem to be independent of miR-424 and miR-503 expression, which overlap the lncRNA locus. Similarly, loss of the CARMN primes vSMCs into a pro-atherogenic proliferative state, while migration or dedifferentiation are regulated through the modulation of the overlapping miR-143 and miR-145.⁴

ESC-derived cardiomyocyte production in regenerative cardiovascular medicine.¹⁴ On the other hand, Modica *et al.* provided evidence for the effectiveness of a novel nanotechnology-based approach for delivering exogenous synthetic miR-133a. The authors demonstrated that intra-tracheal nebulization of miR-133a-nanoconstruct once-a-day on alternate days for 4 consecutive weeks protects against heart failure progression (improved cardiac function parameters and lower fibrosis) in a murine model. This improvement was associated with the restoration of physiological levels of miR-133a in cardiomyocytes without significant accumulation in other myocardial cells or organs.¹⁵

2.2 Single-cell approaches

Single-cell RNA-sequencing (scRNAseq) has emerged as a powerful tool to dissect transcriptional profiles of the complex cardiovascular system at single-cell resolution. scRNAseq has been insightful in our understanding of the earliest stages of cardiac development by identifying the epicardial progenitor field, which is anatomically and transcriptionally distinct from the currently known first and second heart fields.¹⁶ In the formed heart, scRNAseq and spatial transcriptomics were used to show that dysregulation of TBX5, the mutated gene causing septal and conduction defects in patients with Holt–Oram syndrome, leads to transcriptional consequences in specific cardiomyocyte subtypes.¹⁷ The study went on to show using cell-based analyses and mice that the stability of many gene regulatory networks, including those that have been shown to be relevant to congenital heart disease, are sensitive to TBX5 dosage.

At the level of the vasculature, the number of publications of atlas-type human or primate scRNAseq, or Assay for Transposase-Accessible

Chromatin (ATAC) datasets has steadily increased, which provides a valuable, yet often descriptive resource.^{18–21} scRNAseq has been used to identify transcriptional changes upon conditional cell type-specific genetic deletion, otherwise obscured in bulk tissue RNA-sequencing.²² As for immune cells in atherosclerosis, the detection of different subsets has culminated in a consensus on cell type markers,²³ yet to be achieved for the many varieties of vSMCs identified using scRNAseq in recent years, i.e. fibromyocytes, pro-inflammatory or modified vSMCs, SMC-derived intermediate cells.^{21,24–26} scRNAseq has also progressed our understanding of EC,^{27,28} with Rodor *et al.* identifying CD74 as potential target in PAH and showing its capacity to regulate barrier integrity.²⁸

At a cardiac level, the implementation of scRNAseq allowed the impact of heart failure on circulating immune cells to be determined.²⁹ Furthermore, it demonstrated an exacerbated inflamed transcriptome in circulating monocytes and a signature of T-cell activation in heart failure patients harbouring clonal haematopoiesis-driver mutations in DNA methyltransferase DNMT3A, thereby providing further insights into the potential effect of DNMT3A mutations in heart failure progression.³⁰ On the other hand, Hesse *et al.* have defined a high level of heterogeneity of epicardial stromal cells following MI, similar to cardiac fibroblast heterogeneity, with evidence of regenerative capacity and hypoxic signaling.³¹ Tombor *et al.* used scRNAseq of endothelial-lineage traced mice to change the dogma on EndMT in MI, showing this is a transient affair, often without a definite mesenchymal endstage.³²

Moving forward, cardiovascular scientists will benefit greatly from the generation of multi-omics reference atlases, including different layers of information on RNA, protein, spatial anatomy, interactome and cell ontology.^{33–35} Overall, scientific progress can be expedited by open-

access science and data sharing. Thus, the integration of available datasets for mesenchymal cells,³⁶ as previously carried out for immune cells in atherosclerosis,³⁷ and a web-based application by the Miller lab (plaqview.com),³⁸ pave the way for new, meaningful discoveries in cardiovascular biology.

3. Cardiovascular development

2021 witnessed progress in several important aspects of heart development with implications for our understanding of both congenital and acquired heart conditions. Genomic studies of congenital heart malformations now allow the analysis of variants within the context of gene networks. A good example of this is the recent genomic study on hypoplastic left heart syndrome (HLHS),³⁹ where whole-exome sequencing, coupled to nuclear transcriptomics and scRNAseq identified genetic heterogeneity in HLHS that converges to alter fundamental processes (e.g. autophagy, apoptosis, proliferation) in myogenesis.

Despite the relative ease in differentiating functional, if immature, cardiomyocytes from inducible pluripotent stem cell (iPSC), it has proven remarkably difficult to create organoids resembling the cellular and structural complexity of the vertebrate heart *in vitro*. However, Lewis-Israeli et al.⁴⁰ described a robust protocol for producing cardiac organoids from iPSC using a three-step Wnt signalling modulation strategy. These organoids develop a broad range of cardiac cell types, including those that are induced through interactions between distinct primary cardiac cell types, and develop cavities that superficially resemble the lumen of the chambers. Moreover, they are vascularized and display regular beating. Importantly, the transcriptome of the organoids more closely resembles foetal hearts than monolayer cardiomyocytes. This method is an important step on the path to developing a robust human-based *in vitro* model of the heart.

It is increasingly apparent that the majority of valve malformations and dysfunction arise from abnormal development, and yet the mechanisms of valve development are incompletely understood. The study by Fukui et al. focussed on the role of mechanical factors using zebrafish embryos.⁴¹ They identified a critical role for shear stress by showing that ectopic activation of wall shear stress, using agarose beads implanted into the atrium of the early zebrafish heart, resulted in the formation of valve-like structures that expressed the characteristic molecular signature of primitive valves, including the activation of NFATc and klf2a. Downstream of this, they ruled out a number of well-known mechanosensitive pathways, and instead identified adenosine tri-phosphate signalling as a mediator of Ca²⁺ oscillations that were essential for specifying valve cell identity. Overall, the convergence of large-scale genomic network analyses, scRNAseq and spatial transcriptomics and experimental developmental biology is coming close to explaining the mechanisms underlying heart malformations presenting at birth and in adulthood.

4. Vascular disease and repair

4.1 Mechanosensing

The Nobel Prize in Physiology or Medicine 2021 was awarded to David Julius from the University of California San Francisco and Ardem Patapoutian from The Scripps Research Institute La Jolla for explaining the molecular basis for sensing heat, cold, and mechanical force.⁴² Ardem Patapoutian identified PIEZO 1 and 2 as ion channels activated by mechanical force,⁴³ and they are central responders of arterial

responses to flow.⁴⁴ Recently, the protein kinase N2 (PKN2) has been shown to be activated by flow through the mechanosensitive ion channel PIEZO1 and mediate flow-induced endothelial NO synthase activation and vascular tone regulation⁴⁵ (Figure 2). As another important mechanosensor, the glycocalyx modulates the endothelial redox state in response to shear stress and could mediate an atheroprotective synergism between glycocalyx sialic acids and nuclear factor erythroid 2-related factor (NRF2) antioxidant signalling.⁴⁶ The regulation of NRF2 plays also a major role in the reduced EC viability and wound healing in response to cigarette smoke extracts under atherogenic low flow conditions.⁴⁷ The concept of disturbed flow as an initial stimulus for the development of atherosclerotic plaques has led to exciting new therapies to target mechanosensitive genes like *TWIST1*, *GATA4*, and bone morphogenic proteins using small interfering RNA (siRNA)-based technologies in an attempt to slow down the progression of atherosclerosis.^{48,49}

4.2 Atherosclerosis risk factors

The metabolic syndrome—in concert with inflammation—plays a central role in atherosclerosis. In particular, the causal role low-density lipoprotein (LDL) in atherosclerosis is indisputably supported by multiple lines of evidence such as epidemiological studies, Mendelian randomization and genetic analyses, as well as randomized clinical trials and animal model experimentation.

Traditional lipid-lowering drugs such as statins aim to reduce lipid uptake and/or cholesterol synthesis and are still widely used. However, the availability of genetic data and the identification of the genetic cause for rare diseases linked to dyslipidaemias has prompted spectacular advances in the identification of pharmacological targets for the treatment of dyslipidaemias (Figure 3). The most recent advances in lipid-lowering relate to the inhibition of proprotein convertase subtilisin kexin 9 (PCSK9), angiotensin-like 3 (ANGPTL3) and lipoprotein (a) (Lp(a)). Besides monoclonal antibodies, additional options to inhibit PCSK9 are emerging, including gene silencing with an siRNA or gene-editing employing the CRISPR/Cas system. Inclisiran, a siRNA conjugated with *N*-acetylgalactosamine residues ensuring hepatic selectivity, decreases PCSK9 production by promoting the degradation of its mRNA. This approach allows for twice-yearly dosing, with long-term lowering of LDL-cholesterol (LDL-C) (~50%), potentially enhancing patient compliance compared with other cholesterol-lowering drugs.^{50,51} Along the same line of RNA interference, Lp(a)-reducing drugs are being investigated in Phase 2–3 trials.⁵² At earlier stages of development are gene-editing technologies, which introduce permanent genomic changes to alter gene function. A single treatment with PCSK9 gene or base editors has been shown to confer durable LDL-C reduction in primates.⁵³ Evinacumab is a monoclonal antibody targeting ANGPTL3. It reduces significantly triglycerides by up to 80% in hypertriglyceridaemic subjects⁵⁴ and it is highly effective in reducing LDL-C levels in patients with homozygous FH carrying null *LDLR* mutations⁵⁵ providing a new pharmacological tool. In a recent study, membrane Type 1 matrix metalloproteinase (MT1-MMP), in addition to activating MMP-2, was shown to regulate LDL-receptor (LDLR) shedding, affecting circulating lipid concentrations and atherosclerosis.⁵⁶

The past year has further blurred the borders between traditional risk factors and the role of inflammation in atherosclerosis as their connections and interplay become more evident. Diabetes mellitus elevates cardiovascular risk, and hyperglycaemia contributes strongly to metabolic syndrome. Besides these known effects, Edgar et al. elucidated a pro-inflammatory and pro-atherogenic switch in macrophages from diabetic mice persisting even when cultured under normoglycaemic

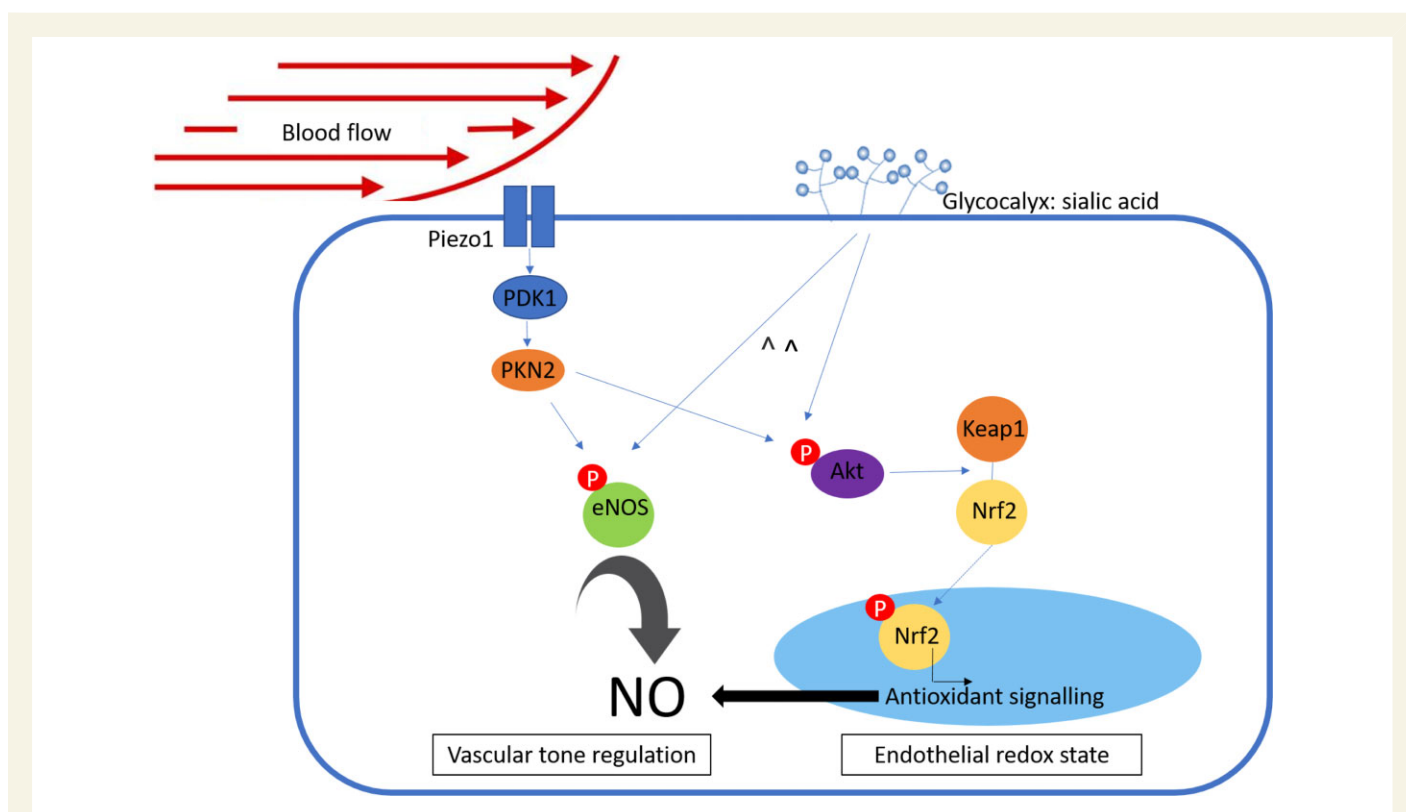


Figure 2 Recent findings on cardiovascular mechanosensing. Newly discovered flow-stimulated mechanosensitive signalling pathways. Flow-activated PIEZO1 was shown to activate the PKN2 via PKD1, resulting in phosphorylation of Akt and eNOS, with subsequent vascular tone regulation via NO.⁴⁵ The glycocalyx component sialic acid, was shown to activate NRF2 antioxidant signalling, via phosphorylation of AKT46, whereby modulating the endothelial redox state in response to shear stress. The pathways are likely to be interconnected as both result in phosphorylation of AKT and eNOS and as NRF2-induced antioxidant signalling is likely to affect NO bioavailability.

conditions.⁵⁷ This persevering effect of earlier hyperglycaemia may explain the relatively low degree of risk reduction upon glucose level normalization in diabetics. The inseparable connection between cholesterol and inflammation and atherosclerosis is further supported by a recent study that showed how sensing of cholesterol crystals by macrophages induces complement component C5aR1 signalling on mitochondrial membranes and results in interleukin (IL)-1 β production and sterile inflammation.⁵⁸ Hence, intracellular C5aR1 targeting may be used to normalize mitochondrial function and reduce IL-1 β release. This has translational relevance since inhibition of IL-1 β production through targeting the inflammasome has been identified as a target in cardiovascular disease previously. Another old acquaintance in cardiovascular disease therapy, rivaroxaban, a direct oral anticoagulant, not only targets factor Xa activity, but may also reduce inflammasome formation. In mice treated with rivaroxaban, macrophage autophagocytic activity increased significantly, which the authors were able to trace back to the Xa-PAR2 axis.⁵⁹

Recent studies show the complex intertwinement between traditional risk factors, vascular biology and immunology. Cardiovascular risk factors can affect haematopoiesis through defective angiogenesis in the bone marrow towards generation of inflammatory leukocytes, thereby creating a self-energizing circle of cardiovascular risk factors—defective angiogenesis—release of inflammatory cells—cardiovascular disease exacerbation.⁶⁰ Sakic *et al.* emphasized crosstalk between vSMCs and vascular inflammation by demonstrating that S100A4 induces vSMC change towards a pro-inflammatory phenotype to drive features of plaque instability.⁶¹ Together, these studies call for an integrated and

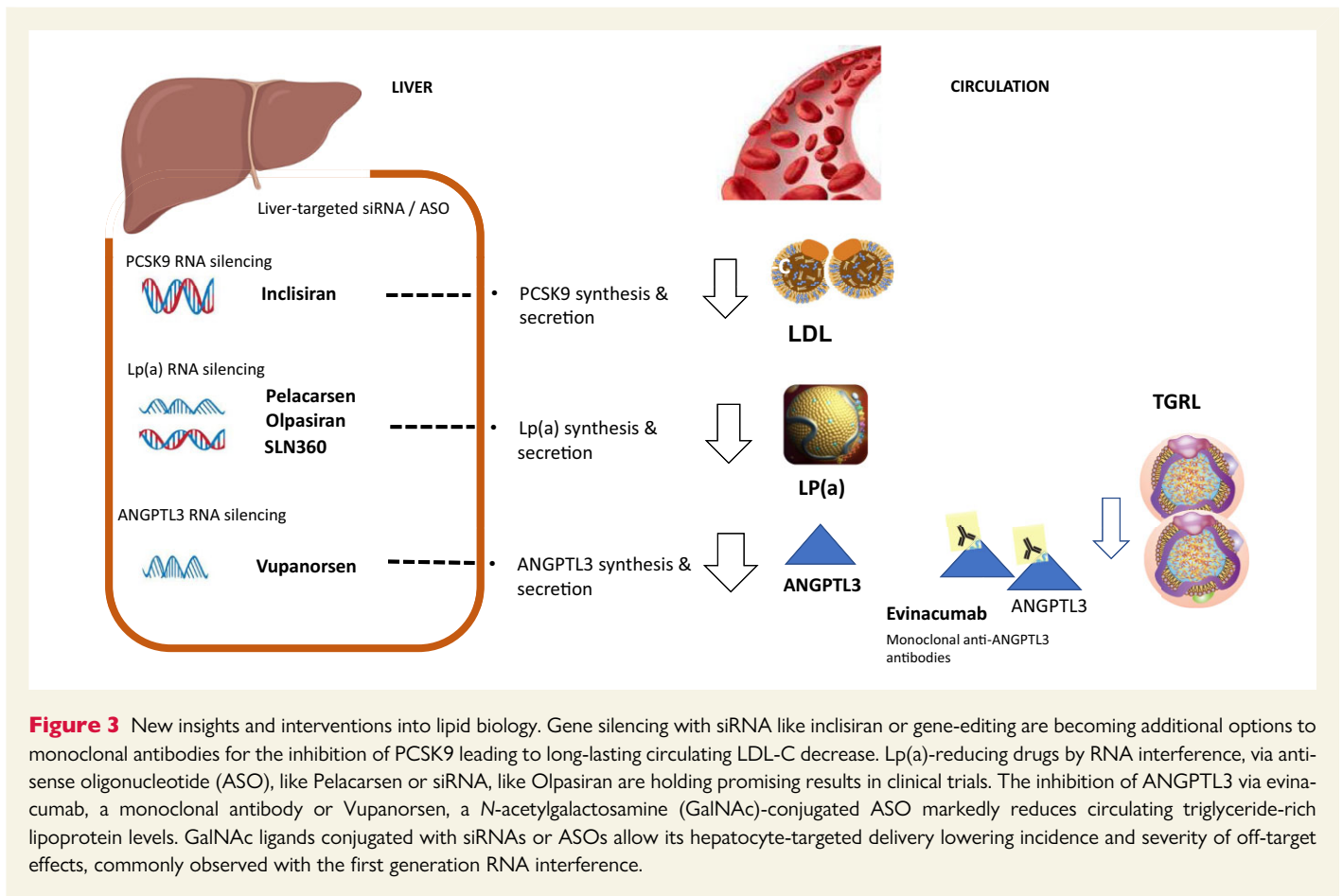
unprejudiced approach in atherosclerosis research to link traditional risk factors with novel molecular mechanisms.

4.3 Inflammation in atherosclerosis

The immune response is critical throughout the development of atherosclerotic lesions, during disease initiation, as a trigger for episodic plaque progression, and a contributor to thrombotic complications.⁶² A failure in the resolution of inflammation can prevent healing and repair of the vascular wall.^{62–64} This concept was advanced by Arnardottir *et al.* who found that lipid-specialized, pro-resolving mediators signalling through G-protein-coupled receptor-32, is critical for inflammatory resolution and atheroprotection.⁶⁴

The proposal that macrophage uptake mechanisms are decisive for the turning point that leads either to inflammation resolution or to chronic inflammation and plaque progression has received further support from analysis pro-resolving pathways⁶⁴ or phagocytic immune checkpoints in murine models.⁶⁵ Focussing on the CD47- signal-regulatory protein (SIRP) α immune checkpoint, loss of SIRP α in macrophages stimulated efferocytosis, attenuated oxidized LDL-induced inflammation and induced an M2 macrophage phenotype.⁶⁵ These findings may pave the way for novel interventions to promote inflammatory resolution through macrophage uptake mechanisms and phenotypic transitions to protect the vasculature.

Adaptive immune responses are critical regulators of atherosclerosis. On a systemic level, pro-inflammatory and cytotoxic T-lymphocytes prevail in atherosclerosis, as demonstrated by a preferential expansion



and function of CD28^{null} T-lymphocytes after *ex vivo* IL-7 and IL-15 stimulation of high-purity sorted CD4⁺ cells isolated from patients with acute coronary syndrome.⁶⁶ The local recruitment of regulatory T-lymphocytes (T_{reg}) is critical for the control of atherosclerotic lesion inflammation and is, in part, regulated by cellular metabolism.⁶⁷ As an approach to use T_{reg} recruitment as a therapeutic strategy to selectively target adaptive immune regulation in the atherosclerotic plaque, adoptive transfer of the fractalkine receptor CX3CR1 overexpressing T_{reg} was shown to increase their recruitment to atherosclerotic lesions and decreased atherosclerosis burden.⁶⁸

However, inhibition of some immune checkpoints can lead to enhanced atherosclerosis. This is exemplified by Poels *et al.* who found that short-term immune checkpoint inhibitors (ICIs) therapy aggravates T-cell-mediated plaque inflammation and drives plaque progression in mice.⁶⁹ Also, ICIs used to treat cancer, such as monoclonal antibodies targeting CTLA-4, PD-1, and PD-L1, have been associated with adverse cardiovascular events.⁷⁰ For example, Michel *et al.* discovered that anti-PD1 therapy in a mouse model of melanoma led to impaired left ventricular function and promoted myocardial infiltration with CD4⁺ and CD8⁺ T cells via a TNF-dependent mechanism.^{71,72} Therefore, the use of ICIs in the treatment of cancer provides exciting new opportunities for therapies but should be pursued with caution.

4.4 Haematopoiesis of indeterminate potential

Clonal haematopoiesis of indeterminate potential (CHIP) has recently emerged as an exciting topic in cardiovascular medicine and biology.

CHIP is defined as positive selection of specific somatic mutations in haematopoietic stem cells that provide a proliferative advantage and finally result in a clonal population carrying the mutation. Besides being associated with a 0.5–1% risk per year to develop leukaemia, CHIP is also associated with aging, smoking, obesity and Type 2 diabetes mellitus, chronic inflammation, infections, sleep deprivation, stress, hyperlipidaemia and atherosclerosis. Most mutations identified in CHIP affect the epigenetic regulators DNA (cytosine-5)-methyltransferase 3A (*DNMT3A*), tet methylcytosine dioxygenase 2 (*TET2*) and ASXL transcriptional regulator 1 (*ASXL1*) and the tyrosine kinase janus kinase 2 (*JAK2*) which result in a pro-inflammatory state that offers a possible explanation for the association of CHIP with a two-fold increase in risk to develop cardiovascular disease.^{73,74} Using mice that express the *JAK2*^{V617F} variant exclusively in macrophages, Fidler *et al.* reported increased proliferation of macrophages in atherosclerotic lesions and greater necrotic cores. These effects were ameliorated when Caspases 1 and 11, which are key components of the inflammasome or gasdermin D, which plays a major role in pyroptosis, were deleted. The authors also noted increased lesional expression of absent in melanoma 2 (*AIM2*) and found that atherosclerosis was reduced in mice deficient in *Aim2*. The authors concluded that enhanced proliferative stress caused by *JAK2*^{V617F} leads to DNA damage and to activation of the *AIM2* inflammasome resulting in IL-1 β activation, which then in turn starts a feed forward loop resulting in even more macrophage proliferation thereby aggravating atherosclerosis.⁷⁵

A new perspective to the field added Heyde *et al.* who recently showed by mathematical modelling and murine models that increased proliferation of haematopoietic stem cells occurs in individuals suffering

from atherosclerosis thereby increasing the risk to develop clonal haematopoiesis by the age of 70 years by 3.5-fold. Based on their findings the authors propose a vicious cycle in which atherosclerosis leads to clonal haematopoiesis, which in turn aggravates atherosclerosis.⁷⁶

5. Cardiac disease and repair

5.1 Extracellular vesicles and nanoparticles

2021 was another exciting year in the field of extracellular vesicle (EV) biology for regenerative medicine, including cardiac repair and regeneration (Figure 4). There was increasing interest in understanding the mechanism of EV-based intercellular communication within the myocardium during ventricular remodelling after acute MI. In terms of the role of EVs in cardiac fibrosis after MI, however, findings differ. For example, Li *et al.* showed that miR-30d is mainly secreted in EVs by cardiomyocytes and inhibits fibroblast proliferation by acting on integrin $\alpha 5$ via paracrine signalling.⁷⁷ Counterbalancing this view, Wang *et al.* evidenced, in a mouse model of MI, that EVs released by myocardial M2 macrophages exacerbate migration, proliferation and myofibroblastic transformation of cardiomyocytes.⁷⁸ By performing mechanistic studies in cocultured primary cardiomyocytes and M2 macrophages, the authors linked these effects to activation of miR-138-5p/RhoC signalling after delivery of the M2 macrophage-derived EVs containing circular RNA *circCUBE3a* into the cardiomyocytes.⁷⁸ These findings may offer an additional therapeutic target to optimize the endogenous mechanism of cardiac repair but suggest that EV function may depend on cell of origin.

There is great interest in the potential for EVs prepared from stem or progenitor cells to enhance cardiac repair. Increasing evidence suggests the mechanism may involve the resolution of inflammation. For example,

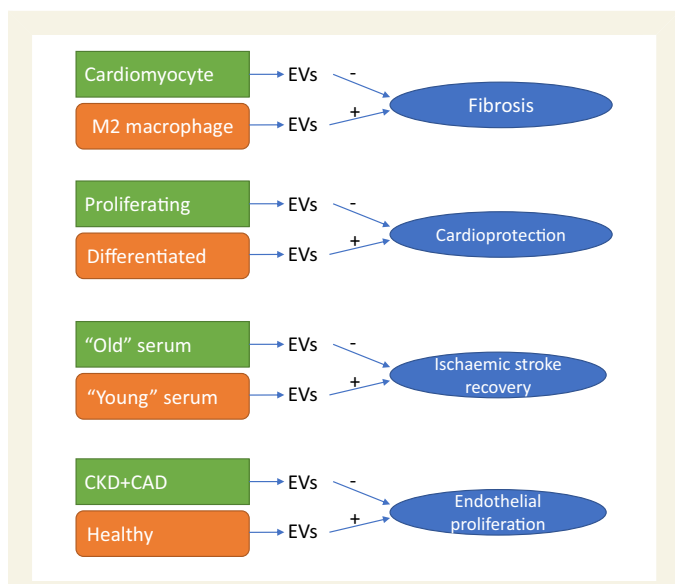


Figure 4 Source of EVs affects their function. Several thought-provoking studies published in 2021/22 demonstrated that the cardiovascular effects of EVs can depend upon their origin. For example, EVs originating from different cell types (cardiomyocytes vs. M2 macrophages), different cellular states (proliferating vs. differentiated), different ages (young vs. old serum) or different health states [chronic kidney disease and coronary artery disease (CKD + CAD) vs. healthy] can have opposite effects.

Correa *et al.* reported that EVs secreted from human iPSC-derived cardiovascular progenitor cells can trigger a pro-resolving immune response in preclinical murine models of either chronic or acute heart failure. Similar results were confirmed *in vitro* on human inflammatory cells, suggesting that this EV formulation can instruct the immune cell response towards a pro-resolving phenotype.⁷⁹ Patil *et al.* showed a similar pro-resolving effect of mesenchymal stem cell (MSC)-derived small EVs, which they attributed to the EVs both enhancing opsonization of dead cells and activating phagocytic signalling, thereby augmenting removal of apoptotic cells, resolution of inflammation, and improving cardiac recovery after injury.⁸⁰

In order to investigate a clinically feasible translational approach, Katsur *et al.* assessed whether cardioprotection could be achieved using a reproducible, clinical-grade preparation of small EVs obtained from the CTX0E03 human neural stem cell line. Systemic administration of small EVs from differentiating CTX0E03 reduced infarct size in mice and prevented *in vitro* cardiomyocyte mitochondrial permeability transition pore opening, which is responsible for cardiomyocyte death during reperfusion injury. These findings provide evidence for considering non-cardiovascular, yet stabilized, cell lines as additional candidate source of therapeutic EVs.⁸¹ Interestingly, however, EVs from proliferating CTX0E03 cells were not cardioprotective, which suggests that the status of cells of origin can impact their secreted EV activity.⁸¹ Further evidence of this is provided by a study showing that systemic administration of serum small EVs from young rats into aged ischaemic rats improved functional outcomes after ischaemic stroke, in contrast to small EVs from aged rats that worsened outcome.⁸² This provides further evidence that EV function is altered in disease, and further suggests that EV-miR-mediated vascular intercellular communication is altered in patients with chronic kidney disease and coronary artery disease.

A major goal in cardiac regenerative medicine is to identify novel methods to reinstate cardiomyocyte renewal. In such a scenario, EVs released from cardiac progenitors have been widely investigated, given the role of cardiac stromal cells such as the epicardium-derived progenitor cells play in cardiac muscle growth during embryonic development, and in heart regeneration in zebrafish and in neonatal mice. Villa del Campo *et al.* reported that epicardial EVs isolated from the secretome of both mouse and human progenitors enhanced the proliferative activity of neonatal murine cardiomyocytes *in vitro* and promoted cell cycle re-entry when injected into the injured area of infarcted neonatal hearts. These EVs also enhanced regeneration in cryoinjured engineered human myocardium constructs, as a novel model of human myocardial injury. Notably, the epicardial EV cargo was found enriched with specific miRNAs, including miR-30a, miR-100, miR-27a, and miR-30e, which recapitulated the EV regenerative influence on human stem cell-derived cardiomyocytes and cryoinjured cardiac constructs *in vitro*.⁸³

The relevance of the content of cardiovascular cell-derived EVs was highlighted by publications showing that miRNAs of the miR-106a-363 cluster,⁸⁴ periostin⁸⁵ and mitochondrial cargoes⁸⁶ can act as effectors of cardiac repair. While such encouraging evidence supports the exploitation of stem/progenitor cell-EVs as candidate therapeutics to promote adult cardiomyocyte proliferation, a general consensus has not been reached yet on their mechanism of action. In fact, Lima Correa *et al.* recently showed that EVs obtained from human iPSC-derived cardiac progenitor cells failed to trigger the generation of new cardiomyocytes in chronically infarcted hearts in mouse models. Despite this negative result, the authors confirmed that EVs from cardiac progenitor cells remained capable of significantly improving cardiac function by non-regenerative mechanisms.⁸⁷

These findings suggest that further analyses and accurate lineage tracing are required to better understand the regenerative potential of

cardiac EVs. At present, the rapid clearance of EVs from circulation is a limitation for their clinical application. During 2021, a number of studies aimed to overcome this barrier by constructing specific nanoparticles and genetically modifying cells to improve retention time of the cell-derived EVs. Thus, Wei *et al.* demonstrated that intravenously injected EV derived from modified mouse bone marrow MSC overexpressing CD47, a transmembrane protein known to elicit blockade of the mononuclear cell phagocytosis, have prolonged retention in the circulation and accumulate at greater levels in the ischaemic heart.⁸⁸

5.2 Cardiotoxicity and regeneration

A wide range of drugs, including but not limited to antineoplastic chemotherapeutic agents, can cause heart electrophysiology dysfunction, muscle damage and other cardiovascular pathologies. For example, anthracyclines such as doxorubicin (DOX) are a cornerstone for the treatment of many cancers, but their use is complicated by cardiotoxicity, especially left ventricular dysfunction.

An interesting 2021 paper reported that transcutaneous vagal nerve stimulation prevented DOX-induced cardiotoxicity in rats by rebalancing autonomic tone, ameliorating cardiac dysfunction and remodelling. It was hypothesized that the mechanism involved crosstalk between autonomic neuromodulation, innate immune cells such as macrophages and chemokines.⁸⁹ Indeed, there are multiple mechanisms responsible for anthracycline cardiotoxicity.^{70,90,91} Chan *et al.* found that two orally available MMP inhibitors ameliorated DOX cardiotoxicity by attenuating intracellular and extracellular matrix remodelling, suggesting that they may be a potential prophylactic strategy to prevent heart injury during chemotherapy.⁹⁰ Remote ischaemic preconditioning can ameliorate DOX-induced cardiotoxicity by preserving mitochondrial integrity⁹² and this is currently the subject of the RESILIENCE clinical trial.⁹³

Other recent studies (discussed in⁹⁴) have identified harmful effects of anticancer therapies on the ability of stem/progenitor cells to repair cardiac damage, through a reduction of stem cell viability and paracrine activity. Thus numerous animal and clinical studies have demonstrated that local or systemic administration of mesenchymal stem cells significantly improve cardiac function, through a reduction in inflammatory responses and myocardial fibrosis.⁹⁵ Antivirals can also induce cardiotoxicity, including the only FDA-approved treatment for hospitalized COVID-19 patients, remdesivir which can induce toxicity in human iPSC-derived cardiomyocytes through mitochondrial fragmentation, electrophysiological alterations and sarcomere disarray.⁹⁶

5.3 Cardiac arrhythmias

Several key insights into fibrillation and re-entrant arrhythmias were obtained in 2021 (Figure 5). Handa *et al.* revealed that the degree of gap junction coupling as well as the pattern of fibrosis influences mechanisms sustaining ventricular fibrillation.⁹⁷ Differentiating between these underlying mechanisms of maintenance of fibrillation may help to guide therapy. Re-entrant arrhythmias may also initiate in the absence of structural abnormalities, shown recently in a study on the spatiotemporal interaction between trigger and electrical substrate in the context of unexplained sudden cardiac arrest (SCA).⁹⁸ Analysis of explanted hearts and observations in survivors of unexplained SCA, identified key elements required for re-entry initiation including the occurrence of an early premature beat from an early repolarizing region of the ventricles, which may block against a steep repolarization time (RT) gradient to start re-entry. They also showed that detection of the origin of premature beats and their relation to RT gradients in patients is possible with non-invasive electrocardiographic imaging (ECGI) and may provide

targets for therapy. ECGI was also employed by Leong *et al.* in survivors of SCA to show that not only repolarization abnormalities, but also underlying conduction abnormalities play a role in the initiation of SCA.⁹⁹ A similar mechanistic reasoning extends to atrial arrhythmias.¹⁰⁰ Bringing these studies together highlights that any cause of steep excitability dispersion—whether resulting from local changes in gap junction coupling, fibrosis, local conduction slowing, or inherent repolarization duration heterogeneity—play a critical role in the initiation and maintenance of re-entry and fibrillation.

New tools are essential to obtain mechanistic insights and recent reports highlight how the field of atrial fibrillation research should transition from a translational approach to an integrative research approach¹⁰¹ and how personalized computer models may provide more individualized insights into disease and guide therapy.¹⁰² Application of novel therapeutic tools also brings new mechanistic insights. Non-invasive radiation therapy for cardiac arrhythmias was initially thought to induce fibrosis, similar to invasive catheter-based therapy.¹⁰³ However, Zhang *et al.* found that transmural fibrosis does not develop in the hearts of patients receiving radiation therapy within the timeframe of its ventricular tachycardia-reducing effects.¹⁰⁴ Interestingly, they showed that irradiating murine hearts results in a persistent supraphysiologic electrical phenotype, mediated by increases in sodium channel function and gap junction function. This functional restoration was confirmed by a shortening of QRS duration in patients receiving radiation therapy, highlighting that radiation-induced reprogramming of cardiac conduction is the potential mechanism beyond the initial success of radiation therapy for refractory ventricular tachycardia. This holds promise for extending the use of non-invasive radiation therapy to other applications, as for example recently demonstrated in heart failure with reduced ejection fraction.¹⁰⁵

6. Cardiovascular precision medicine and iPSC

Precision medicine aims to improve risk stratification and customize the management and therapy of patients based on their clinical and genetic characteristics, on datasets of large populations and the use of advanced technologies.¹⁰⁶ Genome-wide association studies (GWAS) have progressed through advances in genome-wide genotyping technology and large population and patient datasets to explore the role of common variants on phenotypic traits and disease susceptibility. According to the GWAS catalogue database, there are known to be 1329 polymorphism-cardiovascular trait associations. This growing catalogue of genome-wide and nominally significant variants has also opened the door to creating polygenic risk scores that could identify individuals at risk of developing specific cardiovascular diseases or sub-groups of patients with a more severe prognosis.¹⁰⁷ However, this approach must consider numerous confounding factors such as epigenetic and transcriptomic data that may correlate with genetic variants. Boix *et al.* undertook a *tour de force* to create EpiMap, a compendium comprising 10 000 epigenomic maps across 800 samples, which were used to define chromatin states, high-resolution enhancers, enhancer modules, upstream regulators, and downstream target genes.¹⁰⁸ This resource allowed the annotation of 30 000 genetic loci associated with 540 traits, predicting trait-relevant tissues, putative causal nucleotide variants in enriched tissue enhancers and candidate tissue-specific target genes for each of them. These different data integration layers could be essential for understanding the genetic architecture underlying the broad phenotypic traits encountered in common and complex cardiovascular

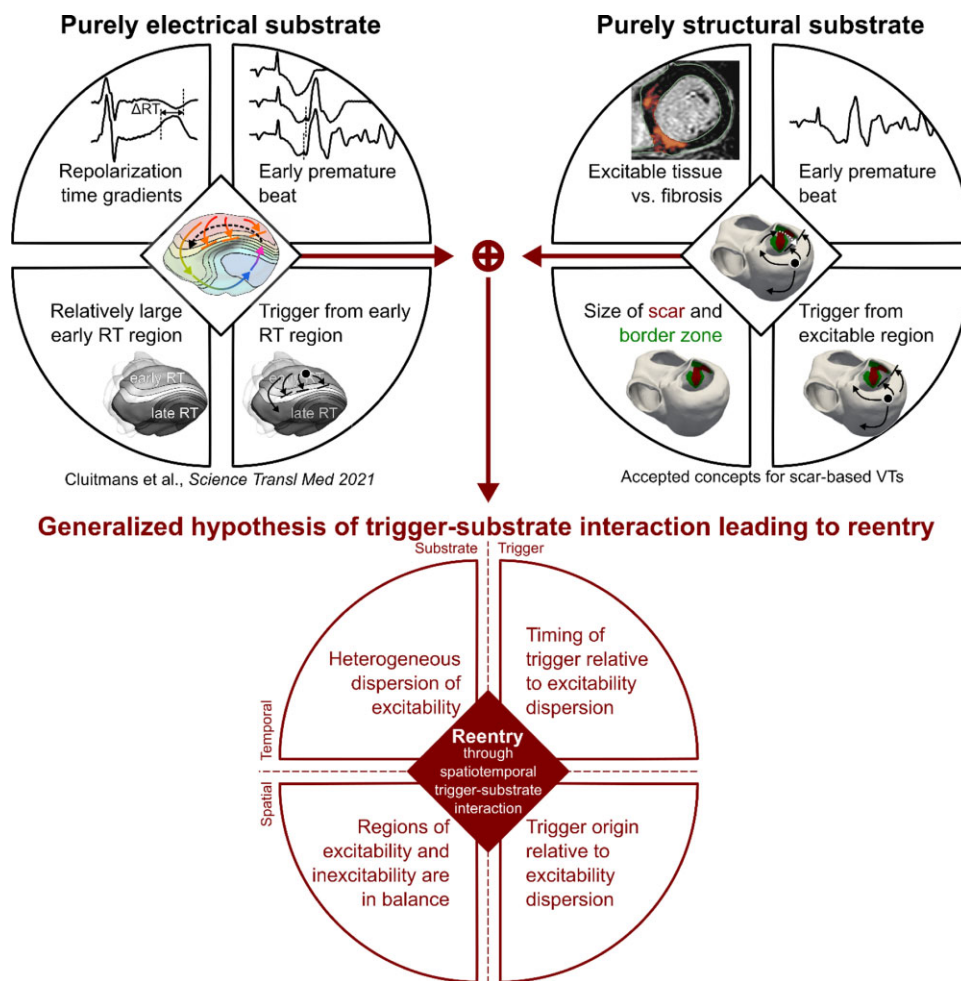


Figure 5 Novel mechanisms of arrhythmia. Recent publications (top left) and accepted concepts (top right) on the mechanisms leading to re-entry may be combined to arrive at a generalized theory of the spatiotemporal interaction between triggers and substrate leading to re-entry arrhythmias (bottom). The generalized hypothesis highlights that re-entry can only initiate when there is a local dispersion of excitability, with some tissue excitable whereas other tissue is (still, or always) refractory at the time when the trigger occurs. The trigger should originate from the excitable tissue, may block and travel around (relatively large) refractory tissue before it arrives at the previously excited tissue again.

diseases such as coronary artery disease. For instance, while ‘only’ 56 ‘unifactorial’ traits were enriched in the case of long-QT syndrome (LQTS), a total of 192 ‘multifactorial’ traits were enriched in an average of five different tissues, and in the case of coronary artery disease, 26 ‘polyfactorial’ traits were enriched in 14 tissues. The study by Boix *et al.* is at the same time a rich scientific resource, but also a lesson regarding the profound and magnificent complexity of the human genome and the causal basis of common diseases like coronary artery disease.

The GENMED consortium conducted a large GWAS study focused on dilated cardiomyopathy (DCM), enrolling 2719 cases and 4440 controls.¹⁰⁹ They identified and replicated two new DCM-associated loci on chromosome 3p25.1 and chromosome 22q11.23. *In silico* annotation and functional 4C-sequencing analyses on cardiomyocytes derived from iPSC-derived cardiomyocytes identified SLC6A6, a gene encoding a taurine, as the most likely DCM candidate at the 3p25.1 locus, and SMARCB1 as the candidate culprit gene at the 22q11.23 locus. The consortium also constructed a genetic risk score for DCM.

In another important study, exome sequencing data from 811 probands with tetralogy of Fallot (TOF) were used to identify rare

loss-of-function and other likely pathogenic variants in genes associated with congenital heart disease.¹¹⁰ The role of some likely pathogenic variants was confirmed and multiple loss-of-function variants provided support for 3 emerging congenital heart disease/TOF candidate genes: *KDR*, *IQGAP1*, and *GDF1*. Moreover, using composite genes in a STRING protein interaction enrichment analysis, a biologically relevant network was revealed, with vascular endothelial growth factor receptor 2 and NOTCH1 representing central nodes.

The use of iPSC technology for disease modelling and drug testing is increasingly used for cardiovascular precision medicine. Last year, for the first time, the combination of patient-specific iPSC-derived cardiomyocytes, genetics and genome editing unveiled the mechanisms of action of modifier genes in subsets of patients affected by LQTS.^{111,112} By comparing patient-specific iPSC-CMs derived from symptomatic and asymptomatic LQT1 carriers of the same mutation, it was shown that genetic variants of *MTMR4*, an upstream regulator of neural precursor cell expressed developmentally downregulated gene 4-like (NEDD4L), control potassium channel turnover, thus influencing the clinical manifestations of the disease. iPSC technology has also been used to gain insights into

the molecular mechanisms of atrial septum defect, a form of congenital heart disease, by implicating a mutation in *GATA4* that modifies *FGF16* induction.¹¹³

Pioneering work from Srivastava and collaborators developed a machine-learning approach to identify small molecules that broadly correct gene networks dysregulated in an iPSC model of aortic valve (AV) disease.¹¹⁴ Correction of the gene network by the most effective therapeutic candidate, XCT790, was sufficient to prevent and treat AV disease *in vivo* in a mouse model. This strategy, made possible by combining iPSC technology, network analytics and machine-learning, may represent an effective path to discovering new therapies.

7. COVID-19

7.1 Cardiovascular tropism and molecular markers

The aetiology of myocarditis caused by cardiotropic viruses has become a major topic of interest during the COVID-19 pandemic.^{115,116} A comparative study revealed that while myocardial injury occurred with a similar frequency in infection with influenza and SARS-CoV-2, the mortality was almost 4-fold higher in COVID-19 compared with influenza.¹¹⁷ Evidence of viral infection was seen mainly in endothelium and rarely in cardiomyocytes,¹¹⁸ however, evidence for stromal cells infection by SARS-CoV-2 has been found.¹¹⁹ Endothelial-dependent dilation in human arterioles is impaired for months after SARS-CoV-2 exposure, and could contribute to long-lasting symptoms of post-COVID-19 infection.¹²⁰ Consistently, Bräuninger *et al.* performed massive analysis of cDNA ends–RNAseq in myocardial tissue from fatal COVID-19 cases with and without cardiac infection to reveal potential SARS-CoV-2-related pro-inflammatory transcriptomic alterations in EC, while no differences were detected in immune cell infiltrations.¹¹⁸ Interestingly, the levels of several known cardiometabolic biomarkers are associated with COVID-19 severity and mortality, particularly myocyte-derived miR-133a and liver-derived miR-122.¹²¹ The potential for the use of cardiovascular RNA markers and artificial intelligence in the setting of COVID-19 has been reviewed in.¹²² In a study of 95 SARS-CoV-2-positive autopsy tissue, cardiac SARS-CoV-2 infection was shown to increase transcription of interferon pathways, originating predominantly from EC.¹¹⁸ The ESC has provided guidance for the diagnosis and management of cardiovascular disease during the COVID-19 pandemic^{123,124} and recommendations for future research.¹²⁵

7.2 Virus- and vaccine-induced thrombotic complications and COVID-19

Accumulating evidence suggests that patients suffering from COVID-19 have an increased risk to experience thrombotic events such as microthrombosis, venous thromboembolism, and ischaemic stroke (for a review see¹²⁶). Two recent studies have found microthrombi in the hearts of patients who succumbed to SARS-CoV-2 infections. Pellegrini *et al.* identified microthrombi as a cause of myocyte necrosis. Interestingly these microthrombi contained more fibrin and more of the complement components C5b-9 than thrombi isolated from the myocardium of patients of COVID-19-negative patients and coronary thrombi aspirated from COVID-19-negative and COVID-19-positive patients with ST-elevation MI.¹²⁷ Bois *et al.* found non-occlusive microthrombi in myocardial arterioles in 12 out of 15 patients who died from SARS-CoV-2 infections. However, no evidence of acute ischaemic injury of the heart was detected in this study.¹²⁸ When tissue factor (TF)-bearing

microvesicles isolated from the plasma of 100 patients with moderate and severe COVID-19 and from the plasma of 28 healthy subjects were studied, the authors found that TF-activity on such microvesicles, which is indicative of a procoagulatory state, was increased in patients suffering from COVID-19 and is significantly linked to disease severity and mortality.¹²⁹

Thrombotic complications have been reported in 1 per 100 000 adenoviral COVID-19 vaccinated irrespective of age, rising to 1 in 50 000 above 50 years vaccinated with ChAdOx1.¹³⁰ This is referred to as vaccine-induced immune thrombotic thrombocytopenia (VITT).^{130,131} Fibrinogen, Age, Platelet count, and the presence of Intracranial haemorrhage, and Cerebral venous sinus thrombosis (the FAPIC score) are significantly associated with mortality in cases of VITT.¹³² Increased levels of anti-PF4 antibodies post-vaccination unrelated to previous heparin exposure implicates an augmentation of the antibody response by unknown PF4 co-factors.¹³¹ The antigenic component with PF4 may be vaccine constituents but remains an unsolved critical question in VITT pathophysiology.¹³¹ The immune complexes transduce platelet activation through the Fcγ receptor IIA (FcγRIIA) resulting in thrombosis with concomitant thrombocytopenia accompanied by a fulminant immune activation.¹³³ Among novel therapeutic options for VITT, inhibitors of Bruton tyrosine kinase (Btk), which is used for B-cell malignancies, have been explored for their ability to block FcγRIIA for preventing the downstream platelet activation and aggregation. The Btk inhibitors ibrutinib and fenebrutinib prevented platelet aggregation induced by serum obtained from patients with VITT.¹³⁴ Additional possibly favourable effects of Btk inhibition in VITT are blocking of neutrophil-platelet complexes and reduced NET release,¹³⁵ which are part of the massive immune activation during VITT.¹³³

7.3 Cardiovascular drugs and COVID-19

In the beginning of the COVID-19 pandemic, the interactions with cardiovascular drugs were focused on ACE-inhibition and anti-thrombotic treatments¹³⁶ and more recently extended to lipid-modulating agents.¹³⁷ In the latter context, omega-3 fatty acids may provide beneficial cardiovascular effects through immunomodulation, anti-thrombosis and improved endothelial function.¹³⁸ Specific cytokine antibodies to dampen the inflammatory storm in COVID-19 exhibit anti-inflammatory strategies explored for cardiovascular prevention and have shown some success in improving survival and clinical outcomes.¹³⁹ The RECOVERY trial tested multiple different therapeutic approaches including anti-viral, immunomodulatory and anti-thrombotic treatments, in a multi-arm factorial design inspired by the International Study of Infarct Survival trials of the 1980s, and demonstrated benefit with tocilizumab and dexamethasone, but not hydroxychloroquine, convalescent plasma or other tested approaches.¹⁴⁰ In a separate study, anticoagulation with low-molecular-weight heparin may curtail viral persistence and reduce mortality.¹⁴¹

7.4 Perspectives

The substantial progress of basic cardiovascular science during the past year has revealed a plethora of novel therapeutic and diagnostic possibilities. Non-coding RNA, scRNAseq, and iPSC are examples of discovery tools to widen the understanding of cardiac and vascular pathophysiology. Through the integration cardiovascular risk factors, genetics, and biomarkers, the basic cardiovascular science field is expanding towards applications in precision medicine. The year was still marked by the COVID-19 pandemic and several important contributions have increased our knowledge of the cardiac and thrombotic effects of SARS-CoV-2, and the underlying pathways behind reported vaccinal

complications. Finally, the mechanistic insights from *in vitro* and *in vivo* basic science models have deepened our understanding of inflammation, CHIP, EVs, regeneration, and mechanosensing in cardiovascular disease.

Conflicts of interest: C.G.T. has received funding from Amgen, and personal fees from Vivalyfe, and is listed as an inventor on two heart failure patents.

References

- Davidson SM, Padro T, Bollini S, Vilahur G, Duncker DJ, Evans PC, Guzik T, Hoefer IE, Waltenberger J, Wojta J, Weber C. Progress in cardiac research: from rebooting cardiac regeneration to a complete cell atlas of the heart. *Cardiovasc Res* 2021;**117**:2161–2174.
- Evans P, Wojta J, Hoefer IE, Waltenberger J, Guzik T, Badimon L, Weber C. The year in basic vascular biology research: from mechanoreceptors and neutrophil extracellular traps to smartphone data and omics. *Cardiovasc Res* 2021;**117**:1814–1822.
- Monteiro JP, Rodor J, Caudrillier A, Scanlon JP, Spiroski AM, Dudnakova T, Pflüger-Müller B, Shmakova A, von Kriegsheim A, Deng L, Taylor RS, Wilson-Kanamorí JR, Chen SH, Stewart K, Thomson A, Mitić T, McClure JD, Iynkkel J, Hadoke PWF, Denby L, Bradshaw AC, Caruso P, Morrell NW, Kovacic JC, Ulitsky I, Henderson NC, Caporali A, Leisegang MS, Brandes RP, Baker AH. MIR503HG Loss promotes endothelial-to-mesenchymal transition in vascular disease. *Circ Res* 2021;**128**:1173–1190.
- Vacante F, Rodor J, Lalwani MK, Mahmoud AD, Bennett M, De Pace AL, Miller E, Van Kuijk K, de Bruijn J, Gijbels M, Williams TC, Clark MB, Scanlon JP, Doran AC, Montgomery R, Newby DE, Giacca M, O'Carroll D, Hadoke PWF, Denby L, Sluimer JC, Baker AH. CARMN Loss regulates smooth muscle cells and accelerates atherosclerosis in mice. *Circ Res* 2021;**128**:1258–1270.
- Gong X, Tian M, Cao N, Yang P, Xu Z, Zheng S, Liao Q, Chen C, Zeng C, Jose PA, Wang DZ, Jian Z, Xiao Y, Jiang DS, Wei X, Zhang B, Wang Y, Chen K, Wu G, Zeng C. Circular RNA circEysyt2 regulates vascular smooth muscle cell remodeling via splicing regulation. *J Clin Invest* 2021;**131**:e147031.
- Liu X, Guo JW, Lin XC, Tuo YH, Peng WL, He SY, Li ZQ, Ye YC, Yu J, Zhang FR, Ma MM, Shang JY, Lv XF, Zhou AD, Ouyang Y, Wang C, Pang RP, Sun JX, Ou JS, Zhou JG, Liang SJ. Macrophage NFATc3 prevents foam cell formation and atherosclerosis: evidence and mechanisms. *Eur Heart J* 2021;**42**:4847–4861.
- Schober A, Blay RM, Saboor Maleki S, Zahedi F, Winklmaier AE, Kakar MY, Baatsch IM, Zhu M, Geissler C, Fusco AE, Eberlein A, Li N, Megens RTA, Banafsche R, Kumbink J, Weber C, Nazari-Jahantigh M. MicroRNA-21 controls circadian regulation of apoptosis in atherosclerotic lesions. *Circulation* 2021;**144**:1059–1073.
- Ramanujam D, Schon AP, Beck C, Vaccarello P, Felician G, Dueck A, Esfandyari D, Meister G, Meitinger T, Schulz C, Engelhardt S. MicroRNA-21-dependent macrophage-to-fibroblast signaling determines the cardiac response to pressure overload. *Circulation* 2021;**143**:1513–1525.
- Hinkel R, Batkai S, Bahr A, Bozoglu T, Straub S, Borchert T, Viereck J, Howe A, Hornaschewitz N, Oberberger L, Jurisch V, Kozlik-Feldmann R, Freudenthal F, Ziegler T, Weber C, Sperandio M, Engelhardt S, Laugwitz KL, Moretti A, Klymiuk N, Thum T, Kupatt C. AntimiR-132 attenuates myocardial hypertrophy in an animal model of percutaneous aortic constriction. *J Am Coll Cardiol* 2021;**77**:2923–2935.
- Blanco-Dominguez R, Sanchez-Diaz R, de la Fuente H, Jimenez-Borreguero LJ, Matesanz-Marin A, Relano M, Jimenez-Alejandro R, Linillos-Pradillo B, Tsilingiri K, Martin-Mariscal ML, Alonso-Herranz L, Moreno G, Martin-Asenjo R, Garcia-Guimaraes MM, Bruno KA, Dauden E, Gonzalez-Alvaro I, Villar-Guimerans LM, Martinez-Leon A, Salvador-Garicano AM, Michelhaugh SA, Ibrahim NE, Januzzi JL, Kottwitz J, Illiceto S, Plebani M, Basso C, Baritussio A, Seguso M, Marcolongo R, Ricote M, Fairweather D, Bueno H, Fernandez-Friera L, Alfonso F, Caforio ALP, Pascual-Figal DA, Heidecker B, Luscher TF, Das S, Fuster V, Ibanez B, Sanchez-Madrid F, Martin P. A novel circulating microRNA for the detection of acute myocarditis. *N Engl J Med* 2021;**384**:2014–2027.
- Eitel I, Adams V, Dieterich P, Fuernau G, de Waha S, Desch S, Schuler G, Thiele H. Relation of circulating microRNA-133a concentrations with myocardial damage and clinical prognosis in ST-elevation myocardial infarction. *Am Heart J* 2012;**164**:706–714.
- Escate R, Padro T, Suades R, Camino S, Muniz O, Diaz-Diaz JL, Sionis A, Mata P, Badimon L. High miR-133a levels in the circulation anticipates presentation of clinical events in familial hypercholesterolemia patients. *Cardiovasc Res* 2021;**117**:109–122.
- Vanhaverbeke M, Attard R, Bartekova M, Ben-Aicha S, Brandenburger T, de Gonzalo-Calvo D, Emanuel C, Farrugia R, Grillari J, Hackl M, Kalocayova B, Martelli F, Scholz M, Wettinger SB, Devaux Y; EU-CardioRNA COST Action CA17129. Peripheral blood RNA biomarkers for cardiovascular disease from bench to bedside: A position paper from the EU-CardioRNA COST Action CA17129. *Cardiovasc Res*; doi: 10.1093/cvr/cvab327. Published online ahead of print 14 October 2021.
- Kay M, Soltani BM, Nemir M, Aghagholizadeh P, Pezzuto I, Chouvardas P, Ruberto F, Movahedi F, Ansari H, Baharvand H, Pedrazzini T. The conserved long noncoding RNA CARM1 regulates cardiomyocyte differentiation. *Cardiovasc Res*; doi: 10.1093/cvr/cvab281. Published online ahead of print 30 August 2021.
- Melica J, Di Mauro V, Barandalla-Sobrados M, Chavez SEP, Carullo P, Nemska S, Anselmo A, Condorelli G, Iafisco M, Miragoli M, Catalucci D. Nano-miR-133a replacement therapy blunts pressure overload-induced heart failure. *Circulation* 2021;**144**:1973–1976.
- Tyser RCV, Mahmammodov E, Nakanoh S, Vallier L, Scialdone A, Srinivas S. Single-cell transcriptomic characterization of a gastrulating human embryo. *Nature* 2021;**600**:285–289.
- Kathiraya IS, Rao KS, Iacono G, Devine WP, Blair AP, Hota SK, Lai MH, Garay BI, Thomas R, Gong HZ, Wesson LK, Goyal P, Sukonnik T, Hu KM, Akgun GA, Bernard LD, Akerberg BN, Gu F, Li K, Speir ML, Haeussler M, Pu WT, Stuart JM, Seidman CE, Seidman JG, Heyn H, Bruneau BG. Modeling human TBX5 haploinsufficiency predicts regulatory networks for congenital heart disease. *Dev Cell* 2021;**56**:292–309.e9.
- Hu Z, Liu W, Hua X, Chen X, Chang Y, Hu Y, Xu Z, Song J. Single-cell transcriptomic atlas of different human cardiac arteries identifies cell types associated with vascular physiology. *Arterioscler Thromb Vasc Biol* 2021;**41**:1408–1427.
- Ma S, Sun S, Li J, Fan Y, Qu J, Sun L, Wang S, Zhang Y, Yang S, Liu Z, Wu Z, Zhang S, Wang Q, Zheng A, Duo S, Yu Y, Belmonte JCI, Chan P, Zhou Q, Song M, Zhang W, Liu GH. Single-cell transcriptomic atlas of primate cardiopulmonary aging. *Cell Res* 2021;**31**:415–432.
- Dawson A, Li Y, Li Y, Ren P, Vasquez HG, Zhang C, Rebello KR, Ageedi W, Azares AR, Mattar AB, Sheppard MB, Lu HS, Coselli JS, Cassis LA, Daugherty A, Shen YH, LeMaire SA. Single-cell analysis of aneurysmal aortic tissue in patients with marfan syndrome reveals dysfunctional TGF-beta signaling. *Genes (Basel)* 2021;**13**:95.
- Wang Y, Gao H, Wang F, Ye Z, Mokry M, Turner AW, Ye J, Koplev S, Luo L, Alsaigh T, Adkar SS, Elishaev M, Gao X, Maegdefessel L, Björkregren JLM, Pasterkamp G, Miller CL, Ross EG, Leeper NJ. Dynamic changes in chromatin accessibility are associated with the atherogenic transition of vascular smooth muscle cells. *Cardiovasc Res*; doi: 10.1093/cvr/cvab347. Published online ahead of print 24 November 2021.
- van Kuijk K, Demandt JAF, Perales-Paton J, Theelen TL, Kuppe C, Marsch E, de Bruijn J, Jin H, Gijbels MJ, Matic L, Mees BME, Reutelingsperger CPM, Hedin U, Biessen EAL, Carmeliet P, Baker AH, Kramann RK, Schurgers JL, Saez-Rodriguez J, Sluimer JC. Deficiency of myeloid PHD proteins aggravates atherogenesis via macrophage apoptosis and paracrine fibrotic signaling. *Cardiovasc Res* 2022;**118**:1232–1246.
- Vallejo J, Cochain C, Zerneck A, Ley K. Heterogeneity of immune cells in human atherosclerosis revealed by scRNA-seq. *Cardiovasc Res* 2021;**117**:2537–2543.
- Wirka RC, Wagh D, Paik DT, Pjanic M, Nguyen T, Miller CL, Kundu R, Nagao M, Collier J, Koyano TK, Fong R, Woo YJ, Liu B, Montgomery SB, Wu JC, Zhu K, Chang R, Alamprese M, Tallquist MD, Kim JB, Quertermous T. Atheroprotective roles of smooth muscle cell phenotypic modulation and the TCF21 disease gene as revealed by single-cell analysis. *Nat Med* 2019;**25**:1280–1289.
- Alencar GF, Owsiany KM, Karnewar S, Sukhavi S, Mocci G, Nguyen AT, Williams CM, Shamsuzzaman S, Mokry M, Henderson CA, Haskins R, Baylis RA, Finn AV, McNamara CA, Zunder ER, Venkata V, Pasterkamp G, Björkregren J, Bekiranov S, Owens GK. Stem cell pluripotency genes Klf4 and Oct4 regulate complex SMC phenotypic changes critical in late-stage atherosclerotic lesion pathogenesis. *Circulation* 2020;**142**:2045–2059.
- Pan H, Xue C, Auerbach BJ, Fan J, Bashore AC, Cui J, Yang DY, Trignano SB, Liu W, Shi J, Huegbo CO, Bush EC, Worley J, Vlahos L, Laise P, Solomon RA, Connolly ES, Califano A, Sims PA, Zhang H, Li M, Reilly MP. Single-cell genomics reveals a novel cell state during smooth muscle cell phenotypic switching and potential therapeutic targets for atherosclerosis in mouse and human. *Circulation* 2020;**142**:2060–2075.
- Chen P-Y, Qin L, Li G, Wang Z, Dahlan JE, Malagon-Lopez J, Gujja S, Cilfone NA, Kauffman KJ, Sun L, Sun H, Zhang X, Aryal B, Canfran-Duque A, Liu R, Kusters P, Sehgal A, Jiao Y, Anderson DG, Gulcher J, Fernandez-Hernando C, Lutgens E, Schwartz MA, Pober JS, Chittenden TW, Tellides G, Simons M. Endothelial TGF-β signaling drives vascular inflammation and atherosclerosis. *Nat Metab* 2019;**1**:912–926.
- Rodor J, Chen SH, Scanlon JP, Monteiro JP, Caudrillier A, Sweta S, Stewart KR, Shmakova A, Dobie R, Henderson BEP, Stewart K, Hadoke PWF, Southwood M, Moore SD, Upton PD, Morrell NW, Li Z, Chan SY, Handen A, Lafyatis R, de Rooij L, Henderson NC, Carmeliet P, Spiroski AM, Brittan M, Baker AH. Single-cell RNA-seq profiling of mouse endothelial cells in response to pulmonary arterial hypertension. *Cardiovasc Res*; doi: 10.1093/cvr/cvab296. Published online ahead of print 16 September 2021.
- Abplanalp WT, John D, Cremer S, Assmus B, Dorsheimer L, Hoffmann J, Becker-Pergola G, Rieger MA, Zeiher AM, Vasa-Nicotera M, Dimmeler S. Single-cell RNA-sequencing reveals profound changes in circulating immune cells in patients with heart failure. *Cardiovasc Res* 2021;**117**:484–494.
- Abplanalp WT, Cremer S, John D, Hoffmann J, Schuhmacher B, Merten M, Rieger MA, Vasa-Nicotera M, Zeiher AM, Dimmeler S. Clonal hematopoiesis-driver DNMT3A mutations alter immune cells in heart failure. *Circ Res* 2021;**128**:216–228.
- Hesse J, Owenier C, Lautwein T, Zalfen R, Weber JF, Ding Z, Alter C, Lang A, Grandoch M, Gerdes N, Fischer JW, Klau GW, Dieterich C, Kohrer K, Schrader J. Single-cell transcriptomics defines heterogeneity of epicardial cells and fibroblasts within the infarcted murine heart. *Elife* 2021;**10**:e65921.
- Tombor LS, John D, Glaser SF, Luxan G, Forte E, Furtado M, Rosenthal N, Baumgarten N, Schulz MH, Wittig J, Rogg EM, Manavski Y, Fischer A, Muhly-Reinholz M, Klee K, Looso M, Selgnow C, Acker T, Bibli SI, Fleming I, Patrick R, Harvey RP, Abplanalp WT, Dimmeler S. Single cell sequencing reveals endothelial plasticity with transient mesenchymal activation after myocardial infarction. *Nat Commun* 2021;**12**:681.
- Borner K, Teichmann SA, Quardokus EM, Gee JC, Browne K, Osumi-Sutherland D, Herr BW, Bueckle A, Paul H, Haniffa M, Jardine L, Bernard A, Ding SL, Miller JA, Lin

- S, Halushka MK, Boppana A, Longacre TA, Hickey J, Lin Y, Valerius MT, He Y, Pryhuber G, Sun X, Jorgensen M, Radtke AJ, Wasserfall C, Ginty F, Ho J, Sunshine J, Beuschel RT, Brusko M, Lee S, Malhotra R, Jain S, Weber G. Anatomical structures, cell types and biomarkers of the human reference atlas. *Nat Cell Biol* 2021;**23**:1117–1128.
34. Osumi-Sutherland D, Xu C, Keays M, Levine AP, Kharchenko PV, Regev A, Lein E, Teichmann SA. Cell type ontologies of the human cell atlas. *Nat Cell Biol* 2021;**23**:1129–1135.
35. Maron BA, Wang RS, Shevtsov S, Drakos SG, Arons E, Wever-Pinzon O, Huggins GS, Samokhin AO, Oldham WM, Aguib Y, Yacoub MH, Rowin EJ, Maron BJ, Maron MS, Loscalzo J. Individualized interactomes for network-based precision medicine in hypertrophic cardiomyopathy with implications for other clinical pathophenotypes. *Nat Commun* 2021;**12**:873.
36. Conklin AC, Nishi H, Schlamp F, Ord T, Ounap K, Kaikkonen MU, Fisher EA, Romanoski CE. Meta-analysis of smooth muscle lineage transcriptomes in atherosclerosis and their relationships to in vitro models. *Immunometabolism* 2021;**3**:e210022.
37. Zernecke A, Winkels H, Cochain C, Williams JW, Wolf D, Soehnlein O, Robbins CS, Monaco C, Park I, McNamara CA, Binder CJ, Cybulsky MI, Scipione CA, Hedrick CC, Galkina EV, Kyaw T, Ghosheh Y, Dinh HQ, Ley K. Meta-analysis of leukocyte diversity in atherosclerotic mouse aortas. *Circ Res* 2020;**127**:402–426.
38. Ma WF, Hodonsky CJ, Turner AW, Wong D, Song Y, Mosquera JV, Ligay AV, Slenders L, Gancayco C, Pan H, Barrientos NB, Mai D, Alencar GF, Owsiany K, Owens GK, Reilly MP, Li M, Pasterkamp G, Mokry M, van der Laan SW, Khomtchouk BB, Miller CL. Enhanced single-cell RNA-seq workflow reveals coronary artery disease cellular crosstalk and candidate drug targets. *Atherosclerosis* 2022;**340**:12–22.
39. Krane M, Dreßen M, Santamaria G, My I, Schneider CM, Dorn T, Laue S, Mastantuono E, Berutti R, Rawat H, Gilsbach R, Schneider P, Lahm H, Schwarz S, Doppler SA, Paige S, Puluca N, Doll S, Neb I, Brade T, Zhang Z, Abou-Ajram C, Northoff B, Holdt LM, Sudhop S, Sahara M, Goedel A, Dendorfer A, Tjong FVY, Rijlaarsdam ME, Cleuziou J, Lang N, Kupatt C, Bezzina C, Lange R, Bowles NE, Mann M, Gelb BD, Crotti L, Hein L, Meitinger T, Wu S, Sinnecker D, Gruber PJ, Laugwitz KL, Moretti A. Sequential defects in cardiac lineage commitment and maturation cause hypoplastic left heart syndrome. *Circulation* 2021;**144**:1409–1428.
40. Lewis-Israeli YR, Wasserman AH, Gabalski MA, Volmert BD, Ming Y, Ball KA, Yang W, Zou J, Ni G, Pajares N, Chatzistavrou X, Li W, Zhou C, Aguirre A. Self-assembling human heart organoids for the modeling of cardiac development and congenital heart disease. *Nat Commun* 2021;**12**:5142.
41. Fukui H, Chow RW, Xie J, Foo YY, Yap CH, Minc N, Mochizuki N, Vermot J. Bioelectric signaling and the control of cardiac cell identity in response to mechanical forces. *Science* 2021;**374**:351–354.
42. Gracheva EO, Bagriantsev SN. Sensational channels. *Cell* 2021;**184**:6213–6216.
43. Kefauver JM, Ward AB, Patapoutian A. Discoveries in structure and physiology of mechanically activated ion channels. *Nature* 2020;**587**:567–576.
44. Li J, Hou B, Tumova S, Muraki K, Bruns A, Ludlow MJ, Sedo A, Hyman AJ, McKeown L, Young RS, Yuldasheva NY, Majeed Y, Wilson LA, Rode B, Bailey MA, Kim HR, Fu Z, Carter DA, Bilton J, Imrie H, Ajuh P, Dear T, Cubbon RM, Kearney MT, Prasad K, Evans PC, Ainscough JF X, Beech DJ. Piezot integration of vascular architecture with physiological force. *Nature* 2014;**515**:279–282.
45. Jin YJ, Chennupati R, Li R, Liang G, Wang S, Iring A, Graumann J, Wettschreck N, Offermanns S. Protein kinase N2 mediates flow-induced endothelial NOS activation and vascular tone regulation. *J Clin Invest* 2021;**131**:e145734.
46. Psefteli PM, Kitscha P, Vizcay G, Fleck R, Chapple SJ, Mann GE, Fowler M, Siow RC. Glycocalyx sialic acids regulate Nrf2-mediated signaling by fluid shear stress in human endothelial cells. *Redox Biol* 2021;**38**:101816.
47. Giebe S, Hofmann A, Brux M, Lowe F, Breheny D, Morawietz H, Brunssen C. Comparative study of the effects of cigarette smoke versus next generation tobacco and nicotine product extracts on endothelial function. *Redox Biol* 2021;**47**:102150.
48. Chiva-Blanch G, Evans PC. Scientists on the spot: a matter of blood flow. *Cardiovasc Res* 2021;**117**:e162–e163.
49. Evans PC, Fragiadaki M, Morris PD, Serbanovic-Canic J. Shear stress: the dark energy of atherosclerotic plaques. *Cardiovasc Res* 2021;**117**:1811–1813.
50. Ray KK, Landmesser U, Leiter LA, Kallend D, Dufour R, Karakas M, Hall T, Troquay RP, Turner T, Visseren FL, Wijngaard P, Wright RS, Kastelein JJ. Inclisiran in patients at high cardiovascular risk with elevated LDL cholesterol. *N Engl J Med* 2017;**376**:1430–1440.
51. Raal FJ, Kallend D, Ray KK, Turner T, Koenig W, Wright RS, Wijngaard PLJ, Curcio D, Jaros MJ, Leiter LA, Kastelein JJP; ORION-9 Investigators. Inclisiran for the treatment of heterozygous familial hypercholesterolemia. *N Engl J Med* 2020;**382**:1520–1530.
52. Tsimikas S, Karwatowska-Prokopczuk E, Gouni-Berthold I, Tardif JC, Baum SJ, Steinhagen-Thiessen E, Shapiro MD, Stroes ES, Moriarty PM, Nordestgaard BG, Xia S, Guerriero J, Viney NJ, O'Dea L, Witztum JL; AKCEA-APO(a)-LRx Study Investigators. Lipoprotein(a) reduction in persons with cardiovascular disease. *N Engl J Med* 2020;**382**:244–255.
53. Katzmann JL, Cupido AJ, Laufs U. Gene therapy targeting PCSK9. *Metabolites* 2022;**12**:70.
54. Ahmad Z, Pordy R, Rader DJ, Gaudet D, Ali S, Gonzaga-Jauregui C, Ponda MP, Shumel B, Banerjee P, Dunbar RL. Inhibition of angiotensin-like protein 3 with evinacumab in subjects with high and severe hypertriglyceridemia. *J Am Coll Cardiol* 2021;**78**:193–195.
55. Raal FJ, Rosenson RS, Reeskamp LF, Hovingh GK, Kastelein JJP, Rubba P, Ali S, Banerjee P, Chan KC, Gipe DA, Khilla N, Pordy R, Weinreich DM, Yancopoulos GD, Zhang Y, Gaudet D; ELIPSE HoFH Investigators. Evinacumab for homozygous familial hypercholesterolemia. *N Engl J Med* 2020;**383**:711–720.
56. Alabi A, Xia XD, Gu HM, Wang F, Deng SJ, Yang N, Adijiang A, Douglas DN, Kneteman NM, Xue Y, Chen L, Qin S, Wang G, Zhang DW. Membrane type 1 matrix metalloproteinase promotes LDL receptor shedding and accelerates the development of atherosclerosis. *Nat Commun* 2021;**12**:1889.
57. Edgar L, Akbar N, Braithwaite AT, Krausgruber T, Gallart-Ayala H, Bailey J, Corbin AL, Khojraty TE, Chai JT, Alkhalil M, Rendeiro AF, Ziberna K, Arya R, Cahill TJ, Bock C, Laurencikienė J, Crabtree J, Lemieux ME, Riksen NP, Netea MG, Wheelock CE, Channon KM, Rydén M, UdaloVA IA, Carnicer R, Choudhury RP. Hyperglycemia induces trained immunity in macrophages and their precursors and promotes atherosclerosis. *Circulation* 2021;**144**:961–982.
58. Niyonzima N, Rahman J, Kunz N, West EE, Freiwald T, Desai JV, Merle NS, Gidon A, Sporsheim B, Lionakis MS, Evensen K, Lindberg B, Skagen K, Skjelland M, Singh P, Haug M, Ruseva MM, Kolev M, Bibby J, Marshall O, O'Brien B, Deeks N, Afzali B, Clark RJ, Woodruff TM, Pryor M, Yang ZH, Remaley AT, Mollnes TE, Hewitt SM, Yan B, Kazemian M, Kiss MG, Binder CJ, Halvorsen B, Espevik T, Kemper C. Mitochondrial C5aR1 activity in macrophages controls IL-1 β production underlying sterile inflammation. *Sci Immunol* 2021;**6**:eabf2489.
59. Ito Y, Maejima Y, Nakagawa S, Shiheido-Watanabe Y, Tamura N, Sasano T, Rivaroxaban, a direct oral factor Xa inhibitor, attenuates atherosclerosis by alleviating factor Xa-PAR2-mediated autophagy suppression. *JACC Basic Transl Sci* 2021;**6**:964–980.
60. Rohde D, Vandoorne K, Lee I-H, Grune J, Zhang S, McAlpine CS, Schloss MJ, Nayar R, Courties G, Frodermann V, Wojtkiewicz G, Honold L, Chen Q, Schmidt S, Iwamoto Y, Sun Y, Cremer S, Hoyer FF, Iborra-Egea O, Muñoz-Guijosa C, Ji F, Zhou B, Adams RH, Wythe JD, Hidalgo J, Watanabe H, Jung Y, van der Laan AM, Piek JJ, Kfoury Y, Désogère PA, Vinegoni C, Dutta P, Sadreyev RI, Caravan P, Bayes-Genis A, Libby P, Scadden DT, Lin CP, Naxerova K, Swirski FK, Nahrendorf M. Bone marrow endothelial dysfunction promotes myeloid cell expansion in cardiovascular disease. *Nat Cardiovasc Res* 2022;**1**:28–44.
61. Sakic A, Chaabane C, Ambartsumian N, Klingelhöfer J, Lemeille S, Kwak BR, Grigorian M, Bochaton-Piallat ML. Neutralization of S100A4 induces stabilization of atherosclerotic plaques: role of smooth muscle cells. *Cardiovasc Res* 2022;**118**:141–155.
62. Libby P. Inflammation during the life cycle of the atherosclerotic plaque. *Cardiovasc Res* 2021;**117**:2525–2536.
63. Fredman G, MacNamara KC. Atherosclerosis is a major human killer and non-resolving inflammation is a prime suspect. *Cardiovasc Res* 2021;**117**:2563–2574.
64. Arnardtortir H, Thul S, Pawelzik SC, Karadimou G, Artiach G, Gallina AL, Mysdotter V, Carracedo M, Tarnawski L, Caravaca AS, Baumgartner R, Ketelhuth DF, Olofsson PS, Paulsson-Berne G, Hansson GK, Bäck M. The resolvin D1 receptor GPR32 transduces inflammation resolution and atheroprotection. *J Clin Invest* 2021;**131**:e142883.
65. Singla B, Lin HP, Ahn W, Xu J, Ma Q, Sghayer M, Dong K, Cherian-Shaw M, Zhou J, Huo Y, White J, Csanyi G. Loss of myeloid cell-specific SIRPalpha, but not CD47, attenuates inflammation and suppresses atherosclerosis. *Cardiovasc Res*; doi: 10.1093/cvr/cvab369. Published online ahead of print 23 December 2021.
66. Bullenkamp J, Mengoni V, Kaur S, Chhetri I, Dimou P, Astroulakis ZMJ, Kaski JC, Dumitriu IE. Interleukin-7 and interleukin-15 drive CD4+ CD28null T lymphocyte expansion and function in patients with acute coronary syndrome. *Cardiovasc Res* 2021;**117**:1935–1948.
67. Amersfoort J, Schaftenaar FH, Douna H, van Santbrink PJ, van Puijvelde GHM, Slutter B, Foks AC, Harms A, Moreno-Gordaliza E, Wang Y, Hankemeier T, Bot I, Chi H, Kuiper J. Diet-induced dyslipidemia induces metabolic and migratory adaptations in regulatory T cells. *Cardiovasc Res* 2021;**117**:1309–1324.
68. Bonacina F, Martini E, Svecla M, Nour J, Cremonesi M, Beretta G, Moregola A, Pellegatta F, Zampolieri V, Catapano AL, Kallikourdis M, Norata GD. Adoptive transfer of CX3CR1 transduced-T regulatory cells improves homing to the atherosclerotic plaques and dampens atherosclerosis progression. *Cardiovasc Res* 2021;**117**:2069–2082.
69. Poels K, van Leent MMT, Boutros C, Tissot H, Roy S, Meerwaldt AE, Toner YCA, Reiche ME, Kusters PJH, Malinova T, Huveneers S, Kaufman AE, Mani V, Fayad ZA, de Winther MPJ, Marabelle A, Mulder WJM, Robert C, Seijkens TTP, Lutgens E. Immune checkpoint inhibitor therapy aggravates T cell-driven plaque inflammation in atherosclerosis. *JACC CardioOncol* 2020;**2**:599–610.
70. Tocchetti CG, Ameri P, de Boer RA, D'Alessandra Y, Russo M, Sorriento D, Ciccarelli M, Kiss B, Bertrand L, Dawson D, Falcao-Pires I, Giacca M, Hamdani N, Linke WA, Mayr M, van der Velden J, Zaccagna S, Ghigo A, Hirsch E, Lyon AR, Gorbe A, Ferdinandy P, Madonna R, Heymans S, Thum T. Cardiac dysfunction in cancer patients: beyond direct cardiomyocyte damage of anticancer drugs. Novel cardio-oncology insights from the joint 2019. Meeting of the ESC working groups of myocardial function and cellular biology of the heart. *Cardiovasc Res* 2020;**116**:1820–1834.
71. Michel L, Helfrich I, Hendgen-Cotta UB, Minciu RI, Korste S, Mrotzek SM, Spomer A, Odersky A, Rischpler C, Herrmann K, Umutlu L, Coman C, Ahrends R, Sickmann A, Loffek S, Livingstone E, Ugurel S, Zimmer L, Gunzer M, Schandendorf D, Totzeck M, Rassaf T. Targeting early stages of cardiotoxicity from anti-PD1 immune checkpoint inhibitor therapy. *Eur Heart J* 2022;**43**:316–329.
72. Varricchi G, Galdiero MR, Tocchetti CG. Novel actors on the stage of cardiac dysfunction induced by anti-PD1 oncological treatments. *Eur Heart J* 2022;**43**:330–332.

73. Marnell CS, Bick A, Natarajan P. Clonal hematopoiesis of indeterminate potential (CHIP): linking somatic mutations, hematopoiesis, chronic inflammation and cardiovascular disease. *J Mol Cell Cardiol* 2021;**161**:98–105.
74. Wang Y, Sano S, Ogawa H, Horitani K, Evans MA, Yura Y, Miura-Yura E, Doviak H, Walsh K. Murine models of clonal hematopoiesis to assess mechanisms of cardiovascular disease. *Cardiovasc Res* 2022;**118**:1413–1432.
75. Fidler TP, Xue C, Yalcinkaya M, Hardaway B, Abramowicz S, Xiao T, Liu W, Thomas DG, Hajebrahimi MA, Pircher J, Silvestre-Roig C, Kotini AG, Luchsinger LL, Wei Y, Westertep M, Snoeck HW, Papapetrou EP, Schulz C, Massberg S, Soehnlein O, Ebert B, Levine RL, Reilly MP, Libby P, Wang N, Tall AR. The AIM2 inflammasome exacerbates atherosclerosis in clonal haematopoiesis. *Nature* 2021;**592**:296–301.
76. Heyde A, Rohde D, McAlpine CS, Zhang S, Hoyer FF, Gerold JM, Cheek D, Iwamoto Y, Schloss MJ, Vandoorne K, Iborra-Egea O, Munoz-Guijosa C, Bayes-Genis A, Reiter JG, Craig M, Swirski FK, Nahrendorf M, Nowak MA, Naxerova K. Increased stem cell proliferation in atherosclerosis accelerates clonal hematopoiesis. *Cell* 2021;**184**:1348–1361.e22.
77. Li J, Salvador AM, Li G, Valkov N, Ziegler O, Yeri A, Yang Xiao C, Meechooet B, Alsop E, Rodosthenous RS, Kundu P, Huan T, Levy D, Tigges J, Pico AR, Ghiran I, Silverman MG, Meng X, Kitchen R, Xu J, Van Keuren-Jensen K, Shah R, Xiao J, Das S. Mir-30d regulates cardiac remodeling by intracellular and paracrine signaling. *Circ Res* 2021;**128**:e1–e23.
78. Wang Y, Li C, Zhao R, Qiu Z, Shen C, Wang Z, Liu W, Zhang W, Ge J, Shi B. Circube3a from M2 macrophage-derived small extracellular vesicles mediates myocardial fibrosis after acute myocardial infarction. *Theranostics* 2021;**11**:6315–6333.
79. Correa B L, Harane N E, Gomez I, Rachid Hocine H, Vilar J, Desgres M, Bellamy V, Keirithana K, Guillas C, Perotto M, Pidial L, Alayrac P, Tran T, Tan S, Hamada T, Charron D, Brisson A, Renault NK, Al-Daccak R, Menasche P, Silvestre JS. Extracellular vesicles from human cardiovascular progenitors trigger a reparative immune response in infarcted hearts. *Cardiovasc Res* 2021;**117**:292–307.
80. Patil M, Saheera S, Dubey PK, Kahn-Krell A, Kumar Govindappa P, Singh S, Tousif S, Zhang Q, Lal H, Zhang J, Qin G, Krishnamurthy P. Novel mechanisms of exosome-mediated phagocytosis of dead cells in injured heart. *Circ Res* 2021;**129**:1006–1020.
81. Katsur M, He Z, Vinokur V, Corteling R, Yellon DM, Davidson SM. Exosomes from neuronal stem cells may protect the heart from ischaemia/reperfusion injury via JAK1/2 and gp130. *J Cell Mol Med* 2021;**25**:4455–4465.
82. Zhang H, Lin S, McElroy CL, Wang B, Jin D, Uteshev VV, Jin K. Circulating pro-inflammatory exosomes worsen stroke outcomes in aging. *Circ Res* 2021;**129**:e121–e140.
83. Villa Del Campo C, Liaw NY, Gunadasa-Rohling M, Matthaei M, Braga L, Kennedy T, Salinas G, Voigt N, Giacca M, Zimmermann WH, Riley PR. Regenerative potential of epicardium-derived extracellular vesicles mediated by conserved miRNA transfer. *Cardiovasc Res* 2022;**118**:597–611.
84. Jung JH, Ikeda G, Tada Y, von Bornstadt D, Santoso MR, Wahlquist C, Rhee S, Jeon YJ, Yu AC, O'Brien C G, Red-Horse K, Appel EA, Mercola M, Woo J, Yang PC. miR-106a-363 Cluster in extracellular vesicles promotes endogenous myocardial repair via Notch3 pathway in ischemic heart injury. *Basic Res Cardiol* 2021;**116**:19.
85. Balbi C, Milano G, Fertig TE, Lazzarini E, Bolis S, Taniyama Y, Sanada F, Di Silvestre D, Mauri P, Gherghiceanu M, Luscher TF, Barile L, Vassalli G. An exosomal-carried short periostin isoform induces cardiomyocyte proliferation. *Theranostics* 2021;**11**:5634–5649.
86. Ikeda G, Santoso MR, Tada Y, Li AM, Vaskova E, Jung JH, O'Brien C, Egan E, Ye J, Yang PC. Mitochondria-rich extracellular vesicles from autologous stem cell-derived cardiomyocytes restore energetics of ischemic myocardium. *J Am Coll Cardiol* 2021;**77**:1073–1088.
87. Lima Correa B, El Harane N, Desgres M, Perotto M, Alayrac P, Guillas C, Pidial L, Bellamy V, Baron E, Autret G, Kamaleswaran K, Pezzana C, Perier MC, Vilar J, Alberdi A, Brisson A, Renault N, Gnechchi M, Silvestre JS, Menasche P. Extracellular vesicles fail to trigger the generation of new cardiomyocytes in chronically infarcted hearts. *Theranostics* 2021;**11**:10114–10124.
88. Wei Z, Chen Z, Zhao Y, Fan F, Xiong W, Song S, Yin Y, Hu J, Yang K, Yang L, Xu B, Ge J. Mononuclear phagocyte system blockade using extracellular vesicles modified with CD47 on membrane surface for myocardial infarction reperfusion injury treatment. *Biomaterials* 2021;**275**:121000.
89. Lai Y, Zhou X, Guo F, Jin X, Meng G, Zhou L, Chen H, Liu Z, Yu L, Jiang H. Non-invasive transcutaneous vagal nerve stimulation improves myocardial performance in doxorubicin-induced cardiotoxicity. *Cardiovasc Res* 2021;**118**:1821–1834.
90. Chan BYH, Ruckzkowsky A, Cho WJ, Poirier M, Sergi C, Keschrumus V, Churko JM, Granzier H, Schulz R. MMP Inhibitors attenuate doxorubicin cardiotoxicity by preventing intracellular and extracellular matrix remodelling. *Cardiovasc Res* 2021;**117**:188–200.
91. Galan-Arriola C, Vilchez-Tschischke JP, Lobo M, Lopez GJ, de Molina-Iracheta A, Perez-Martinez C, Villena-Gutierrez R, Macias A, Diaz-Rengifo IA, Oliver E, Fuster V, Sanchez-Gonzalez J, Ibanez B. Coronary microcirculation damage in anthracycline cardiotoxicity. *Cardiovasc Res* 2022;**118**:531–541.
92. Galan-Arriola C, Villena-Gutierrez R, Higuero-Verdejo MI, Diaz-Rengifo IA, Pizarro G, Lopez GJ, Molina-Iracheta A, Perez-Martinez C, Garcia RD, Gonzalez-Calle D, Lobo M, Sanchez PL, Oliver E, Cordoba R, Fuster V, Sanchez-Gonzalez J, Ibanez B. Remote ischaemic preconditioning ameliorates anthracycline-induced cardiotoxicity and preserves mitochondrial integrity. *Cardiovasc Res* 2021;**117**:1132–1143.
93. Heusch G, Rassaf T. Protection from cardiotoxicity of cancer chemotherapy: a novel target for remote ischaemic conditioning? *Cardiovasc Res* 2021;**117**:985–986.
94. Smith AJ. Effects of cardiotoxins on cardiac stem and progenitor cell populations. *Front Cardiovasc Med* 2021;**8**:624028.
95. Lan HR, Xue Q, Liu YY, Jin KT, Fang XL, Shao H. The emerging therapeutic role of mesenchymal stem cells in anthracycline-induced cardiotoxicity. *Cell Tissue Res* 2021;**384**:1–12.
96. Kwok M, Lee C, Li HS, Deng R, Tsoi C, Ding Q, Tsang SY, Leung KT, Yan BP, Poon EN. Remdesivir induces persistent mitochondrial and structural damage in human induced pluripotent stem cell derived cardiomyocytes. *Cardiovasc Res*; doi: 10.1093/cvr/cvab311. Published online ahead of print 5 October 2021.
97. Handa BS, Li X, Baxan N, Roney CH, Shchendrygina A, Mansfield CA, Jabbour RJ, Pitcher DS, Chowdhury RA, Peters NS, Ng FS. Ventricular fibrillation mechanism and global fibrillatory organization are determined by gap junction coupling and fibrosis pattern. *Cardiovasc Res* 2021;**117**:1078–1090.
98. Cluitmans MJM, Bear LR, Nguyen UC, van Rees B, Stoks J, Ter Bekke RMA, Muhl C, Heijman J, Lau KD, Vigmond E, Bayer J, Belterman CNW, Abell E, Labrousse L, Rogier J, Bernus O, Haissaguerre M, Hassink RJ, Dubois R, Coronel R, Volders PGA. Noninvasive detection of spatiotemporal activation-repolarization interactions that prime idiopathic ventricular fibrillation. *Sci Transl Med* 2021;**13**:eab9317.
99. Leong KMW, Ng FS, Shun-Shin MJ, Koa-Wing M, Qureshi N, Whinnett ZI, Linton NF, Lefroy D, Francis DP, Harding SE, Davies DW, Peter NS, Lim PB, Behr E, Lambiase PD, Varnava A, Kanagaratnam P. Non-invasive detection of exercise-induced cardiac conduction abnormalities in sudden cardiac death survivors in the inherited cardiac conditions. *Europace* 2021;**23**:305–312.
100. Quintanilla JG, Shpun S, Jalife J, Figueiras-Rama D. Novel approaches to mechanism-based atrial fibrillation ablation. *Cardiovasc Res* 2021;**117**:1662–1681.
101. Schotten U. From translation to integration: how to approach the complexity of atrial fibrillation mechanisms. *Cardiovasc Res* 2021;**117**:e88–e90.
102. Heijman J, Sutanto H, Crijns H, Nattel S, Trayanova NA. Computational models of atrial fibrillation: achievements, challenges, and perspectives for improving clinical care. *Cardiovasc Res* 2021;**117**:1682–1699.
103. Cuculich PS, Schill MR, Kashani R, Mutic S, Lang A, Cooper D, Faddis M, Gleva M, Noheria A, Smith TW, Hallahan D, Rudy Y, Robinson CG. Noninvasive cardiac radiation for ablation of ventricular tachycardia. *N Engl J Med* 2017;**377**:2325–2336.
104. Zhang DM, Navara R, Yin T, Szymanski J, Goldsztejn J, Kenkel C, Lang A, Mpooy C, Lipovsky CE, Qiao Y, Hicks S, Li G, Moore KMS, Bergom C, Rogers BE, Robinson CG, Cuculich PS, Schwarz JK, Rentschler SL. Cardiac radiotherapy induces electrical conduction reprogramming in the absence of transmural fibrosis. *Nat Commun* 2021;**12**:5558.
105. Dusi V, Vitolo V, Frigerio L, Totaro R, Valentini A, Barcellona A, Mirandola A, Perego GB, Coccia M, Greco A, Ghio S, Valvo F, De Ferrari GM, Gnechchi M, Oltrona Visconti L, Rordorf R. First-in-man case of non-invasive proton radiotherapy for the treatment of refractory ventricular tachycardia in advanced heart failure. *Eur J Heart Fail* 2021;**23**:195–196.
106. Gnechchi M, Sala L, Schwartz PJ. Precision Medicine and cardiac channelopathies: when dreams meet reality. *Eur Heart J* 2021;**42**:1661–1675.
107. Barc J, Kovacic JC. From polygenic risk scores to integrative epigenomics: the Dawn of a new era for cardiovascular precision medicine. *Cardiovasc Res* 2021;**117**:e73–e75.
108. Boix CA, James BT, Park YP, Meuleman W, Kellis M. Regulatory genomic circuitry of human disease loci by integrative epigenomics. *Nature* 2021;**590**:300–307.
109. Garnier S, Harakalova M, Weiss S, Mokry M, Regitz-Zagrosek V, Hengstenberg C, Cappola TP, Isnard R, Arbustini E, Cook SA, van Setten J, Calis JJA, Hakonarson H, Morley MP, Stark K, Prasad SK, Li J, O'Regan DP, Grasso M, Muller-Nurasyid M, Meitinger T, Empana JP, Strauch K, Waldenberger M, Margulies KB, Seidman CE, Kararigas G, Meder B, Haas J, Boutouyrie P, Lacollet P, Jouven X, Erdmann J, Blankenberg S, Wichter T, Ruppert V, Tavazzi L, Dubourg O, Roizes G, Dorent R, de Groote P, Fauchier L, Trochu JN, Aupetit JF, Bilinska ZT, Germain M, Volker U, Hemerich D, Raji I, Bacq-Daian D, Proust C, Remior P, Gomez-Bueno M, Lehnert K, Maas R, Olaso R, Saripella GV, Felix SB, McGinn S, Duboscq-Bidot L, van Mil A, Besse C, Fontaine V, Blanche H, Ader F, Keating B, Curjol A, Boland A, Komajda M, Cambien F, Deleuze JF, Dorr M, Asselbergs FW, Villard E, Tregouet DA, Charron P, Consortium G. Genome-wide association analysis in dilated cardiomyopathy reveals two new players in systolic heart failure on chromosomes 3p25.1 and 22q11.23. *Eur Heart J* 2021;**42**:2000–2011.
110. Reuter MS, Chaturvedi RR, Jobling RK, Pellicchia G, Hamdan O, Sung WWL, Nalpathamkalam T, Attaluri P, Silversides CK, Wald RM, Marshall CR, Williams SG, Keavney BD, Thiruvahindrapuram B, Scherer SW, Bassett AS. Clinical genetic risk variants inform a functional protein interaction network for tetralogy of fallot. *Circ Genom Precis Med* 2021;**14**:e003410.
111. Lee YK, Sala L, Mura M, Rocchetti M, Pedrazzini M, Ran XR, Mak TSH, Crotti L, Sham PC, Torre E, Zaza A, Schwartz PJ, Tse HF, Gnechchi M. MTMR4 SNVs modulate ion channel degradation and clinical severity in congenital long QT syndrome: insights in the mechanism of action of protective modifier genes. *Cardiovasc Res* 2021;**117**:767–779.

112. Ronchi C, Bernardi J, Mura M, Stefanello M, Badone B, Rocchetti M, Crotti L, Brink P, Schwartz PJ, Gnecci M, Zaza A. NOS1AP Polymorphisms reduce NOS1 activity and interact with prolonged repolarization in arrhythmogenesis. *Cardiovasc Res* 2021;**117**:472–483.
113. Ye L, Yu Y, Zhao ZA, Zhao D, Ni X, Wang Y, Fang X, Yu M, Wang Y, Tang JM, Chen Y, Shen Z, Lei W, Hu S. Patient-specific iPSC-derived cardiomyocytes reveal abnormal regulation of FGF16 in a familial atrial septal defect. *Cardiovasc Res* 2021;**118**:859–871.
114. Theodoris CV, Zhou P, Liu L, Zhang Y, Nishino T, Huang Y, Kostina A, Ranade SS, Gifford CA, Uspenskiy V, Malashicheva A, Ding S, Srivastava D. Network-based screen in iPSC-derived cells reveals therapeutic candidate for heart valve disease. *Science* 2021;**371**:eabd0724.
115. Schultheiss HP, Baumeier C, Pietsch H, Bock CT, Poller W, Escher F. Cardiovascular consequences of viral infections: from COVID to other viral diseases. *Cardiovasc Res* 2021;**117**:2610–2623.
116. Raman B, Bluemke DA, Lüscher TF, Neubauer S. Long COVID: post-acute sequelae of COVID-19 with a cardiovascular focus. *Eur Heart J* 2022;**43**:1157–1172.
117. Biasco L, Klersy C, Beretta GS, Valgimigli M, Valotta A, Gabutti L, Bruna RD, Pagnamenta A, Tersalvi G, Ruinelli L, Artero A, Senatore G, Jüni P, Pedrazzini GB. Comparative frequency and prognostic impact of myocardial injury in hospitalized patients with COVID-19 and influenza. *Eur Heart J Open* 2021;**1**:oeb025.
118. Bräuninger H, Stoffers B, Fitzek ADE, Meißner K, Aleshcheva G, Schweizer M, Weimann J, Rotter B, Warnke S, Edler C, Braun F, Roedel K, Scherschel K, Escher F, Kluge S, Huber TB, Ondruschka B, Schultheiss HP, Kirchhof P, Blankenberg S, Püschel K, Wvstermann D, Lindner D. Cardiac SARS-CoV-2 infection is associated with pro-inflammatory transcriptomic alterations within the heart. *Cardiovasc Res* 2022;**118**:542–555.
119. Amendola A, Garoffolo G, Songia P, Nardacci R, Ferrari S, Bernava G, Canzano P, Myasoedova V, Colavita F, Castilletti C, Sberna G, Capobianchi MR, Piacentini M, Agrifoglio M, Colombo GI, Poggio P, Pesce M. Human cardiosphere-derived stromal cells exposed to SARS-CoV-2 evolve into hyper-inflammatory/pro-fibrotic phenotype and produce infective viral particles depending on the levels of ACE2 receptor expression. *Cardiovasc Res* 2021;**117**:1557–1566.
120. Nishijima Y, Hader SN, Hanson AJ, Zhang DX, Sparapani R, Gutterman DD, Beyer AM. Prolonged endothelial-dysfunction in human arterioles following infection with SARS-CoV-2. *Cardiovasc Res* 2022;**118**:18–19.
121. Gutmann C, Khamina K, Theofilatos K, Diendorfer AB, Burnap SA, Nabeebaccus A, Fish M, McPhail MJW, O'Gallagher K, Schmidt LE, Cassel C, Auzinger G, Napoli S, Mujib SF, Trovato F, Sanderson B, Merrick B, Roy R, Edgeworth JD, Shah AM, Hayday AC, Traby L, Hackl M, Eichinger S, Shankar-Hari M, Mayr M. Association of cardiometabolic microRNAs with COVID-19 severity and mortality. *Cardiovasc Res* 2022;**118**:461–474.
122. Badimon L, Robinson EL, Jusic A, Carpusca I, DeWindt LJ, Emanuelli C, Ferdinandy P, Gu W, Gyongyosi M, Hackl M, Karadzovic-Hadziabdic K, Lustrek M, Martelli F, Nham E, Potocnjak I, Satagopam V, Schneider R, Thum T, Devaux Y. Cardiovascular RNA markers and artificial intelligence may improve COVID-19 outcome: a position paper from the EU-CardioRNA COST Action CA17129. *Cardiovasc Res* 2021;**117**:1823–1840.
123. Task Force for the management of COVID-19 of the European Society of Cardiology; Baigent C, Windecker S, Andreini D, Arbelo E, Barbato E, Bartorelli AL, Baumbach A, Behr ER, Berti S, Bueno H, Capodanno D, Cappato R, Chieffo A, Collet JP, Cuisset T, de Simone G, Delgado V, Dendale P, Dudek D, Edvardsen T, Elvan A, González-Juanatey JR, Gori M, Grobbee D, Guzik TJ, Halvorsen S, Haude M, Heidbuchel H, Hindricks G, Ibanez B, Karam N, Katus H, Klok FA, Konstantinides SV, Landmesser U, Leclercq C, Leonardi S, Lettino M, Marenzi G, Mauri J, Metra M, Morici N, Mueller C, Petronio AS, Polovina MM, Potpara T, Praz F, Prendergast B, Prescott E, Price S, Pruszczyk P, Rodríguez-Leor O, Roffi M, Romaguera R, Rosenkranz S, Sarkozy A, Scherrenberg M, Seferovic P, Senni M, Spera FR, Stefanini G, Thiele H, Tomasoni D, Torracca L, Touyz RM, Wilde AA, Williams B. European Society of Cardiology guidance for the diagnosis and management of cardiovascular disease during the COVID-19 pandemic: part 1-epidemiology, pathophysiology, and diagnosis. *Cardiovasc Res* 2022;**118**:1385–1412.
124. Task Force for the management of COVID-19 of the European Society of Cardiology. ESC Guidance for the diagnosis and management of cardiovascular disease during the COVID-19 pandemic: part 2-care pathways, treatment, and follow-up. *Cardiovasc Res* 2021;**118**:1618–1666.
125. Pesce M, Agostoni P, Botker HE, Brundel B, Davidson SM, De Caterina R, Ferdinandy P, Giraio H, Gyongyosi M, Hulot JS, Lecour S, Perrino C, Schulz R, Sluijter JP, Steffens S, Tancevski I, Gollmann-Tepekoylu C, Tschope C, van Linthout S, Madonna R. COVID-19-related cardiac complications from clinical evidences to basic mechanisms: opinion paper of the ESC Working Group on Cellular Biology of the Heart. *Cardiovasc Res* 2021;**117**:2148–2160.
126. McFadyen JD, Stevens H, Peter K. The emerging threat of (micro)thrombosis in COVID-19 and its therapeutic implications. *Circ Res* 2020;**127**:571–587.
127. Pellegrini D, Kawakami R, Guagliumi G, Sakamoto A, Kawai K, Gianatti A, Nasr A, Kutys R, Guo L, Cornelissen A, Faggi L, Mori M, Sato Y, Pescetelli I, Brivio M, Romero M, Virmani R, Finn AV. Microthrombi as a major cause of cardiac injury in COVID-19: a pathologic study. *Circulation* 2021;**143**:1031–1042.
128. Bois MC, Boire NA, Layman AJ, Aubry MC, Alexander MP, Roden AC, Hagen CE, Quinton RA, Larsen C, Erben Y, Majumdar R, Jenkins SM, Kipp BR, Lin PT, Maleszewski JJ. COVID-19-associated nonocclusive fibrin microthrombi in the heart. *Circulation* 2021;**143**:230–243.
129. Rosell A, Havervall S, von Meijenfeldt F, Hisada Y, Aguilera K, Grover SP, Lisman T, Mackman N, Thålin C. Patients with COVID-19 have elevated levels of circulating extracellular vesicle tissue factor activity that is associated with severity and mortality-brief report. *Arterioscler Thromb Vasc Biol* 2021;**41**:878–882.
130. Marchandot B, Curtiaud A, Trimaille A, Sattler L, Grunebaum L, Morel O. Vaccine-induced immune thrombotic thrombocytopenia: current evidence, potential mechanisms, clinical implications, and future directions. *Eur Heart J Open* 2021;**1**:oeb014.
131. McFadyen JD, Peter K. The known knowns and known unknowns of vaccine-induced thrombotic thrombocytopenia. *Cardiovasc Res* 2021;**117**:e147–e150.
132. Hwang J, Park SH, Lee SW, Lee SB, Lee MH, Jeong GH, Kim MS, Kim JY, Koyanagi A, Jacob L, Jung SY, Song J, Yon DK, Shin JJ, Smith L. Predictors of mortality in thrombotic thrombocytopenia after adenoviral COVID-19 vaccination: the FAPIC score. *Eur Heart J* 2021;**42**:4053–4063.
133. Holm S, Kared H, Michelsen AE, Kong XY, Dahl TB, Schultz NH, Nyman TA, Fladeby C, Seljeflot I, Ueland T, Stensland M, Mjåaland S, Goll GL, Nissen-Meyer LS, Aukrust P, Skagen K, Gregersen I, Skjelland M, Holme PA, Munthe LA, Halvorsen B. Immune complexes, innate immunity, and NETosis in ChAdOx1 vaccine-induced thrombocytopenia. *Eur Heart J* 2021;**42**:4064–4072.
134. Weber C, von Hundelshausen P, Siess W. VITT After ChAdOx1 nCoV-19 vaccination. *N Engl J Med* 2021;**385**:2203–2204.
135. von Hundelshausen P, Lorenz R, Siess W, Weber C. Vaccine-induced immune thrombotic thrombocytopenia (VITT): targeting pathomechanisms with bruton tyrosine kinase inhibitors. *Thromb Haemost* 2021;**121**:1395–1399.
136. Cenko E, Badimon L, Bugiardini R, Claeys MJ, De Luca G, de Wit C, Derumeaux G, Dorobantu M, Duncker DJ, Eringa EC, Gorog DA, Hassager C, Heinzl FR, Huber K, Manfrini O, Milicic D, Oikonomou E, Padro T, Trifunovic-Zamaklar D, Vasiljevic-Pokrajac Z, Vavlukis M, Vilahur G, Tousoulis D. Cardiovascular disease and COVID-19: a consensus paper from the ESC Working Group on Coronary Pathophysiology & Microcirculation, ESC Working Group on Thrombosis and the Association for Acute Cardiovascular Care (ACVC), in collaboration with the European Heart Rhythm Association (EHRA). *Cardiovasc Res* 2021;**117**:2705–2729.
137. Talasz AH, Sadeghipour P, Aghakouchakzadeh M, Dreyfus I, Kakavand H, Ariannejad H, Gupta A, Madhavan MV, Van Tassel BW, Jimenez D, Monreal M, Vaduganathan M, Fanikos J, Dixon DL, Piazza G, Parikh SA, Bhatt DL, Lip GYH, Stone GW, Krumholz HM, Libby P, Goldhaber SZ, Bickdeli B. Investigating lipid-modulating agents for prevention or treatment of COVID-19 JACC state-of-the-art review. *J Am Coll Cardiol* 2021;**78**:1635–1654.
138. Zhu YF, Wen L, Wang S, Zhang K, Cui Y, Zhang C, Feng L, Yu F, Chen YQ, Wang RX, Ma X. Omega-3 fatty acids improve flow-induced vasodilation by enhancing TRPV4 in arteries from diet-induced obese mice. *Cardiovasc Res* 2021;**117**:2450–2458.
139. Ridker PM. Targeting cytokine storm in COVID-19: what have we learned? *Eur Heart J Open* 2021;**1**:oeb005.
140. Pessoa-Amorim G, Mafham MM. The RECOVERY trial: cardiovascular implications of a large, simple randomized trial in COVID-19. *Cardiovasc Res* 2021;**117**:e110–e113.
141. Pereyra D, Heber S, Schrottmaier WC, Santol J, Pirabe A, Schmuckenschlager A, Kammerer K, Ammon D, Sorz T, Fritsch F, Hayden H, Pawelka E, Krüger P, Rumpf B, Traugott MT, Glaser P, Firbas C, Schörghofer C, Seitz T, Karolyi M, Pabinger I, Brostjan C, Starlinger P, Weiss G, Bellmann-Weiler R, Salzer HJF, Jilma B, Zoufaly A, Assinger A. Low-molecular-weight heparin use in coronavirus disease 2019 is associated with curtailed viral persistence: a retrospective multicentre observational study. *Cardiovasc Res* 2021;**117**:2807–2820.